## 1 Exploration of 6-Methyl-7-(Hetero)Aryl-7-Deazapurine Ribonucleosides as Antileishmanial Agents

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## 10 Graphic

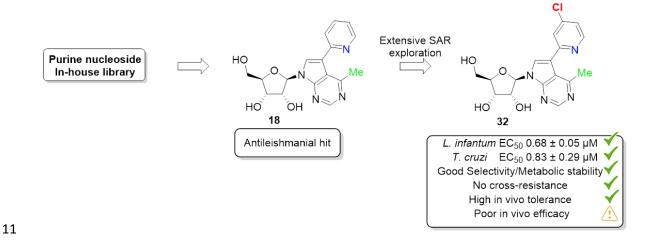
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#### Abstract

Leishmaniasis causes high mortality and morbidity in tropical and subtropical regions of Africa, Asia, the Americas and southern Europe, and is characterized by diverse clinical manifestations. As a neglected tropical disease, limited resources are allocated for antileishmanial drug discovery. The *Leishmania* parasite is deficient in *de novo* purine synthesis, and therefore acquires purines from the host and processes these using a purine salvage pathway. By making use of purine transport systems and interfering with this salvage pathway, purine (nucleoside) analogues might exert a selective detrimental impact on its growth and survival. *In vitro* screening of an in-house purine nucleoside library and analogue synthesis afforded the 6-methyl-7-(2-pyridyl)-7-deazapurine ribonucleoside analogue 18 as a promising hit. Optimization of the 7-substituent afforded 31 and 32 which displayed potent activity against wild-type and resistant *L. infantum*, intracellular amastigote and extracellular promastigote forms, and favorable selectivity versus primary mouse macrophages (Mφ) and MRC-5 cells. Encouraged by the favorable *in vitro* metabolic stability of 32, an *in vivo* study was performed using an early curative *L. infantum* hamster model. When orally administrated at 50 mg/kg once daily (s.i.d) for 10 days, 32 was devoid of side effects, however, it only poorly reduced amastigote burdens in the major target organs.

#### 1. Introduction

Leishmaniasis is a poverty-related and neglected tropic disease (NTD) endemic in 98 countries in Asia, Africa, southern Europe, and South and Central America.[1] According to the World Health Organization, leishmaniasis is classified as a Category I (emerging or uncontrolled) disease responsible for 25,000 deaths and 700,000 to one million new cases each year.[2] *Leishmania* is an obligate intracellular protozoan of macrophages that is transmitted during a blood meal of infected female sand flies.[3, 4] The parasite has a complex life cycle, involving macrophage invasion as a promastigote and multiplication within host

41 phagolysosomes in an amastigote form. Intra-macrophage *Leishmania* may evade the host immune defenses 42 by remodeling the phagosomal compartments and disturbing signaling pathways that generate lysosomal 43 enzymes and toxic metabolites.[5] 44 Leishmaniasis has three main clinical manifestations: cutaneous (CL), mucocutaneous (MCL) and visceral 45 (VL).[1] Unless treated, VL is fatal due to severe invasion of spleen, liver and bone marrow.[6] CL may 46 potentially cause lifelong scarring. In the absence of vaccines for human leishmaniasis, [7] antileishmanial chemotherapy remains the sole front-line strategy to combat the disease. Several treatments are available 47 48 such as the pentavalent antimonials, amphotericin B, miltefosine (MIL) and allopurinol (Figure 1A). MIL 49 affects the parasite membrane composition through inhibition of phospholipid synthesis by blocking the 50 transport of the choline precursor from the host, thereby impeding the synthesis of phosphatidylcholine and 51 phosphatidylethanolamine.[8]Allopurinol is metabolized into aminopurinol riboside triphosphate followed 52 by incorporation into RNA.[9] None of these drugs is ideal because of toxicity, cost, route of administration 53 or drug resistance. Hence, there is an urgent and continuous need to identify new drugs. 54 Since Leishmania species (spp.) cannot synthesize purine rings de novo, they evolved an extensive set of 55 transporters[10, 11] and salvage enzymes[12, 13] to scavenge external purines.[13] This salvage pathway enables Leishmania to dephosphorylate extracellular nucleotides into nucleosides via 3' and 5'-56 57 nucleotidases/nucleases[14] prior to uptake and intracellular conversion to the required nucleotides.[12, 15] 58 This absolute reliance on scavenging host purine nucleobases or nucleosides renders *Leishmania* vulnerable 59 to nucleoside analogues that interfere with active import processes or behave as subversive substrates as 60 mentioned above for allopurinol.[13] Our steadily growing in-house synthesized library of nucleoside 61 analogues earlier proved a valuable source of antitrypanosomal in vitro hits (Figure 1B). Notably, 62 modifications of tubercidin on C7[16] (e.g. 1), C-3'(e.g. 2[17] and 3[18]), C6 and C7 (e.g. 4[19] and 5[20]), 1.7-dideazapurines (pyrrolo[2,3-b]pyridine)[21] (e.g. 63 as well structurally related pyrazolopyrimidines[22] (e.g. 7) and C-nucleosides[23] (e.g. 8) displayed promising activity against 64 65 different kinetoplastids. Also Michal Hocek's team has recently identified 6-alkoxy-7-methyltubercidin

derivatives (e.g. 9, Figure 1A) with significant activity against Trypanosoma brucei (T. b.) brucei and T.

67 b. gambiense.[24] Remarkably, the hit rate against Leishmania is comparatively low.

7-Deazapurine nucleoside analogues that do show in vitro antileishmanial activity include the 7-(3,4-

dichlorophenyl) analogue 10,[16] and the 6-methyl-7-3,6-(dihydro-2*H*-pyranyl) analogue 11

(Figure 1C).[26] The promising activity of 10 (7-aryl analogue) and 11 (6-Me analogue) led us to evaluate

known[20] and new hybrid 6-methyl-7-aryl analogues against *L. infantum* in Mφ.

(A) Known antileishmanial agents

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(B) Previously reported representative antitrypanosomal nucleoside analogues by us and others

(C) Previously identified antileishmanial nucleoside analogues

CI ON NH2 HO 10 OH NNN HO 11 OH NNN HO 11 OH PMM 
$$CC_{50} = 8.00; 32.0 \ \mu\text{M}$$
 PMM  $CC_{50} > 64.0 \ \mu\text{M}$ 

Figure 1. Known antileishmanial agents (A), representative examples of previously identified nucleoside
analogues with antitrypanosomal activity (base and sugar modification highlighted in green and orange,

respectively) (B), and previously identified nucleoside antileishmanial nucleoside analogues (C). Throughout the text, purine numbering is used as depicted for compound 1, while in the experimental section, systematic numbering is used.

# Table 1. In vitro activity of 6-methyl-7-aryl derivatives against Leishmania intracellular amastigotes<sup>a</sup>

		L. infantum	Мф	
Cpd.b	Ar	$EC_{50}$ ( $\mu$ M)/5 days	$CC_{50}$ ( $\mu M$ ) )/5 days	SI
12	- <del> </del> -CI	$16.1 \pm 5.4$	> 64	> 3.5
13	CF <sub>3</sub>	$11.3 \pm 9.3$	> 64	> 5.6
14	OMe 	$8.76 \pm 0.76$	> 64	> 7.3
15	F CI	$7.9 \pm 0.70$	> 64	> 8.0
16	SO <sub>2</sub> Me	$6.7 \pm 3.9$	> 64	> 9.5
17	NH <sub>2</sub>	$6.7 \pm 1.3$	> 64	> 9.5
18	- -\(\sigma\)	$2.3 \pm 1.1$	> 64	> 27
19	- -S	12.7	32	2.5
MIL		$8.3 \pm 1.6$		

<sup>a</sup>Evaluation of drug sensitivity against *L. infantum* (MHOM/MA (BE)/67) of known and new 6-methyl-7-aryl analogues (above and below dotted line respectively). Cytotoxicity was assayed on Mφ. Values represent mean  $\pm$  SEM, which originate from 2 to 3 independent experiments and are expressed in μM. SI, *in vitro* selectivity index is the ratio of the EC<sub>50</sub> for the host cell (Mφ) and the EC<sub>50</sub> of the parasite. MIL was included as a reference (EC<sub>50</sub> = 8.30  $\pm$  1.64 μM). Values represent *in italics* indicate the data of a single experiment due to too low selectivity or activity. <sup>b</sup>Compounds **12**, **13**, **14** and **15** were reported in our previous study,[20] while **16**, **17**, **18** and **19** were synthesized for this study.

Compounds **12-15** showed reasonable *in vitro* activity and were devoid of cytotoxicity but failed to approximate **10** or **11** (**Table 1**). This led us to explore alternative aryl substituents and to introduce a heteroatom into the aromatic moiety. The 6-methyl-7-(2-pyridyl)-7-deazapurine nucleoside analogue **18** showed improved antileishmanial activity with a promising selectivity. To further expand the SAR different alternatives for the pyridin-2-yl ring were explored (**Figure 2**).

Figure 2. Overview of heteroaromatic optimization in seven devised structural modules

#### 2. Results and discussion

# **2.1. Chemistry**

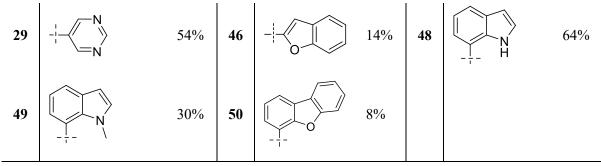
The target 6-methyl-7-(hetero)aryl nucleoside analogues were synthesized via transition-metal catalyzed coupling reactions from either benzoyl-protected or unprotected 6-methyl-7-deaza-7-iodopurine ribonucleoside precursors.[20, 27] Suzuki condition[28] employing Pd(OAc)<sub>2</sub> and TPPTS allowed to couple aryl boronic acids or their pinacol ester (**Scheme 1**). Alternative conditions were required to couple pyridin-4-ylboronic acid and Pd<sub>2</sub>(dba)<sub>3</sub>, P(c-Hex)<sub>3</sub> ligand and K<sub>3</sub>PO<sub>4</sub> in a mixture of water/1,4-dioxane (1:2) at 100 °C gave satisfactory yields.[29] The 2-pyrrol-2-yl analogues **38** and **46** were obtained after removal of the Boc-protecting group under basic conditions. A Stille reaction proved favorable to produce the 2-pyridyl (**18**) and 2-thienyl (**19**) analogues, while the (*E*)-2-pyridin-2-ylethenyl group was introduced via a Heck reaction and a thiophenyl group under Ullmann conditions.[30, 31]

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#### Scheme 1

a: arylboronic acid/arylboronic pinacol ester, Pd(OAc)<sub>2</sub>, TPPTS, Na<sub>2</sub>CO<sub>3</sub>, MeCN / H<sub>2</sub>O (1/2 ratio), 100 °C

16	-i-SO <sub>2</sub> Me	50%	22	F	65%	23	CI	31%
24				S				69%
37	CI	26%	40	N-NH	39%	41	N	71%
42	S	32%	43	N-NH 	53%	20	- -\(\bigc\)	36%



b: pyridin-4-ylboronic acid, K<sub>3</sub>PO<sub>4</sub>, Pd<sub>2</sub>(dba)<sub>3</sub>, P(c-Hex)<sub>3</sub>, H<sub>2</sub>O/dioxane (1/2 ratio), 100 °C

21	- N	45%		

c: (i) arylboronic acid, Pd(OAc)<sub>2</sub>, TPPTS, Na<sub>2</sub>CO<sub>3</sub>, MeCN / H<sub>2</sub>O (1/2 ratio), 100 °C; (ii) 0.5 M NaOMe in MeOH

d: Ar-Sn(nBu)<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>, CuI, DMF, 100 °C

e: 2-vinylpyridine, Pd(OAc)<sub>2</sub>, TPPTS, TEA, DMF, 100 °C



f: thiophenol, CuI,  $K_2CO_3$ , ethyleneglyol, iPrOH, 130 °C

54	- -s-	92%		

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Next, the obtained nitro (26 and 56[20]) and (2-pyridyl)vinyl analogues (52) were further hydrogenated into the corresponding amino (17 and 27) and (2-pyridyl)ethyl congeners (53) under Pd(OH)<sub>2</sub>/C system (Scheme 2), respectively.

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Scheme 2

	Ar	yield	Ar'	
56	-i-NO <sub>2</sub>	92%	-i-NH <sub>2</sub>	17
26	O <sub>2</sub> N	69%	H <sub>2</sub> N	27
52	† N	64%		53

Substituted 2-pyridyl, diazinyl and quinolinyl analogues were prepared by Negishi coupling (Scheme

with heteroaromatic bromides. Due to the instability of 2-pyridyl organometallics,[32] the overall yields

were low to moderate. The 2-thiazolyl derivative was obtained via Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub>-catalyzed Stille cross-

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3).[16] Magnesium-iodine exchange with iPrMgCl·LiCl and subsequent transmetallation with ZnCl<sub>2</sub> solution of perbenzoylated 6-Me-7-iodopurine ribonucleoside **57**[28] afforded the zinc partner to couple

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## Scheme 3

coupling followed by deprotection in 7 N NH<sub>3</sub>/MeOH.

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a: (i) iPrMgCl·LiCl, ZnCl<sub>2</sub>, THF, -65 °C to r.t.; (ii) Ar-Br, Pd<sub>2</sub>(dba)<sub>3</sub>, RuPhos, THF, 60 °C, overnight; (iii) 7 N NH<sub>3</sub> in MeOH

28	- -\(\sum_{N}^{N}-\)	63%	29	-I-N-N	37%	31	CI 	33%
32	CI 	29%	33		23%	34		22%
35	- -\(\sum_{N}\)	18%	36	- -\(\)N=\(\)F	5%	44	N	30%
45	N	19%	48	N N	46%			

b: (i) 2-(tripropylstannyl)thiazole, Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub>, DMF, 100 °C; (ii) 7 N NH<sub>3</sub> in MeOH

39 -	N 56%	ó			
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#### 2.2. Biological evaluation

#### 2.2.1. *In vitro* activity profile

All prepared compounds were evaluated for *in vitro* activity against intracellular amastigotes of *L. infantum* (MHOM/MA(BE)/67 strain), using MIL and allopurinol as the reference drug (**Table 2**). In parallel, cytotoxicity was checked against the M $\phi$  and MRC-5 fibroblasts.

In contrast to the inactive phenyl analogue,[26] the pyridin-2-yl analogue showed relevant activity. Shuffling of the nitrogen atom has a negative impact on the activity. *Ortho*-modification of the 7-phenyl substituent with a fluoride (22) or chloride (23) or other hetero-containing substituents (24, 25, 26 and 27) failed to improve the activity, underscoring the importance of the nitrogen atom in 18. Introduction of an

additional nitrogen atom in *ortho* or *para* position gave a 4- to 5-fold drop in activity (28, 29). Interestingly, the 3,5-diazinyl derivative 30 (without ortho N atom) displayed similar micromolar activity as the above diazinyl analogues. Further derivatization focused on increasing the electron deficiency of the pyridine moiety. Incorporating a chloro in *ortho* position (31) slightly increased activity while remaining non-toxic for the host cells. Moving the chloro atom to the *meta* position afforded 32 with submicromolar activity and a favorable SI (> 94.1). Introduction of an electron-donating methoxy group in this position resulted in a complete loss of activity. Moving the chloro atom to the para position (34) further increased the activity and was 20-fold better than MIL. The compound with the highest antileishmanial in vitro activity (34) showed moderate cytotoxicity on MRC-5 cells while 31 and 32 were devoid of notable cytotoxicity. Further rotation of the chloro-atom (35) had a negative impact. Replacement in 35 of the chloro by a fluoro atom (36) and shifting the nitrogen of 34 to the *meta* position (37) proved detrimental for activity. Changing the pyridin-2-yl moiety for a 2-pyrrol-2-yl (38) boosted the activity (EC<sub>50</sub> =  $0.35 \mu M$ ), however, at the expense of selectivity. Three azolo analogues (39, 40 and 41) showed similar potency as the diazinyl analogues. Since the 2-thienyl 19 was only moderately active and the 2-furanyl derivative proved cytotoxic, [27] 3thioenyl (42) and 3-furanyl (43) derivatives were investigated but failed to show activity. Fusion of the pyridine substituent with a phenyl to afford the 2- and 3-quinolinyl analogues 44 and 45 failed as well. Interestingly, the indolyl and benzofuran-2-yl analogues 46 and 47, resulting from the fusion of a phenyl ring to the pyrrol-2-yl and furan-2-yl substituents, conferred lower cytotoxicity and the former showed low micromolar activity. Surprisingly, the 8-quinolinyl analogue 48 showed a similar potency as the 2-pyridyl analogue. The 7-indolyl analogue 49 was equipotent to 48. The N-methyl-substituted analogue of 49 (50) was inactive. Analogues in which the heterocyclic moiety was attached to the purine via a short spacer (52, 53 and 54) also proved inactive. The SAR trends of 7-heteroaryl modified 6-methyl nucleoside analogues are summarized in **Figure 3**.

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Table 2. In vitro activity of 7-heteroaromatic derivatives against Leishmania intracellular amastigotes<sup>a</sup>

Cpd.	L. infantum EC <sub>50</sub> (μM)/5 days	Mφ CC <sub>50</sub> (μM) )/5 days	MRC-5 CC <sub>50</sub> (μM) )/3 days	SI vs. Mφ
18	$2.3 \pm 1.1$	> 64	> 64	> 27
3- and 4-pyridine	L			
20	22.6	> 64	> 64	> 2.8
21	> 64	> 64	> 64	ND
Ortho-substituted	phenyl			
22	> 64	> 64	> 64	ND
23	$20.4 \pm 9.0$	> 64	> 64	> 3.1
24	21.1	> 64	> 64	> 3.0
25	> 64	> 64	> 64	ND
26	> 64	> 64	> 64	ND
27	> 64	> 64	> 64	ND
Diazine				
28	$8.2 \pm 3.1$	> 64	> 64	> 7. 8
29	10.1	> 64	> 64	> 6.3
30	$6.5 \pm 0.5$	> 64	> 64	> 9.9
Substituted pyriding	ne			
31	$1.2 \pm 0.4$	> 64	> 64	> 50
32	$0.68 \pm 0.05$	> 64	> 64	> 94
33	> 64	> 64	> 64	ND
34	$0.40\pm0.05$	> 64	$26.3 \pm 20.7$	> 160
35	$14.2 \pm 7.0$	> 64	> 64	> 4.51
36	> 64	> 64	> 64	ND
37	> 64	> 64	> 64	ND

38	0.35	32	12.9	91.4				
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39	2.07	32	6.06	15.5				
40	8.0	> 64	27.9	> 8				
41	$8.0 \pm 1.0$	> 64	> 64	> 8				
42	> 64	> 64	> 64; 22.6	ND				
43	> 64	> 64	> 64	ND				
Multiple-cycle with one heteroatom								
44	> 64	> 64	> 64	ND				
45	> 64	> 64	> 64	ND				
46	$1.7\pm0.0$	> 64	23.6	> 36				
47	20.3	32	45.8	1.58				
48	$3.7 \pm 1.7$	> 64	> 64	> 17				
49	$2.1\pm0.3$	> 64	> 64	> 30				
50	> 64	> 64	> 64	ND				
51	50.8	> 64	12.7	> 1.2				
(Hetero)aryl space	r							
52	19.0	> 64	> 64	> 3.37				
53	> 64	> 64	> 64	ND				
54	> 64	> 64	> 64	ND				
MIL	$8.3 \pm 1.6$	ND	ND	ND				
allopurinol*	$3.5 \pm 1.8$	> 64	> 64	> 18				

<sup>a</sup>Evaluation of drug sensitivity against *L. infantum* (MHOM/MA(BE)/67/ITMAP263). Cytotoxicity was assayed on M $\phi$  and MRC-5 fibroblasts. Values represent mean  $\pm$  SEM which originate from 2 to 3 independent experiments and are expressed in  $\mu$ M. *In vitro* selectivity index (SI) is the ratio of the CC<sub>50</sub> for the host cell (M $\phi$ ) and the EC<sub>50</sub> of the parasite. Values *in italics* indicate the data of a single experiment due to too low selectivity or activity. MIL and allopurinol were included as references. \*The EC<sub>50</sub> of allopurinol corresponds with that reported in our previous study.[22]

**Figure 3**. Summary of SAR trends for *L. infantum* 

### 2.2.2. *In vitro* profiling against drug-resistant strains

The promising analogues **31** and **32** were evaluated against three *L. infantum* strains that are resistant to potassium antimonyl [Sb(III)], sodium stibogluconate [Sb(V)], MIL and paromomycin (PMM) after 96 h incubation (**Table 3**). **31** retained activity against all resistant strains. **32** retained activity against LEM3323-Cl4 MIL5Cl3 and LEM3323-Cl4 PMM strains but exerted slightly lower activity against the parent Sbresistant LEM3323-Cl4 clinical isolate. It can be concluded that both compounds show no obvious cross-resistance to the established drugs and retained potent activity against extracellular promastigotes.

Table 3. In vitro evaluation of cross-persistence of selected nucleoside analogues<sup>a</sup>

Cpd.	LEM3323-Cl4 EC <sub>50</sub> (μM)	LEM3323-Cl4 MIL5Cl3 EC <sub>50</sub> (μM)	LEM3323-Cl4 PMM EC <sub>50</sub> (μM)	L. infantum EC <sub>50</sub> (μM)	Μφ CC <sub>50</sub> (μM)
31	0.67	0.73	1.46	$1.2 \pm 0.4$	> 64
32	2.21	0.73	0.79	$0.68 \pm 0.05$	> 64

<sup>a</sup> Susceptibility of drug-resistant *L. infantum* promastigotes for analogues **31** and **32**. Strain MHOM/FR/96/LEM3323-Cl4 is resistant to sodium stibogluconate [Sb(V)] and potassium antimonyl tartrate [Sb(III)]; LEM3323-Cl4 MIL5Cl3 strain is resistant to Sb(V), Sb(III) and MIL; LEM3323-Cl4 PMM is resistant to Sb(V), Sb(III) and PMM.[33]

## 2.2.3. *In vitro* evaluation against *Trypanosoma cruzi* (*T. cruzi*)

As some 6-methyl-7-aromatic-7-deazapurine nucleoside analogues possessed favorable activity against *T. cruzi*[20], *e.g.* **12**, analogues with the most promising *in vitro* activity against *Leishmania* were also evaluated for anti-*T. cruzi* activity (**Table 4**). The 2-pyridyl (**18**), 8-quinolinyl (**48**) and 7-indolyl analogue (**49**) displayed micromolar activity, while the chloropyridyl derivatives (**31** and **32**) exhibited submicromolar potency without apparent cytotoxicity on MRC-5 cells.

Table 4. In vitro activity of selected nucleoside analogues against T. cruzi intracellular amastigotes<sup>a</sup>

	T. cruzi	MRC-5	
Cpd.	$EC_{50} (\mu M)/7 days$	$CC_{50}$ ( $\mu M$ )/3 days	SI
18	$1.6 \pm 0.6$	> 64	> 39
28	$5.9 \pm 3.9$	> 64	> 10
30	$7.3 \pm 1.5$	> 64	> 8.8
31	$0.70 \pm 0.07$	> 64	> 91
32	$0.83 \pm 0.29$	> 64	> 77
34	$0.49 \pm 0.01$	$26.3\pm20.7$	53
38	0.68	12.9	19
39	0.66	6.06	9.2
46	$0.62 \pm 0.08$	$14.80 \pm 8.77$	24
48	$9.6 \pm 0.5$	> 64	> 6.7

49	$3.3 \pm 0.4$	> 64	> 19
Benznidazole	$2.28 \pm 0.05$	ND	ND

<sup>a</sup>Evaluation of drug sensitivity against *T. cruzi* (Tulahuen strain expressing β-galactosidase). Cytotoxicity was assayed on MRC-5 fibroblasts. Values represent mean  $\pm$  SEM which originate from 2 to 3 independent experiments and are expressed in μM. *In vitro* selectivity index (SI) is the ratio of the CC<sub>50</sub> of MRC-5 and the EC<sub>50</sub> of the parasite. Benznidazole was included as a reference (EC<sub>50</sub> = 2.28  $\pm$  0.05 μM). Values *in italics* indicate the data of a single experiment due to too low selectivity or activity.

#### 2.2.4. *In vitro* metabolic stability

Compounds **31** and **32** were exposed to liver microsomal fractions originating from mouse, hamster and human to assess their metabolic stability (**Table 5**). The percentage of remaining parent compound was determined at three time points. Both were hardly metabolized under incubation with either NADPH fractions (Phase-I) or UGT fractions (Phase-II) regardless the originating species. The more potent **32** was selected for *in vivo* follow-up in the *L. infantum* hamster model.

Table 5. *In vitro* metabolic stability of selected nucleoside analogues in mouse, hamster and pooled human S9 microsomal fractions<sup>a</sup>

			Amount of compounds remaining (%)				
Microsomes	Phase I / II	Time		31		32	
			Mean	STDEV	Mean	STDEV	
Mouse		0	100	-	100	-	
	CYP – NADPH	15 98	98	5.7	109	9.2	
	CII -NADIII	30	83	11.9	86	14.3	
		60	74	16.8	94	5.9	

		0	100	-	100	-
	UGT	15	92	0.6	103	2.2
		30	91	8.5	102	4.9
		60	98	3.7	100	8.9
		0	100	-	100	-
	CYP – NADPH	15	86	4.7	96	13.7
	CIP-NADPH	30	98	1.6	95	16.4
Hamster		60	90	1.1	87	9.0
Humster	UGT	0	100	-	100	-
		15	95	0.8	98	9.5
		30	96	5.2	92	7.2
		60	96	5.5	89	12.0
		0	100	-	100	-
	CYP – NADPH	15	107	4.0	92	3.6
	CIF - NADFH	30	100	5.4	86	4.5
Human		60	99	7.4	75	0.8
Human		0	100	-	100	-
	UGT	15	93	7.5	93	4.4
	001	30	95	9.2	96	1.3
		60	101	11.7	98	5.3

<sup>a</sup>Values (mean and STDEV) represent the remaining percentage of parent compound after 0–15–30–60 min of incubation, based on two independent assays. CYP – NADPH refers to Phase-I metabolism, and UGT refers to Phase-II metabolism. Diclofenac, susceptible to Phase-I and Phase-II metabolism, was used as reference (*data not shown*). Mouse and hamster microsomal fractions were selected since these laboratory rodent species are respectively used for *T. cruzi* and *L. infantum* infection.

# 2.2.5.1. Efficacy in the target organs liver, spleen, and bone-marrow in the early curative hamster model of *L. infantum*

Inspired by the encouraging *in vitro* activity against intracellular amastigotes, 32 was selected for *in vivo* evaluation in the early curative hamster model of *L. infantum*. Treatment was initiated at 21 days post-infection (dpi). 32 was orally administrated at 50 mg/kg s.i.d for 10 consecutive days and the efficacy was determined by the percentage reduction of amastigote burdens in the major target organs, namely liver, spleen and bone-marrow (**Table 6**). Percentage reduction compared to the burdens in the vehicle-treated infected control animals (VIC) was used as a measure for activity. MIL was included as reference (40 mg/kg, PO, s.i.d, 5 consecutive days). Unfortunately, 32 showed poor efficacy with organ burden reductions in liver, spleen and bone-marrow of 52.7%, -1.8% and 30.8%, respectively. (Supplementary figure in supporting information) A 'viable' residual burdens assay was performed for the individual organ samples using the promastigote back-transformation assay (**Table 7**). As could be expected, all samples turned positive, confirming that no sterile cure could be obtained.

Table 6. Percentage reduction in amastigote burdens in target organs with 32 in the early curative L. infantum infected hamster model<sup>a</sup>

	% reduction of amastigote burdens					
Experimental Group	in target organs					
	Liver	Spleen	Bone-marrow			
<b>G1</b> : VIC: 10% (v/v) PEG 400 and 1% Tween 80 (v/v) in water	-	-	-			
G2: MIL 40 mg/kg PO SID for 5 days	94.0	99.2	93.1			
<b>G3</b> : <b>32</b> : 50 mg/kg PO SID for 10 days	52.7	-1.8	30.8			

<sup>a</sup>G1: Vehicle-treated control (VIC), G2: MIL-treated group, G3: 32-treated group. Each experimental group was allocated randomly with five infected hamsters; Dosing started at 21 dpi; Amastigote burdens in the

different target organs (liver, spleen, bone-marrow) were determined 9 days after the last treatment (*i.e.* day 39 of the experiment).

## Table 7. Promastigote back-transformation assay for the individual organ samples<sup>a</sup>

		Response of promastigotes in target organs				
<b>Experimental Group</b>	n	Liver	Spleen	Bone-marrow		
	1	+++	+++	+++		
<b>G1</b> : VIC: 10% (v/v) PEG	2	+++	+++	+++		
400 and 1% Tween 80 (v/v)	3	+++	++	+++		
in water	4	+++	+++	+++		
	5	+++	+++	++		
	1	++	+	++		
	2	++	++	+		
G2: MIL 40 mg/kg PO SID for 5 days	3	++	++	++		
•	4	++	++	++		
	5	+	+	+		
	1	+++	+++	+++		
	2	+++	++	+++		
<b>G3</b> : <b>32</b> : 50 mg/kg PO SID for 10 days	3	+++	+++	++		
<del></del>	4	+++	+++	+++		
	5	+++	+++	+++		

<sup>&</sup>lt;sup>a</sup>Incubation of small tissue samples in promastigote medium at room temperature with qualitative assessment of the presence of promastigotes after two weeks of incubation.

# 2.2.5.2. Pharmacokinetic (PK) profile

The absorption and elimination kinetics of 32 were studied by LC-MS/MS on dried blood spots (DBS) collected after the first dose on day 1 and after the morning dose of day 9. After dosing at 50 mg/kg, the maximum blood concentrations in infected hamsters were reached at 0.5 hours ( $T_{max}$ ) with maximum blood levels ( $C_{max}$ ) exceeding the *in vitro* IC<sub>50</sub> value by about 4-fold, inferring that the concentration of 32 should in principle be sufficient to inhibit the growth of *L. infantum* (**Table 8**). With respect to elimination, Cl, AUC and  $T_{1/2}$  indicated that 32 was not quickly cleared. Compared to single-dose treatment, the AUC after repeated-dose treatment increased, while the Cl decreased, indicating that a greater amount of 32 became systemically available. However, a decrease in volume of distribution (Vd) and  $T_{1/2}$  suggests that the higher systemic exposure of 32 is unexpectedly accompanied by an inferior tissue distribution. The low Vd in particular in the Day 9 (Vd < 20 L/kg) indicates that 32 is predominately localized to plasma or extracellular fluid or is highly bound rather than extensively distributed throughout tissues, which probably accounts for the poor efficacy in the target organs. Besides, the nucleoside transporters in rodents are known to be unevenly distributed over different tissues,[34] which may explain the inconsistent reduction percentages in the different organs, despite the favorable *in vitro* activity and metabolic stability. A liposomal formulation[35] could be considered to improve the distribution and macrophage targeting of 32.

Table 8. Non-compartmental pharmacokinetic parameters of 32 in blood after a single dose (<u>Day 1</u>: 50 mg/kg) and after repeated-dose administration (<u>Day 9</u>: 50 mg/kg s.i.d. for 10 days) in infected hamsters<sup>a</sup>

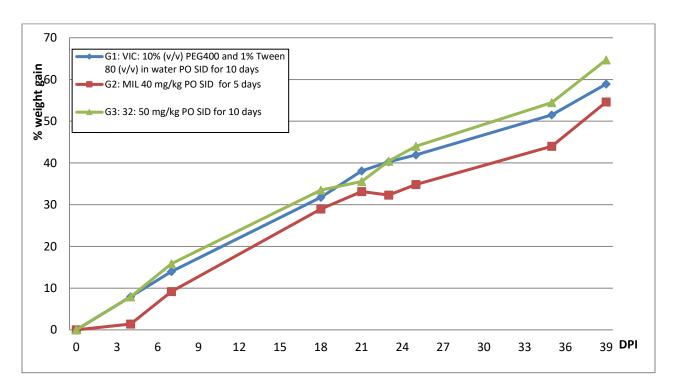
Experimental	T <sub>max</sub>	$\mathbf{C}_{max}$	$C_{max}$	T <sub>1/2</sub>	AUC <sub>0→8</sub>	AUC <sub>0→24</sub>	Cl	Vd
Group	h	ng/mL	μΜ	h	ng.h/mL	ng.h/mL	mL/min/kg	L/kg
G4 Day 1-50 mg/kg	0.5	1096	2.9	2.7	1664	2080	56.6	13.0
G4 Day 9-50 mg/kg	0.5	1213	3.2	1.7	2844	3380	34.7	5.14

<sup>a</sup>Blood samples taken until 8 hours after morning dose on day 1. All samples were collected as DBS for analytical detection of parent compound. The calculated PK parameters are based upon the mean values

per time point using non-compartmental analysis. The areas under the curve ( $AUC_{0-8h}$  and  $AUC_{0-24h}$ ) were calculated using the linear trapezoidal rule. The standard PK parameters were determined using appropriate software (TopFit):  $C_{max}$ ,  $T_{max}$ , AUC, Cl and Vd.

## **2.2.5.3.** *In vivo* toxicity

After oral dosing of 50 mg/kg for 10 days, no clinical adverse effects, gross pathological changes nor severe weight losses (**Figure 4**) were noted in all **32**-treated hamsters.



**Figure 4**. Evolution of body weight gain for 39 dpi. The percentage gain was compared to the start of the experiment, based on the mean from the five hamsters.

#### 3. Conclusion

Phenotypic drug discovery is widely considered as an important strategy for the discovery of new agents for NTD.[36-38] In the present work, screening of a nucleoside library and further heteroaromaticity led to the identification of 6-methyl-7-(2-pyridyl)-7-deazapurine ribonucleoside as an antileishmanial hit. Heteroaromatic optimization afforded the 4-chloropyridyl analogue 32, which combined submicromolar activity and high selectivity towards host cells. In view of its *in vitro* metabolic stability, 32 was evaluated in an early curative hamster model of *L. infantum*. Although 32 was well tolerated, it showed poor efficacy in eliminating the parasite from the target organs.

#### 4. Materials and methods

#### 4.1. General chemistry

All chemical reagents and solvents were supplied by standard commercial sources and had analytical grade. Compounds 12, 13, 14 and 15 were obtained from the previous stock.[20] Machery–Nagel precoated F254 aluminum plates were used for analytical thin-layer chromatography (TLC), which was visualized under UV light at 254 nm. Column chromatography was performed for the purification, using Machery-Nagel 60 M silica gel (40-63 µm) or on a Reveleris X2 (Grace) automated Flash unit. NMR spectra were recorded with a Bruker Avance Neo 400 MHz or a Varian Mercury 300 MHz spectrometer and referenced to the residual solvent peak signal. High-resolution mass spectrometry (HRMS) was performed with a Waters LCT Premier XE time-to-flight (TOF) mass spectrometer equipped with a standard electrospray ionization (ESI) and modular LockSpray interface. Purities of the final compounds were assessed via analytical LC-MS on a Waters AutoPurification system (equipped with ACQUITY QDa (mass; 100–1000 amu)) and 2998 Photodiode Array (220–400 nm) equipped with a Waters Cortecs C18 column (2.7 µm, 100 × 4.6 mm) and a gradient system of formic acid in H<sub>2</sub>O (0.2% v/v)/MeCN at a flow rate of 1.44 mL/min and a gradient of 95:5 to 0:100 in 6.5 min. All the purities of final compounds for biological evaluation were > 95%. Purifications of final compounds using preparative High-performance liquid chromatography (HPLC) was

in the same system equipped with a Phenomenex Luna Omega Polar column (5  $\mu$ m, 250 mm  $\times$  21 mm) with a gradient system of formic acid in H<sub>2</sub>O (0.2% v/v)/MeCN at a flow rate of 20 mL/min.

#### 4.2 The synthesis of target compounds

#### 4.2.1. General procedure A of Suzuki coupling reaction (adapted from Ref.[27])

**Iodo-nucleoside** (1 eq.), boronic acid reagent (1.5 eq.), Na<sub>2</sub>CO<sub>3</sub> (3 eq.), Pd(OAc)<sub>2</sub> (0.1 eq.) and TPPTS (0.2 eq.) were added to a 10 mL round-bottom flask, flushed with argon. Next, a mixed solution of MeCN/water (1/2 ratio, 6 mL/mmol SM) was added via syringe. The reaction mixture was stirred at ambient temperature for 5 min, and then stirred at 100 °C. When completion of the reaction was observed via LC-MS analysis (~0.5 to 3 h), the reaction mixture was cooled to room temperature, neutralized with 0.5 M aq. HCl, and evaporated. The residue was resuspended in MeOH and evaporated *in vacuo*, which was repeated three times. The residue was adsorbed by Celite® and washed through a short silica pad using 20% MeOH/DCM. The resulting solution was evaporated *in vacuo* and purified by column chromatography using MeOH/DCM gradient.

#### 4.2.2. General procedure B of Negishi coupling reaction[16]

**Iodo-nucleoside** (1 eq.) was added in a 25 mL oven-dried round-bottom flask and co-evaporated three times with anhydrous toluene (10 mL). The resulting solid was dissolved in anhydrous THF (8.5 mL/mmol SM) under argon and cooled to -65 °C. iPrMgCl·LiCl solution (1.3 M in THF, 1.1 eq.) was added in one portion with a syringe. The reaction mixture was stirred at -65 °C for 30 min. Consumption of iodo-nucleoside was monitored via TLC analysis after quenching a small sample with sat. NH<sub>4</sub>Cl solution. Then, ZnCl<sub>2</sub> solution (0.5 M in THF, 1.2 eq.) was added. The reaction mixture was stirred at -65 °C for 5 min and for 1 h at ambient temperature. Pd<sub>2</sub>(dba)<sub>3</sub> (0.02 eq.), RuPhos (0.08 eq.) and the appropriate bromo-pyridine or bromo-quinoline (1.4 eq.) were added in a flame-dried Schlenk-tube (5 mL) purged with argon and dissolved in anhydrous THF (3 mL/mmol SM). The mixture was stirred for 5 min and then transferred via a syringe to the flask containing the nucleoside-zinc reagent. The mixture was stirred at 60 °C overnight,

328 cooled, quenched by adding water (~ 5 mL) and transferred to a separatory funnel. EA (10 mL) and aq. 1M

EDTA (pH = 8) solution (5 mL) were added. The layers were separated and the water layer was extracted

with EA two more times. The organic layers were combined, dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated and purified

by column chromatography unless indicated. The collected fractions were evaporated and the residue

dissolved in 7 N methanolic NH<sub>3</sub> solution. The mixture was stirred at ambient temperature overnight after

which it was evaporated until dryness and purified by column chromatography ( $0 \rightarrow 10\%$  MeOH/DCM).

- 4.2.3. 4-Methyl-5-(4-(methylsulfonyl)phenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (16).
- 335 16 was prepared according to General procedure A. 55 (100 mg, 0.26 mmol) and (4-
- (methylsulfonyl)phenyl)boronic acid (62 mg, 0.31 mmol) gave rise to **16** (55 mg, 0.13 mmol) as a white
- solid in 50% yield.  ${}^{1}$ H NMR (300 MHz, DMSO- $d_6$ )  $\delta$ : 2.65 (s, 3 H, CH<sub>3</sub>), 3.28 (s, 3 H, CH<sub>3</sub>), 3.50 3.60
- 338 (m, 1 H, H-5"), 3.60 3.73 (m, 1 H, H-5"), 3.94 (q, J = 3.7 Hz, 1 H, H-4"), 4.07 4.21 (m, 1 H, H-3"), 4.48
- 339 (t, J = 5.4 Hz, 1 H, H-2'), 5.05 (br. s., 1 H, OH-5'), 5.18 (br. s., 1 H, OH-3'), 5.40 (br. s., 1 H, OH-2'), 6.29
- 340 (d, J = 5.9 Hz, 1 H, H-1'), 7.80 (d, J = 8.5 Hz, 2 H, H-Ph), 7.93 8.09 (m, 3 H, H-Ph and H-6), 8.75 (s, 1
- 341 H, H-2). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 23.3 (CH<sub>3</sub>), 43.6 (CH<sub>3</sub>), 61.4 (C-5'), 70.5 (C-3'), 74.1 (C-2'),
- 342 85.3 (C-4'), 86.7 (C-1'), 115.5 (C-4a), 115.5 (C-5), 126.0 (C-6), 127.0 (2 C, C-Ph), 130.4 (2 C, C-Ph),
- 343 139.2 (C-Ph), 139.6 (C-Ph), 150.9 (C-2), 151.0 (C-7a), 159.3 (C-4). HRMS (ESI): calculated for
- 344  $C_{19}H_{22}N_3O_6S$  ([M+H]<sup>+</sup>): 420.1224, found: 420.1218.

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- 345 4.2.4. 4-Methyl-5-(4-aminophenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (17). The p-
- nitrophenyl compound **56** (70 mg, 0.18 mmol) was dissolved in MeOH (5 mL) and H<sub>2</sub>O (1 mL) and purged
- with  $N_2$  atmosphere. A catalytic amount of Pd(OH)<sub>2</sub>/C was added and the  $N_2$  was exchanged with  $H_2$  gas
- (balloon; bubbling). The reaction mixture was stirred at ambient temperature until LC-MS analysis showed
- full conversion of **56** (5 h). The reaction mixture was filtered over Celite® and rinsed with MeOH. The
- filtrate was evaporated and the residue was purified by column chromatography ( $5 \rightarrow 20\%$  MeOH/DCM)
- 351 to give 17 (60 mg, 0.17 mmol) as a white solid in 92% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.45 (s, 3)
- 352 H, CH<sub>3</sub>), 3.51 3.56 (m, 1 H, H-5''), 3.60 3.65 (m, 1 H, H-5'), 3.91 (q, J = 3.6 Hz, 1 H, H-4'), 4.09 4.13

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353 (m, 1 H, H-3'), 4.45 (q, J = 6.2 Hz, 1 H, H-2'), 5.04 (t, J = 5.4 Hz, 1 H, OH-5'), 5.14 (d, J = 4.5 Hz, 1 H,
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- 354 OH-3'), 5.17 (s, 2 H, NH<sub>2</sub>), 5.33 (d, J = 6.1 Hz, 1 H, OH-2'), 6.24 (d, J = 6.3 Hz, 1 H, H-1'), 6.64 (d, J = 6.1 Hz, 1 H, OH-2'), 6.24 (d, J = 6.3 Hz, 1 H, H-1'), 6.64 (d, J = 6.1 Hz, 1 H, OH-2'), 6.25 (d, J = 6.3 Hz, 1 H, H-1'), 6.64 (d, J = 6.1 Hz, 1 H, OH-2'), 6.25 (d, J = 6.3 Hz, 1 H, H-1'), 6.64 (d, J = 6.1 Hz, 1 H, OH-2'), 6.25 (d, J = 6.3 Hz, 1 H, H-1'), 6.64 (d, J = 6.1 Hz, 1 H, OH-2'), 6.25 (d, J = 6.3 Hz, 1 H, H-1'), 6.65 (d, J = 6.1 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, H-1'), 6.65 (d, J = 6.1 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, H-1'), 6.65 (d, J = 6.1 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, H-1'), 6.65 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, H-1'), 6.65 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, OH-2'), 6.27 (d, J = 6.3 Hz, 1 H, OH-2'), 6.28
- 355 8.4 Hz, 2 H, H-Ph), 7.12 (d, J = 8.5 Hz, 2 H, H-Ph), 7.64 (s, 1 H, H-6), 8.63 (s, 1 H, H-2).  $^{13}$ C NMR (100)
- 356 MHz, DMSO-d<sub>6</sub>) δ: 22.6 (CH<sub>3</sub>), 61.6 (C-5'), 70.6 (C-3'), 73.9 (C-2'), 85.1 (C-4'), 86.4 (C-1'), 113.6 (2 C-1')
- 357 Ph), 116.2 (C-4a), 117.7 (C-5), 121.1 (C-Ph), 123.5 (C-6), 130.5 (2 C-Ph), 147.9 (C-Ph), 150.5 (C-7a),
- 358 150.5 (C-2), 159.2 (C-4). HRMS (ESI): calculated for  $C_{18}H_{21}N_4O_4$  ([M+H]<sup>+</sup>): 357.1557, found: 357.1553.
- 359 4.2.5. 4-Methyl-5-(pyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (18). The iodo-
- 360 precursor 55 (100 mg, 0.26 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (30 mg, 0.026 mmol, 0.1 eq.) were added to a 10 mL two-
- neck round-bottom flask purged with argon. Anhydrous DMF (2 mL) and 2-(tributylstannyl)pyridine (125
- 362 μl, 0.39 mmol, 1.5 eq.) were added with syringes. The mixture was heated to 100 °C. When full conversion
- of **55** was observed via LC-MS analysis (1 h), the reaction mixture was cooled and evaporated to dryness.
- The residue was purified by column chromatography ( $0 \rightarrow 10\%$  MeOH/DCM) to give **18** (70 mg, 0.20
- 365 mmol) as a white solid in 79% yield. <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ )  $\delta$ : 2.71 (s, 3 H, CH<sub>3</sub>), 3.57 (ddd, J =
- 366 12.0, 5.8, 4.0 Hz, 1 H, H-5"), 3.66 (ddd, *J* = 12.0, 5.2, 3.9 Hz, 1 H, H-5"), 3.94 (q, *J* = 3.8 Hz, 1 H, H-4"),
- 367 4.15 (td, J = 4.8, 3.5 Hz, 1 H, H-3'), 4.48 (dd, J = 11.5, 6.2 Hz, 1 H, H-2'), 5.08 (t, J = 5.6 Hz, 1 H, OH-
- 368 5'), 5.17 (d, J = 4.7 Hz, 1 H, OH-3'), 5.40 (d, J = 6.4 Hz, 1 H, OH-2'), 6.29 (d, J = 6.2 Hz, 1 H, H-1'), 7.35
- 369 (ddd, J = 7.5, 4.9, 1.0 Hz, 1 H, H-Pyr), 7.71 (dt, J = 7.9, 1.0 Hz, 1 H, H-Pyr), 7.88 (td, J = 7.7, 1.9 Hz, 1 H,
- 370 H-Pyr), 8.14 (s, 1 H, H-6), 8.66 8.70 (m, 1 H, H-Pyr), 8.71 (s, 1 H, H-2). <sup>13</sup>C NMR (75 MHz, DMSO-d<sub>6</sub>)
- δ: 24.2 (CH<sub>3</sub>), 61.4 (C-5'), 70.5 (C-3'), 74.0 (C-2'), 85.2 (C-4'), 86.6 (C-1'), 115.4 (C-4a), 117.2 (C-5),
- 372 121.7 (C-Pyr), 123.3 (C-Pyr), 126.7 (C-6), 136.7 (C-Pyr), 149.0 (C-Pyr), 150.9 (C-Pyr), 151.1 (C-7a), 153.2
- 373 (C-2), 160.3 (C-4). HRMS (ESI): calculated for  $C_{17}H_{19}N_4O_4$  ([M+H]<sup>+</sup>): 343.1401, found: 343.1405.
- **4.2.6.** 4-Methyl-5-(thiophen-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (19[27]). 55 (100
- mg, 0.26 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (30 mg, 0.026 mmol, 0.1 eq.) were added to a 10 mL two-neck round-
- 376 bottom flask flushed with argon to which anhydrous DMF (2 mL) was added, followed by
- tributyl(thiophen-2-yl)stannane (124 µl, 0.39 mmol, 1.5 eq.). The mixture was heated at 100 °C. When full

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378 conversion of 55 was observed via LC-MS analysis (1 h), the reaction mixture was cooled and evaporated
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- to dryness. The residue was purified by column chromatography ( $0 \rightarrow 10\%$  MeOH/DCM) to give 19 (70
- 380 mg, 0.20 mmol) as a white solid in 78% yield. <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ )  $\delta$ : 2.54 (s, 3 H, CH<sub>3</sub>), 3.56
- 381 (ddd, J = 11.8, 5.7, 4.0 Hz, 1 H, H-5''), 3.65 (ddd, J = 11.8, 5.2, 4.0 Hz, 1 H, H-5'), 3.94 (q, J = 3.8 Hz, 1 Hz)
- 382 H, H-4'), 4.06 4.19 (m, 1 H, H-3'), 4.45 (dd, J = 11.4, 6.2 Hz, 1 H, H-2'), 5.08 (t, J = 5.4 Hz, 1 H, OH-
- 383 5'), 5.17 (d, J = 4.7 Hz, 1 H, OH-3'), 5.39 (d, J = 6.4 Hz, 1 H, OH-2'), 6.26 (d, J = 6.4 Hz, 1 H, H-1'), 7.18
- 384 (dd, J = 5.3, 3.5 Hz, 1 H, H-Thio), 7.21 7.28 (m, 1 H, H-Thio), 7.61 (dd, J = 5.0, 1.2 Hz, 1 H, H-Thio),
- 385 7.95 (s, 1 H, H-6), 8.71 (s, 1 H, H-2).  $^{13}$ C NMR (75 MHz, DMSO- $d_6$ )  $\delta$ : 22.5 (CH<sub>3</sub>), 61.4 (C-5'), 70.5 (C-
- 386 3'), 74.1 (C-2'), 85.3 (C-4'), 86.5 (C-1'), 108.9 (C-5), 116.1 (C-4a), 126.0 (C-6), 126.2 (C-Thio), 127.6 (C-
- 387 Thio), 128.1 (C-Thio), 134.7 (C-Thio), 150.5 (C-7a), 151.1 (C-2), 159.4 (C-4). HRMS (ESI): calculated for
- 388  $C_{16}H_{18}N_3O_4S$  ([M+H]<sup>+</sup>): 348.1013, found: 348.1010.
- 389 4.2.7. 4-Methyl-5-(pyridin-3-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (20). The title
- compound was prepared according to General procedure A. **55** (200 mg, 0.51 mmol) gave rise to **20** (63
- 391 mg, 0.18 mmol) as a white solid in 36% yield. H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.47 (s, 3 H, CH<sub>3</sub>), 3.55
- 392 (dd,  $J = 11.4, 3.1 \text{ Hz}, 1 \text{ H}, \text{H-5}^{"}$ ), 3.65 (d,  $J = 11.9, 3.6 \text{ Hz}, 1 \text{ H}, \text{H-5}^{"}$ ), 3.94 (q,  $J = 3.7 \text{ Hz}, 1 \text{ H}, \text{H-4}^{"}$ ), 4.14
- 393 (br. s., 1 H, H-3'), 4.48 (br. s., 1 H, H-2'), 5.05 (br. s., 1 H, OH-5'), 5.18 (br. s., 1 H, OH-3'), 5.39 (d, J =
- 394 4.6 Hz, 1 H, OH-2'), 6.28 (d, J = 6.1 Hz, 1 H, H-1'), 7.51 (dd, J = 7.8, 4.8 Hz, 1 H, CH), 7.96 (t, J = 1.8
- 395 Hz, 1 H, H-Pyr), 7.98 (s, 1 H, H-6), 8.60 (dd, J = 4.8, 1.5 Hz, 1 H, H-Pyr), 8.73 (s, 1 H, CH, H-2), 8.74 (d,
- 396 J = 2.1 Hz, 1 H, CH, H-Pyr). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 23.1 (CH<sub>3</sub>),  $\delta$ 1.7 (C-5'), 70.7 (C-3'), 74.2
- 397 (C-2'), 85.4 (C-4'), 86.8 (C-1'), 113.5 (C-5), 116.0 (C-4a), 123.5 (C-Pyr), 125.8 (C-6), 130.4 (C-Pyr), 137.4
- 398 (C-Pyr), 148.4 (C-Pyr), 150.0 (C-Pyr), 151.0 (C-7a), 151.1 (C-2), 159.4 (C-4). HRMS (ESI) calculated for
- 399  $C_{17}H_{19}N_4O_4^+([M+H^+])$ : 343.1401, found: 343.1407.
- 400 4.2.8. 4-Methyl-5-(pyridin-4-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (21). The iodo-
- 401 precursor 55 (150 mg, 0.38 mmol), pyridin-4-ylboronic acid (71 mg, 0.58 mmol, 1.5 eq.), Pd<sub>2</sub>(dba)<sub>3</sub> (17 mg,
- 402 0.019 mmol, 0.05 eq.), P(c-Hexyl)<sub>3</sub> (13 mg, 0.046 mmol, 0.12 eq.) and K<sub>2</sub>PO<sub>4</sub> (1.27 M, 0.5 mL, 1.33

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       mL/mmol SM) were added to a 10 mL round-bottom flask purged with with argon. 1,4-Dioxane (1 mL,
       2.67 mL/mmol SM) was added via syringe and the reaction mixture was stirred at ambient temperature for
404
405
       5 min, and then stirred at 100 °C. Upon completion of the reaction as observed via LC-MS analysis, the
406
       reaction mixture was cooled to room temperature, neutralized with 0.5 M aq. HCl, and evaporated. The
407
       residue was resuspended in MeOH and then evaporated in vacuo, and this was repeated three times. The
408
       residue was adsorbed on Celite® and washed through a short silica pad using 20% MeOH/DCM. The
409
       resulting solution was evaporated in vacuo and purified by column chromatography (0 -> 10%
410
       MeOH/DCM) to give 21 (59 mg, 0.17 mmol) as a white solid in 45% yield. <sup>1</sup>H NMR (400 MHz, DMSO-
       d_6) \delta: 2.54 (s, 3 H, CH<sub>3</sub>), 3.54 - 3.57 (m, 1 H, H-5"), 3.64 - 3.67 (m, 1 H, H-5"), 3.94 (q, J = 3.6 Hz, 1 H,
411
412
       H-4'), 4.14 (br. s., 1 H, H-3'), 4.47 (q, J = 5.4 Hz, 1 H, H-2'), 5.08 (br. s., 1 H, OH-5'), 5.20 (d, J = 3.4 Hz,
413
       1 H, OH-3'), 5.41 (d, J = 5.9 Hz, 1 H, OH-2'), 6.28 (d, J = 6.0 Hz, 1 H, H-1'), 7.56 - 7.57 (m, 2 H, H-pyr),
414
       8.06 (s, 1 H, H-6), 8.63 - 8.65 (m, 2 H, H-Pyr), 8.74 (s, 1 H, H-2). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 23.4
       (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.1 (C-2'), 85.3 (C-4'), 86.7 (C-1'), 114.6 (C-5), 115.3 (C-4a), 124.5 (2
415
       C, C-Pyr), 126.3 (C-6), 142.2 (C-Pyr), 149.4 (2 C, C-Pyr), 151.0 (C-7a), 151.1 (C-2), 159.4 (C-4). HRMS
416
417
       (ESI): calculated for C_{17}H_{19}N_4O_4 ([M+H]<sup>+</sup>): 343.1401, found: 343.1402.
418
       4.2.9. 4-Methyl-5-(2-fluorophenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (22). Compound
419
       22 was prepared according to General procedure A. 55 (100 mg, 0.26 mmol) and (2-fluorophenyl)boronic
420
       acid (44 mg, 0.31 mmol) gave rise to 22 (60 mg, 0.17 mmol) as a white solid in 60% yield. <sup>1</sup>H NMR (300
421
       MHz, DMSO-d_0) \delta: 2.38 (s, 1 H, CH<sub>3</sub>), 3.55 (ddd, J = 12.0, 5.6, 4.0 Hz, 1 H, H-5''), 3.64 (ddd, J = 12.0,
       5.2, 4.0 Hz, 1 H, H-5'), 3.94 (q, J = 3.7 Hz, 1 H, H-4'), 4.08 - 4.19 (m, 1 H, H-3'), 4.48 (dd, J = 11.5, 6.4
422
423
       Hz, 1 H, H-2'), 5.05 (t, J = 5.4 Hz, 1 H, OH-5'), 5.18 (d, J = 4.7 Hz, 1 H, OH-3'), 5.39 (d, J = 6.4 Hz, 1 H,
424
       OH-2'), 6.27 (d, J = 6.2 Hz, 1 H, H-1'), 7.24 - 7.42 (m, 2 H, H-Ph), 7.44 - 7.58 (m, 2 H, H-Ph), 7.91 (s, 1
       H, H-6), 8.71 (s, 1 H, H-2). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 21.3 (CH<sub>3</sub>), 61.5 (C-5'), 70.6 (C-3'), 74.0
425
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(C-2'), 85.3 (C-4'), 86.7 (C-1'), 109.5 (C-5), 115.5 (d, J = 21.8 Hz, 1 C, C-Ph), 116.5 (C-4a), 122.0 (d, J = 21.8 Hz, 1 C, C-Ph)

16.0 Hz, 1 C, C-Ph), 124.6 (d, *J* = 2.9 Hz, 1 C, C-Ph), 125.8 (C-6), 130.0 (d, *J* = 8.0 Hz, 1 C, C-Ph), 132.6

426

- 428 (d, J = 2.9 Hz, 1 C, C-Ph), 150.5 (C-7a), 150.9 (C-2), 159.2 (C-4), 159.8 (d, J = 244.1 Hz, 1 C, C-Ph). <sup>19</sup>F
- NMR (376 MHz, DMSO- $d_6$ ) δ: -114.29. HRMS (ESI): calculated for  $C_{18}H_{19}FN_3O_4$  ([M+H]<sup>+</sup>): 360.1354,
- 430 found: 360.1356.
- 431 4.2.10. 4-Methyl-5-(2-chlorophenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (23).
- Compound 23 was prepared according to General procedure A. 55 (100 mg, 0.26 mmol) and (2-
- chlorophenyl)boronic acid (61 mg, 0.39 mmol) gave rise to 23 (30 mg, 0.08 mmol) as a white solid in 31%
- 434 yield. <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ ) δ: 2.28 (s, 3 H, CH<sub>3</sub>), 3.55 (ddd, J = 11.8, 5.6, 4.2 Hz, 1 H, H-5''),
- 3.65 (ddd,  $J = 11.8, 5.3, 4.0 \,\text{Hz}, 1 \,\text{H}, \text{H--5'}$ ), 3.94 (q,  $J = 3.8 \,\text{Hz}, 1 \,\text{H}, \text{H--4'}$ ), 4.07 4.18 (m, 1 H, H-3'), 4.47
- 436 (dd, J = 11.5, 6.1 Hz, 1 H, H-2'), 5.04 (t, J = 5.4 Hz, 1 H, OH-5'), 5.18 (d, J = 5.0 Hz, 1 H, OH-3'), 5.38
- 437 (d, J = 6.4 Hz, 1 H, OH-2'), 6.26 (d, J = 6.2 Hz, 1 H, H-1'), 7.39 7.55 (m, 3 H, H-Ph), 7.56 7.70 (m, 1
- 438 H, H-Ph), 7.86 (s, 1 H, H-6), 8.70 (s, 1 H, H-2).  $^{13}$ C NMR (75 MHz, DMSO- $d_6$ )  $\delta$ : 21.1 (CH<sub>3</sub>), 61.5 (C-5'),
- 439 70.5 (C-3'), 74.1 (C-2'), 85.2 (C-4'), 86.8 (C-1'), 113.4 (C-5), 116.7 (C-4a), 125.4 (C-6), 127.2 (C-Ph),
- 440 129.3 (2 C, C-Ph), 129.8 (C-Ph), 133.3 (C-Ph), 134.0 (C-Ph), 150.2 (C-7a), 150.9 (C-2), 159.0 (C-4).
- 441 HRMS (ESI): calculated for  $C_{18}H_{19}ClN_3O_4$  ([M+H]<sup>+</sup>): 376.1059, found: 376.1066.
- 442 4.2.11. 4-Methyl-5-(2-methoxyphenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (24).
- 443 Compound 24 was prepared according to General procedure A. 55 (150 mg, 0.38 mmol) and (2-
- methoxyphenyl)boronic acid (87 mg, 0.58 mmol) gave rise to **24** (100 mg, 0.27 mmol) as a white solid in
- 445 71% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ) δ: 2.28 (s, 3 H, CH<sub>3</sub>), 3.50 3.57 (m, 1 H, H-5''), 3.58 3.67
- 446 (m, 1 H, H-5'), 3.72 (s, 3 H, OCH<sub>3</sub>), 3.92 (q, J = 3.6 Hz, 1 H, H-4'), 4.10 4.13 (m, 1 H, H-3'), 4.48 (q, J
- 447 = 6.3 Hz, 1 H, H-2', 5.04 (t, J = 5.5 Hz, 1 H, OH-5', 5.16 (d, J = 4.8 Hz, 1 H, OH-3'), 5.36 (d, J = 6.5 Hz, 1 H, OH-3')
- 448 1 H, OH-2'), 6.24 (d, J = 6.4 Hz, 1 H, H-1'), 7.03 (td, J = 7.4, 0.9 Hz, 1 H, H-Ph), 7.11 (d, J = 7.8 Hz, 1 H,
- 449 H-Ph), 7.29 (dd, J = 7.4, 1.8 Hz, 1 H, H-Ph), 7.37 7.47 (m, 1 H, H-Ph), 7.70 (s, 1 H, H-6), 8.65 (s, 1 H,
- 450 H-2). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ: 20.8 (CH<sub>3</sub>), 55.0 (OCH<sub>3</sub>), 61.6 (C-5'), 70.6 (C-3'), 73.8 (C-2'),
- 451 85.2 (C-4'), 86.5 (C-1'), 110.8 (C-Ph), 113.1 (C-5), 117.2 (C-4a), 120.3 (C-Ph), 123.1 (C-Ph), 124.7 (C-6),

- 452 129.4 (C-Ph), 131.6 (C-Ph), 150.4 (C-7a), 150.6 (C-2), 157.2 (C-Ph), 159.5 (C-4). HRMS (ESI): calculated
- 453 for  $C_{19}H_{22}N_3O_5$  ([M+H]<sup>+</sup>): 372.1554, found: 372.1558.
- 454 4.2.12. 4-Methyl-5-(2-(methylthio)phenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (25).
- Compound 25 was prepared according to General procedure A. 55 (150 mg, 0.38 mmol) and (2-
- (methylthio)phenyl)boronic acid (97 mg, 0.58 mmol) gave rise to 25 (115 mg, 0.30 mmol) as a white solid
- 457 in 78% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.23 (s, 3 H, CH<sub>3</sub>), 2.37 (s, 3 H, SCH<sub>3</sub>), 3.49 3.58 (m, 1
- 458 H, H-5''), 3.60 3.68 (m, 1 H, H-5'), 3.93 (q, J = 3.6 Hz, 1 H, H-4'), 4.09 4.14 (m, 1 H, H-3'), 4.44 (q, J
- $459 = 6.0 \text{ Hz}, 1 \text{ H}, \text{H-}2'), 5.05 \text{ (t, } J = 5.1 \text{ Hz}, 1 \text{ H}, \text{OH-}5'), 5.17 \text{ (d, } J = 4.8 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ H}, \text{OH-}3'), 5.36 \text{ (d, } J = 6.4 \text{ Hz}, 1 \text{ Hz$
- 460 1 H, OH-2'), 6.24 (d, J = 6.0 Hz, 1 H, H-1'), 7.20 7.25 (m, 1 H, H-Ph), 7.26 7.30 (m, 1 H, H-Ph), 7.34
- 461 (d, J = 7.5 Hz, 1 H, H-Ph), 7.41 7.48 (m, 1 H, H-Ph), 7.76 (s, 1 H, H-6), 8.67 (s, 1 H, H-2). <sup>13</sup>C NMR (100)
- 462 MHz, DMSO-*d*<sub>6</sub>) δ: 14.2 (SCH<sub>3</sub>), 21.0 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.1 (C-2'), 85.1 (C-4'), 86.8 (C-
- 463 1'), 114.0 (C-5), 116.7 (C-4a), 123.8 (C-Ph), 124.0 (C-Ph), 125.2 (C-6), 128.8 (C-Ph), 131.2 (C-Ph), 132.1
- 464 (C-Ph), 139.9 (C-Ph), 150.2 (C-7a), 150.8 (C-2), 159.2 (C-4). HRMS (ESI): calculated for C<sub>19</sub>H<sub>22</sub>N<sub>3</sub>O<sub>4</sub>S
- 465 ([M+H]<sup>+</sup>): 388.1326, found: 388.1313.
- 466 4.2.13. 4-Methyl-5-(2-nitrophenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (26). Compound
- 467 **26** was prepared according to General procedure A. **55** (200 mg, 0.51 mmol) and (2-nitrophenyl)boronic
- acid (129 mg, 0.77 mmol) gave rise to **26** (135 mg, 0.35 mmol) as a yellow solid in 69% yield. <sup>1</sup>H NMR
- 469 (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.22 (s, 3 H, CH<sub>3</sub>), 3.51 3.56 (m, 1 H, H-5''), 3.61 3.66 (m, 1 H, H-5'), 3.93
- 470 (q, J = 3.6 Hz, 1 H, H-4'), 4.10 (q, J = 4.2 Hz, 1 H, H-3'), 4.42 (q, J = 5.8 Hz, 1 H, H-2'), 5.03 (t, J = 5.4
- 471 Hz, 1 H, OH-5'), 5.18 (d, J = 4.9 Hz, 1 H, OH-3'), 5.39 (d, J = 6.3 Hz, 1 H, OH-2'), 6.23 (d, J = 6.0 Hz, 1
- 472 H, H-1'), 7.65 (d, J = 7.5 Hz, 1 H, H-Ph), 7.70 7.75 (m, 1 H, H-Ph), 7.80 7.83 (m, 1 H, H-Ph), 7.87 (s,
- 473 1 H, H-6), 8.12 (d, J = 7.9 Hz, 1 H, H-Ph), 8.71 (s, 1 H, H-2). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ) δ: 21.3
- 474 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.1 (C-2'), 85.2 (C-4'), 86.9 (C-1'), 111.1 (C-5), 116.9 (C-4a), 124.1 (C-5)
- 475 Ph), 125.2 (C-6), 128.4 (C-Ph), 129.6 (C-Ph), 132.9 (C-Ph), 133.9 (C-Ph), 149.8 (C-Ph), 150.2 (C-7a),
- 476 151.1 (C-2), 158.9 (C-4). HRMS (ESI): calculated for C<sub>18</sub>H<sub>19</sub>N<sub>4</sub>O<sub>6</sub> ([M+H]<sup>+</sup>): 387.1299, found: 387.1310.

- 4.2.14. 4-Methyl-5-(2-aminophenyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (27).
- 478 Compound 26 (60 mg, 0.16 mmol) was dissolved in MeOH (2 mL). The flask was purged into a N<sub>2</sub>
- atmosphere. Next, a cat. amount of Pd(OH)<sub>2</sub>/C was added into the reaction solution. Then, the N<sub>2</sub> gas was
- exchanged with H<sub>2</sub> gas (balloon; bubbling). The reaction mixture was stirred at ambient temperature until
- 481 LC-MS analysis showed full conversion of 26. Then, the balloon was removed and the mixture was filtered
- over Celite® and rinsed by MeOH. The filtrate was evaporated until dryness. The residue was purified by
- column chromatography (5  $\rightarrow$  20% MeOH/DCM) to give 27 (38 mg, 0.11 mmol) as a white solid in 69%
- 484 yield. H NMR (400 MHz, DMSO- $d_6$ ) δ: 2.35 (s, 3 H, CH<sub>3</sub>), 3.52 3.58 (m, 1 H, H-5''), 3.62 3.67 (m, 1
- 485 H, H-5'), 3.93 (q, J = 3.5 Hz, 1 H, H-4'), 4.14 (q, J = 4.2 Hz, 1 H, H-3'), 4.44 (q, J = 5.8 Hz, 1 H, H-2'),
- 486 4.71 (br. s., 2 H, NH<sub>2</sub>), 5.05 (t, J = 5.4 Hz, 1 H, OH-5'), 5.14 (d, J = 4.9 Hz, 1 H, OH-3'), 5.33 (d, J = 5.9
- 487 Hz, 1 H, OH-2'), 6.26 (d, J = 5.8 Hz, 1 H, H-1'), 6.61 (td, J = 7.3, 0.9 Hz, 1 H, H-Ph), 6.76 (d, J = 7.5 Hz,
- 488 1 H, H-Ph), 7.02 (dd, *J* = 7.4, 1.4 Hz, 1 H, H-Ph), 7.08 7.12 (m, 1 H, H-Ph), 7.69 (s, 1 H, H-6), 8.65 (s, 1
- 489 H, H-2).  ${}^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 20.8 (CH<sub>3</sub>), 61.4 (C-5'), 70.5 (C-3'), 74.1 (C-2'), 85.0 (C-4'),
- 490 86.8 (C-1'), 113.3 (C-5), 114.2 (C-Ph), 115.8 (C-Ph), 117.0 (C-4a), 118.4 (C-Ph), 124.9 (C-6), 128.7 (C-
- 491 Ph), 131.4 (C-Ph), 147.2 (C-Ph), 150.5 (C-7a), 150.7 (C-2), 159.4 (C-4), HRMS (ESI): calculated for
- 492  $C_{18}H_{21}N_4O_4$  ([M+H]<sup>+</sup>): 357.1557, found: 357.1546.
- 493 4.2.15. 4-Methyl-5-(pyrimidin-2-yl)-N7-( $\beta$ -D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (28).
- Compound 28 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 28 (55
- mg, 0.16 mmol) as a white solid in 63% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.99 (s, 3)
- 496 H, CH<sub>3</sub>), 3.56 3.61 (m, 1 H, H-5''), 3.63 3.68 (m, 1 H, H-5'), 3.97 (q, *J* = 3.3 Hz, 1 H, H-4'), 4.14 (q, *J*
- 497 = 4.2 Hz, 1 H, H-3'), 4.48 (q, J = 6.0 Hz, 1 H, H-2'), 5.11 (t, J = 5.2 Hz, 1 H, OH-5'), 5.19 (d, J = 4.6 Hz,
- 498 1 H, OH-3'), 5.41 (d, J = 6.3 Hz, 1 H, OH-2'), 6.32 (d, J = 6.3 Hz, 1 H, H-1'), 7.38 (t, J = 4.8 Hz, 1 H, H-
- 499 pyrimidine), 8.51 (s, 1 H, H-6), 8.73 (s, 1 H, H-2), 8.88 (d, J = 4.9 Hz, 2 H, H-pyrimidine).  $^{13}$ C NMR (100
- 500 MHz, DMSO- $d_6$ )  $\delta$ : 25.4 (CH<sub>3</sub>), 61.4 (C-5'), 70.6 (C-3'), 74.3 (C-2'), 85.4 (C-4'), 86.6 (C-1'), 115.0 (C-
- 501 4a), 116.2 (C-5), 118.7 (C-pyrimidine), 130.2 (C-6), 151.1 (C-2), 151.6 (C-7a), 157.3 (2 C, C-pyrimidine),

- 502 161.0 (C-4), 161.6 (C-pyrimidine). HRMS (ESI): calculated for C<sub>16</sub>H<sub>18</sub>N<sub>5</sub>O<sub>4</sub> ([M+H]<sup>+</sup>): 344.1353, found:
- 503 344.1359.
- 4.2.16. 4-Methyl-5-(pyrazin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (29). Compound
- 29 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 29 (35 mg, 0.10
- 506 mmol) as a white solid in 37% yield for two steps.  $^{1}$ H NMR (400 MHz, DMSO- $d_{6}$ )  $\delta$ : 2.73 (s, 3 H, CH<sub>3</sub>),
- 507  $3.55 3.60 \text{ (m, 1 H, H-5'')}, 3.65 3.71 \text{ (m, 1 H, H-5')}, 3.95 \text{ (d, } J = 3.4 \text{ Hz, 1 H, H-4')}, 4.16 \text{ (q, } J = 4.2 \text{ Hz, } J = 4.2 \text{ Hz,$
- 508 1 H, H-3'), 4.49 (q, J = 5.7 Hz, 1 H, H-2'), 5.09 (t, J = 5.4 Hz, 1 H, OH-5'), 5.20 (d, J = 4.9 Hz, 1 H, OH-
- 509 3'), 5.43 (d, J = 6.3 Hz, 1 H, OH-2'), 6.30 (d, J = 5.9 Hz, 1 H, H-1'), 8.36 (s, 1 H, H-6), 8.59 (d, J = 2.1 Hz,
- 510 1 H, H-pyrazin), 8.74 8.75 (m, 2 H, H-pyrazin and H-2), 9.02 (s, 1 H, H-pyrazin). <sup>13</sup>C NMR (100 MHz,
- 511 DMSO- $d_6$ )  $\delta$ : 24.4 (CH<sub>3</sub>),  $\delta$ 1.4 (C-5'), 70.4 (C-3'), 74.1 (C-2'), 85.3 (C-4'), 86.8 (C-1'), 113.6 (C-4a), 115.3
- 512 (C-5), 127.8 (C-6), 142.2 (C-pyrazin), 143.7 (C-pyrazin), 144.1 (C-pyrazin), 149.2 (C-pyrazin), 151.3 (C-
- 513 2), 151.3 (C-7a), 160.4 (C-4). HRMS (ESI): calculated for  $C_{16}H_{18}N_5O_4$  ([M+H]<sup>+</sup>): 344.1353, found:
- 514 344.1350.
- 515 4.2.17. 4-Methyl-5-(pyrimidin-5-yl)-N7-( $\beta$ -D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (30).
- 516 Compound 30 was prepared according to General procedure A. 55 (150 mg, 0.38 mmol) gave rise to 30 (70
- 517 mg, 0.20 mmol) as a white solid in 54% yield.  $^{1}$ H NMR (400 MHz, DMSO- $d_{6}$ )  $\delta$ : 3.54 3.58 (m, 1 H, H-
- 518 5''), 3.63 3.68 (m, 1 H, H-5'), 3.94 (q, J = 3.8 Hz, 1 H, H-4'), 4.14 (q, J = 4.2 Hz, 1 H, H-3'), 4.47 (q, J = 4.2 Hz, J = 4.2 Hz
- 5.5 = 5.8 Hz, 1 H, H-2', 5.04 (t, J = 5.5 Hz, 1 H, OH-5', 5.20 (d, J = 4.9 Hz, 1 H, OH-3'), 5.41 (d, J = 6.3 Hz, 1 H, OH-3')
- 520 1 H, OH-2'), 6.29 (d, J = 6.0 Hz, 1 H, H-1'), 8.10 (s, 1 H, H-6), 8.75 (s, 1 H, H-2), 9.02 (s, 2 H, 2H-
- 521 pyrimdine), 9.22 (s, 1 H, H-pyrimdine). (the peak of CH<sub>3</sub> is covered in the solvent peak). <sup>13</sup>C NMR (100
- 522 MHz, DMSO-d<sub>6</sub>) δ: 23.0 (CH<sub>3</sub>), 61.5 (C-5'), 70.4 (C-3'), 74.1 (C-2'), 85.3 (C-4'), 86.7 (C-1'), 109.6 (C-1')
- 523 5), 115.7 (C-4a), 126.4 (C-6), 128.6 (C-pyrimdine), 150.9 (C-7a), 151.2 (C-2), 156.8 (2 C, C-pyrimdine),
- 524 156.9 (C-pyrimdine), 159.3 (C-4). HRMS (ESI): calculated for  $C_{16}H_{18}N_5O_4$  ([M+H]<sup>+</sup>): 344.1353, found:
- 525 344.1359.

- 526 4.2.18. 4-Methyl-5-(3-chloropyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (31).
- 527 Compound 31 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 31 (54
- 528 mg, 0.14 mmol) as a white solid in 33% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.35 (s, 3
- 529 H, CH<sub>3</sub>), 3.53 3.59 (m, 1 H, H-5''), 3.62 3.68 (m, 1 H, H-5'), 3.95 (q, J = 3.5 Hz, 1 H, H-4'), 4.13 (q, J
- 530 = 4.2 Hz, 1 H, H-3'), 4.46 (q, J = 6.0 Hz, 1 H, H-2'), 5.07 (t, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz,
- 531 1 H, OH-3'), 5.43 (d, J = 6.4 Hz, 1 H, OH-2'), 6.29 (d, J = 6.0 Hz, 1 H, H-1'), 7.49 (dd, J = 8.2, 4.7 Hz, 1
- 532 H, H-Pyr), 8.10 (dd, J = 8.1, 1.5 Hz, 1 H, H-Pyr), 8.12 (s, 1 H, H-6), 8.66 (dd, J = 4.6, 1.5 Hz, 1 H, H-Pyr),
- 533 8.73 (s, 1 H, H-2).  ${}^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 22.2 (CH<sub>3</sub>), 61.4 (C-5'), 70.5 (C-3'), 74.3 (C-2'),
- 534 85.3 (C-4'), 86.9 (C-1'), 113.1 (C-5), 116.3 (C-4a), 124.1 (C-Pyr), 127.1 (C-6), 131.2 (C-Pyr), 137.7 (C-
- 535 Pyr), 147.7 (C-Pyr), 150.3 (C-7a), 150.9 (C-2), 151.1 (C-Pyr), 159.3 (C-4). HRMS (ESI): calculated for
- 536  $C_{17}H_{18}CIN_4O_4$  ([M+H]<sup>+</sup>): 377.1011, found: 377.1003.
- 537 4.2.19. 4-Methyl-5-(4-chloropyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (32).
- Compound 32 was prepared according to General procedure B. 57 (2.00 g, 2.84 mmol) gave rise to a slightly
- impure 32 (500 mg, 1.32 mmol) as a white solid. 32 was purified by preparative RP-HPLC gradient: 0.2%
- formic acid in water:MeCN at a flow rate of 20 mL/min; The initial gradient composition (95% A/05% B)
- was held for 2.0 min, increased to 57% B in 10 min, then increased to 100% B in 1.5 min as a white solid
- 542 (310 mg, 0.45 mmol) in 29% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ: 2.74 (s, 3 H, CH<sub>3</sub>),
- 543 3.55 3.59 (m, 1 H, H-5''), 3.64 3.70 (m, 1 H, H-5'), 3.94 (q, J = 3.9 Hz, 1 H, H-4'), 4.15 (q, J = 4.2 Hz,
- 544 1 H, H-3'), 4.48 (q, J = 6.1 Hz, 1 H, H-2'), 5.06 (t, J = 5.6 Hz, 1 H, OH-5'), 5.19 (d, J = 5.0 Hz, 1 H, OH-
- 3'), 5.42 (d, J = 6.3 Hz, 1 H, OH-2'), 6.29 (d, J = 6.0 Hz, 1 H, H-1'), 7.49 (dd, J = 5.4, 2.0 Hz, 1 H, H-Pyr),
- 546 7.90 (d, J = 1.9 Hz, 1 H, H-Pyr), 8.27 (s, 1 H, H-6), 8.66 (d, J = 5.4 Hz, 1 H, H-Pyr), 8.73 (s, 1 H, H-2). <sup>13</sup>C
- NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ: 24.5 (CH<sub>3</sub>), 61.4 (C-5'), 70.4 (C-3'), 74.0 (C-2'), 85.2 (C-4'), 86.6 (C-1'),
- 548 115.2 (C-4a), 116.0 (C-5), 121.6 (C-Pyr), 122.7 (C-Pyr), 127.7 (C-6), 143.2 (C-Pyr), 150.4 (C-Pyr), 151.1
- 549 (C-2), 151.2 (C-7a), 154.9 (C-Pyr), 160.5 (C-4). HRMS (ESI): calculated for C<sub>17</sub>H<sub>18</sub>ClN<sub>4</sub>O<sub>4</sub> ([M+H]<sup>+</sup>):
- 550 377.1011, found: 377.1004.

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4.2.20. 4-Methyl-5-(4-methoxypyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (33).
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- 552 Compound 33 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 33 (28
- mg, 0.075 mmol) as a white solid in 23% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.71 (s, 3)
- 554 H, CH<sub>3</sub>), 3.54 3.59 (m, 1 H, H-5''), 3.64 3.69 (m, 1 H, H-5'), 3.90 (s, 3 H, OCH<sub>3</sub>), 3.94 (q, J = 3.8 Hz,
- 555 1 H, H-4'), 4.15 (q, J = 4.2 Hz, 1 H, H-3'), 4.48 (q, J = 6.0 Hz, 1 H, H-2'), 5.09 (t, J = 5.4 Hz, 1 H, OH-5'),
- 556 5.18 (d, J = 4.9 Hz, 1 H, OH-3'), 5.41 (d, J = 6.3 Hz, 1 H, OH-2'), 6.28 (d, J = 6.0 Hz, 1 H, H-1'), 6.95 (dd,
- 557 J = 5.8, 2.5 Hz, 1 H, H-Pyr, 7.25 (d, <math>J = 2.4 Hz, 1 H, H-Pyr), 8.15 (s, 1 H, H-6), 8.49 (d, <math>J = 5.8 Hz, 1 H, H-Pyr)
- 558 H-Pyr), 8.70 (s, 1 H, H-2).  ${}^{13}$ C NMR (100 MHz, DMSO- $d_6$ ) δ: 24.2 (CH<sub>3</sub>), 55.4 (CH<sub>3</sub>), 61.4 (C-5'), 70.4
- 559 (C-3'), 74.0 (C-2'), 85.1 (C-4'), 86.6 (C-1'), 108.2 (C-Pyr), 109.1 (C-Pyr), 115.5 (C-4a), 117.3 (C-5), 126.7
- 560 (C-6), 150.3 (C-Pyr), 150.9 (C-2), 151.0 (C-7a), 154.8 (C-Pyr), 160.3 (C-4), 165.6 (C-Pyr). HRMS (ESI):
- 561 calculated for  $C_{18}H_{21}N_4O_5$  ([M+H]<sup>+</sup>): 373.1506, found: 373.1500.

#### 562 4.2.21. 4-Methyl-5-(5-chloropyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (34).

- Compound 34 was prepared according to General procedure B. 57 (2.00 g, 2.84 mmol) gave rise to 34 (430
- mg, 1.14 mmol) as a white solid. 34 was purified by preparative RP-HPLC gradient: 0.2% formic acid in
- water:MeCN at a flow rate of 20 mL/min; The initial gradient composition (95% A/05% B) was held for
- 2.0 min, increased to 57% B in 10 min, then increased to 100% B in 1.5 min as a white solid (250 mg, 0.66
- 567 mmol) in 22% yield for two steps. H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.71 (s, 3 H, CH<sub>3</sub>), 3.54 3.59 (m, 1
- 568 H, H-5''), 3.64 3.69 (m, 1 H, H-5'), 3.94 (q, J = 3.8 Hz, 1 H, H-4'), 4.15 (q, J = 4.2 Hz, 1 H, H-3'), 4.48
- 569 (q, J = 5.8 Hz, 1 H, H-2'), 5.07 (t, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 4.8 Hz, 1 H, OH-3'), 5.41 (d, J = 4.8 Hz, 1 Hz, 1
- 570 6.3 Hz, 1 H, OH-2'), 6.29 (d, J = 6.0 Hz, 1 H, H-1'), 7.77 (d, J = 8.5 Hz, 1 H, H-Pyr), 8.02 (dd, J = 8.4, 2.6
- 571 Hz, 1 H, H-Pyr), 8.20 (s, 1 H, H-6), 8.72 8.73 (m, 2 H, H-Pyr and H-2).  ${}^{13}$ C NMR (100 MHz, DMSO- $d_6$ )
- δ: 24.4 (CH<sub>3</sub>), 61.4 (C-5'), 70.4 (C-3'), 74.0 (C-2'), 85.2 (C-4'), 86.6 (C-1'), 115.2 (C-4a), 116.0 (C-5),
- 573 124.4 (C-Pyr), 127.2 (C-6), 129.0 (C-Pyr), 136.6 (C-Pyr), 147.5 (C-Pyr), 151.1 (C-7a), 151.2 (C-2), 151.8
- 574 (C-Pyr), 160.3 (C-4). HRMS (ESI): calculated for  $C_{17}H_{18}ClN_4O_4$  ([M+H]<sup>+</sup>): 377.1011, found: 377.1019.

- 575 4.2.22. 4-Methyl-5-(6-chloropyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (35).
- 576 Compound 35 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 35 (25
- mg, 0.066 mmol) as a white solid in 18% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.76 (s, 3)
- 578 H, CH<sub>3</sub>), 3.57 (ddd, J = 11.9, 5.7, 4.1 Hz, 1 H, H-5''), 3.65 3.69 (m, 1 H, H-5'), 3.95 (q, J = 3.8 Hz, 1 H,
- 579 H-4'), 4.15 (q, J = 4.2 Hz, 1 H, H-3'), 4.47 (q, J = 6.0 Hz, 1 H, H-2'), 5.08 (t, J = 5.5 Hz, 1 H, OH-5'), 5.19
- 580 (d, J = 5.0 Hz, 1 H, OH-3'), 5.42 (d, J = 6.3 Hz, 1 H, OH-2'), 6.29 (d, J = 6.0 Hz, 1 H, H-1'), 7.46 (dd, J = 6.0 Hz, 1 H, OH-3')
- 581 7.9, 0.5 Hz, 1 H, H-Pyr), 7.76 (dd, J = 7.8, 0.5 Hz, 1 H, H-Pyr), 7.93 7.96 (m, 1 H, H-Pyr), 8.28 (s, 1 H,
- 582 H-6), 8.73 (s, 1 H, H-2).  $^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 24.6 (CH<sub>3</sub>), 61.4 (C-5'), 70.4 (C-3'), 74.1 (C-
- 583 2'), 85.2 (C-4'), 86.7 (C-1'), 115.1 (C-4a), 115.6 (C-5), 121.8 (C-Pyr), 127.6 (C-6), 140.3 (C-Pyr), 149.4
- 584 (C-Pyr), 151.2 (C-2), 151.2 (C-7a), 153.8 (C-Pyr), 160.4 (C-4). HRMS (ESI): calculated for C<sub>17</sub>H<sub>18</sub>ClN<sub>4</sub>O<sub>4</sub>
- 585 ([M+H]<sup>+</sup>): 377.1011, found: 377.1019.
- 586 4.2.23. 4-Methyl-5-(6-fluoropyridin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (36).
- Compound **36** was prepared according to General procedure B. **57** (200 mg, 0.28 mmol) gave rise to **36** (4
- 588 mg, 0.011 mmol) as a white solid in 5% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.76 (s, 3
- 589 H, CH<sub>3</sub>), 3.54 3.60 (m, 1 H, H-5''), 3.64 3.70 (m, 1 H, H-5'), 3.95 (q, J = 3.7 Hz, 1 H, H-4'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), 4.15 (q, J = 3.7 Hz, 1 H, H-5'), J = 3.7
- = 4.3 Hz, 1 H, H-3'), 4.47 (q, J = 5.8 Hz, 1 H, H-2'), 5.09 (t, J = 5.5 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 Hz, 1
- 591 1 H, OH-3'), 5.42 (d, J = 6.3 Hz, 1 H, OH-2'), 6.29 (d, J = 6.0 Hz, 1 H, H-1'), 7.13 (dd, J = 8.1, 2.6 Hz, 1
- 592 H, H-Pyr), 7.69 (dd, J = 7.4, 2.4 Hz, 1 H, H-Pyr), 8.07 (q, J = 8.3 Hz, 1 H, H-Pyr), 8.26 (s, 1 H, H-6), 8.73
- 593 (s, 1 H, H-2).  ${}^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 24.5 (CH<sub>3</sub>), 61.4 (C-5'), 70.4 (C-3'), 74.1 (C-2'), 85.3 (C-
- 594 4'), 86.7 (C-1'), 106.9 (d, *J* = 37.8 Hz, 1 C, C-Pyr), 115.1 (C-4a), 115.5 (C-5), 120.7 (d, *J* = 3.6 Hz, 1 C,
- 595 C-Pyr), 127.6 (C-6), 142.6 (d, J = 8.0 Hz, 1 C, C-Pyr), 151.1 (C-2), 151.2 (C-7a), 151.8(d, J = 14.5 Hz, 1
- 596 C, C-Pyr), 160.2 (C-4), 162.3 (d, J = 235.4 Hz, 1 C, C-Pyr). <sup>19</sup>F NMR (376 MHz, DMSO- $d_6$ )  $\delta$ : -68.00.
- 597 HRMS (ESI): calculated for  $C_{17}H_{18}FN_4O_4$  ([M+H]<sup>+</sup>): 361.1307, found: 361.1319.
- 598 4.2.24. 4-Methyl-5-(6-chloropyridin-3-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (37).
- 599 Compound 37 was prepared according to General procedure A. 55 (100 mg, 0.26 mmol) and (6-

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600 chloropyridin-3-yl)boronic acid (49 mg, 0.31 mmol) gave rise to 37 (25 mg, 0.066 mmol) as a light grey
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- solid in 26% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.48 (s, 3 H, CH<sub>3</sub>), 3.50 3.60 (m, 1 H, H-5''), 3.60
- 602 3.71 (m, 1 H, H-5'), 3.94 (q, J = 3.8 Hz, 1 H, H-4'), 4.07 4.17 (m, 1 H, H-3'), 4.46 (t, J = 5.4 Hz, 1 H,
- 603 H-2'), 5.05 (br. s., 1 H, OH-5'), 5.21 (br. s., 1 H, OH-3'), 5.42 (br. s., 1 H, OH-2'), 6.28 (d, J = 6.0 Hz, 1
- 604 H, H-1'), 7.63 (d, J = 8.3 Hz, 1 H, H-Pyr), 8.01 (s, 1 H, H-6), 8.05 (dd, J = 8.2, 2.6 Hz, 1 H, H-Pyr), 8.58
- 605 (d, J = 2.4 Hz, 1 H, H-Pyr), 8.73 (s, 1 H, H-2). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 23.0 (CH<sub>3</sub>), 61.5 (C-5'),
- 606 70.4 (C-3'), 74.1 (C-2'), 85.2 (C-4'), 86.7 (C-1'), 111.9 (C-5), 115.7 (C-4a), 123.8 (C-Pyr), 125.9 (C-6),
- 607 129.7 (C-Pyr), 140.7 (C-Pyr), 149.0 (C-Pyr), 149.9 (C-Pyr), 150.8 (C-7a), 151.1 (C-2), 159.3 (C-4). HRMS
- 608 (ESI): calculated for  $C_{17}H_{18}ClN_4O_4$  ([M+H]<sup>+</sup>): 377.1011, found: 377.1006.
- 4.2.25. 4-Methyl-5-(1*H*-pyrrol-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-*d*]pyrimidine (38). The iodo-
- precursor 55 (150 mg, 0.38 mmol), (1-(tert-butoxycarbonyl)-1*H*-pyrrol-2-yl)boronic acid (122 mg, 0.57
- 611 mmol, 1.5 eq.), Pd(OAc)<sub>2</sub> (9 mg, 0.038 mmol, 0.1 eq.), TPPTS (43 mg, 0.076 mmol, 0.2 eq.) and Na<sub>2</sub>CO<sub>3</sub>
- 612 (121 mg, 1.14 mmol, 3 eq.) were added to a 10 mL round-bottom flask, purged with argon. Next, a
- 613 MeCN/water (1/2 ratio, 6 mL/mmol SM) was added via syringe. The reaction mixture was stirred at ambient
- temperature for 5 min, and then at 100 °C. Upon completion of the reaction as monitored by LC-MS analysis,
- the reaction mixture was cooled to room temperature and evaporated *in vacuo*. The residue was treated with
- 616 0.5 M NaOMe in MeOH (5 mL) and stirred at ambient temperature for 1 h, after which it was neutralized
- with 0.5 M aq. HCl, and evaporated. The residue was purified by column chromatography (5  $\rightarrow$  20%
- 618 MeOH/DCM) to give 38 (35 mg, 0.11 mmol) as a yellow solid in 28% yield for two steps. <sup>1</sup>H NMR (400
- 619 MHz, DMSO- $d_6$ )  $\delta$ : 3.52 3.57 (m, 1 H, H-5''), 3.60 3.66 (m, 1 H, H-5'), 3.92 (q, J = 3.6 Hz, 1 H, H-4'),
- 620 4.10 4.13 (m, 1 H, H-3'), 4.43 (q, J = 6.2 Hz, 1 H, H-2'), 5.05 (t, J = 5.4 Hz, 1 H, OH-5'), 5.17 (d, J = 4.9
- 621 Hz, 1 H, OH-3'), 5.34 (d, J = 6.5 Hz, 1 H, OH-2'), 6.12 (q, J = 2.8 Hz, 1 H, H- pyrrole), 6.16 (br. s., 1 H,
- H-pyrrole), 6.25 (d, J = 6.1 Hz, 1 H, H-1'), 6.86 (br. s., 1 H, H-pyrrole), 7.76 (s, 1 H, H-6), 8.66 (s, 1 H, H-
- 2), 11.04 (br. s., 1 H, NH). (the peak of CH<sub>3</sub> is covered in the solvent peak). <sup>13</sup>C NMR (100 MHz, DMSO-
- 624 *d*<sub>6</sub>) δ: 22.1 (CH<sub>3</sub>), 61.6 (C-5'), 70.6 (C-3'), 74.1 (C-2'), 85.2 (C-4'), 86.4 (C-1'), 108.1 (C-pyrrole), 108.6

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                (C-pyrrole), 109.3 (C-5), 116.3 (C-4a), 118.2 (C-pyrrole), 123.5 (C-6), 124.6 (C-pyrrole), 150.3 (C-7a),
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                 150.8 (C-2), 159.5 (C-4). HRMS (ESI): calculated for C_{16}H_{19}N_4O_4 ([M+H]+): 331.1401, found: 331.1414.
                4.2.26. 4-Methyl-5-(thiazol-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (39). Compound
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                57 (200 mg, 0.28 mmol) and Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub> (8 mg, 0.011 mmol, 0.04 eq.) were added to a 10 mL two-neck
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                round-bottom flask. The flask was charged with argon. Next, anhydrous DMF (2 mL) was added into the
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                flask via syringe, followed by the addition of 2-tributylstannyltriazole (179 µl, 0.57 mmol, 2.0 eq.). The
                mixture was heated at 100 °C. When full conversion of 57 was observed via LC-MS analysis (2 days), the
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632
                reaction mixture was cooled and evaporated until dryness. The residue was dissolved in 7 N NH<sub>3</sub> methanol
                solution. The mixture was stirred at ambient temperature overnight. Next, the reaction solution was
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634
                evaporated until dryness and purified by column chromatography (0 \rightarrow 10\% MeOH/DCM) to give 39 (55
                mg, 0.16 mmol) as a white solid in 56% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO-d_6) \delta: 2.86 (s, 3
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636
                H, CH<sub>3</sub>), 3.56 - 3.62 (m, 1 H, H-5''), 3.69 (dt, J = 11.9, 4.5 Hz, 1 H, H-5'), 3.96 (q, J = 3.4 Hz, 1 H, H-4'),
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                4.16 (q, J = 4.4 Hz, 1 H, H-3'), 4.47 (q, J = 5.7 Hz, 1 H, H-2'), 5.15 (t, J = 5.3 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 H, OH-5'), 5.19 (d, J = 5.4 Hz, 1 Hz,
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                = 4.9 \text{ Hz}, 1 H, OH-3'), 5.45 (d, J = 6.1 \text{ Hz}, 1 H, OH-2'), 6.28 (d, J = 5.9 \text{ Hz}, 1 H, H-1'), 7.76 (d, J = 3.3
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                Hz, 1 H, H-thiazole), 7.96 (d, J = 3.3 Hz, 1 H, H-thiazole), 8.37 (s, 1 H, H-6), 8.75 (s, 1 H, H-2). ^{13}C NMR
                (100 MHz, DMSO-d<sub>6</sub>) δ: 23.8 (CH<sub>3</sub>), 61.2 (C-5'), 70.3 (C-3'), 74.2 (C-2'), 85.3 (C-4'), 86.9 (C-1'), 110.3
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# 4.2.27. 4-Methyl-5-(1*H*-pyrazol-3-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-*d*]pyrimidine (40).

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644 Compound 40 was prepared according to General procedure A. 55 (120 mg, 0.31 mmol) and (1*H*-pyrazol-

3-yl)boronic acid (51 mg, 0.46 mmol) gave rise to **40** (40 mg, 0.12 mmol) as a white solid in 39% yield. <sup>1</sup>H

(C-5), 114.6 (C-4a), 119.9 (C-thiazole), 128.0 (C-6), 143.3 (C-thiazole), 151.0 (C-7a), 151.5 (C-2), 160.6

(C-4), 160.8 (C-thiazole). HRMS (ESI): calculated for  $C_{15}H_{17}N_4O_4S$  ([M+H]<sup>+</sup>): 349.0965, found: 349.0960.

NMR (400 MHz, DMSO-d<sub>6</sub>) δ: 2.77 (s, 3 H, CH<sub>3</sub>), 3.53 - 3.59 (m, 1 H, H-5''), 3.62 - 3.67 (m, 1 H, H-5'),

3.93 (d, J = 3.1 Hz, 1 H, H-4'), 4.13 (q, J = 4.3 Hz, 1 H, H-3'), 4.45 (q, J = 5.9 Hz, 1 H, H-2'), 5.08 (br. s., 1 H, H-2'), 5.08 (br. s., 2 Hz, 2 H

648 1 H, OH-5'), 5.17 (br. s., 1 H, OH-3'), 5.38 (d, J = 6.3 Hz, 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, H-1'),

649 6.52 (br. s., 1 H, H-pyrazole), 7.82 (s, 1 H, H-pyrazole), 7.96 (s, 1 H, H-6), 8.67 (s, 1 H, H-2), 12.91 (br. s.,

- 650 1 H, NH). ). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ: 23.4 (CH<sub>3</sub>), 61.5 (C-5'), 70.6 (C-3'), 74.0 (C-2'), 85.2 (C-
- 651 4'), 86.5 (C-1'), 104.8 (C-pyrazole), 110.3 (C-5), 115.8 (C-4a), 125.3 (C-6), 129.1 (C-pyrazole), 144.7 (C-
- 652 pyrazole), 150.7 (C-2), 150.8 (C-7a), 160.1 (C-4). HRMS (ESI): calculated for  $C_{15}H_{18}N_5O_4$  ([M+H]<sup>+</sup>):
- 653 332.1353, found: 332.1334.
- 4.2.28. 4-Methyl-5-(1-methyl-1*H*-pyrazol-4-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-*d*]pyrimidine
- 655 (41). Compound 41 was prepared according to General procedure A. 55 (150 mg, 0.38 mmol) and (1-
- methyl-1*H*-pyrazol-4-yl)boronic acid (73 mg, 0.58 mmol) gave rise to **41** (14 mg, 0.041 mmol) as a white
- solid in 11% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.57 (s, 3 H, CH<sub>3</sub>), 3.51 3.57 (m, 1 H, H-5''), 3.60
- -3.65 (m, 1 H, H-5'), 3.90 3.93 (m, 4 H, CH<sub>3</sub> and H-4'), 4.09 4.13 (m, 1 H, H-3'), 4.43 (q, J = 5.9 Hz, 1
- 659 H, H-2'), 5.04 (t, J = 5.4 Hz, 1 H, OH-5'), 5.15 (d, J = 4.8 Hz, 1 H, OH-3'), 5.35 (d, J = 6.3 Hz, 1 H, OH-
- 660 2'), 6.23 (d, *J* = 6.3 Hz, 1 H, H-1'), 7.60 (s, 1 H, H-pyrazole), 7.73 (s, 1 H, H-6), 7.90 (s, 1 H, H-pyrazole),
- 8.65 (s, 1 H, H-2).  ${}^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 22.5 (CH<sub>3</sub>), 38.6 (CH<sub>3</sub>), 61.5 (C-5'), 70.6 (C-3'),
- 74.0 (C-2'), 85.1 (C-4'), 86.4 (C-1'), 107.5 (C-pyrazole), 113.7 (C-5), 116.1 (C-4a), 124.3 (C-6), 130.2
- 663 (C-pyrazole), 139.0 (C-pyrazole), 150.5 (C-7a), 150.7 (C-2), 159.3 (C-4). HRMS (ESI): calculated for
- 664  $C_{16}H_{20}N_5O_4$  ([M+H]<sup>+</sup>): 346.1510, found: 346.1503.
- 665 4.2.29. 4-Methyl-5-(thiophen-3-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (42[27]).
- 666 Compound 42 was prepared according to General procedure A. 55 (200 mg, 0.51 mmol) and thiophen-3-
- ylboronic acid (98 mg, 0.77 mmol) gave rise to 42 (56 mg, 0.16 mmol) as a white solid in 32% yield. <sup>1</sup>H
- NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.52 (s, 3 H, CH<sub>3</sub>), 3.55 (d, J = 11.4 Hz, 1 H, H-5''), 3.60 3.68 (m, 1 H,
- 669 H-5'), 3.89 3.98 (m, 1 H, H-4'), 4.12 (br. s., 1 H, H-3'), 4.45 (br. s., 1 H, H-2'), 5.06 (br. s., 1 H, OH-5'),
- 5.17 (br. s., 1 H, OH-3'), 5.36 (br. s., 1 H, OH-2'), 6.26 (d, J = 6.3 Hz, 1 H, H-1'), 7.31 (dd, J = 4.9, 1.3 Hz,
- 671 1 H, H-Thio), 7.59 (dd, J = 2.9, 1.3 Hz, 1 H, H-Thio), 7.67 (dd, J = 4.9, 3 Hz, 1 H, H-Thio), 7.86 (s, 1 H,
- 672 H-6), 8.69 (s, 1 H, H-2).  $^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 22.6 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.0 (C-
- 673 2'), 85.2 (C-4'), 86.5 (C-1'), 111.7 (C-5), 116.0 (C-4a), 123.6 (C-Thio), 124.8 (C-6), 126.1 (C-Thio), 129.7

- 674 (C-Thio), 134.2 (C-Thio) 150.5 (C-7a), 150.7 (C-2), 159.3 (C-4). HRMS (ESI) calculated for  $C_{16}H_{18}N_3O_4S^+$
- 675 ([M+H<sup>+</sup>]): 348.1013, found: 348.1030.
- 4-Methyl-5-(furan-3-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (43[27]).
- 677 Compound 43 was prepared according to General procedure A. 55 (100 mg, 0.26 mmol) gave rise to 43 (46
- 678 mg, 0.14 mmol) as a white solid in 53% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.60 (s, 3 H, CH<sub>3</sub>), 3.52
- -3.57 (m, 1 H, H-5''), 3.61 3.66 (m, 1 H, H-5'), 3.92 (q, J = 3.7 Hz, 1 H, H-4'), 4.12 (q, J = 4.2 Hz, 1 H,
- 680 H-3'), 4.43 (q, J = 6.1 Hz, 1 H, H-2'), 5.05 (t, J = 5.4 Hz, 1 H, OH-5'), 5.16 (d, J = 4.9 Hz, 1 H, OH-3'),
- 5.35 (d, J = 6.4 Hz, 1 H, OH-2'), 6.24 (d, J = 6.3 Hz, 1 H, H-1'), 6.77 (d, J = 1.0 Hz, 1 H, H-furan), 7.78 -
- 7.80 (m, 1 H, H-furan), 7.82 (s, 1 H, H-6), 7.87 (s, 1 H, H-furan), 8.67 (s, 1 H, H-2). <sup>13</sup>C NMR (100 MHz,
- 683 DMSO- $d_6$ )  $\delta$ : 22.6 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.0 (C-2'), 85.2 (C-4'), 86.4 (C-1'), 107.1 (C-furan),
- 684 112.7 (C-5), 116.1 (C-4a), 118.2 (C-furan), 124.7 (C-6), 140.5 (C-furan), 143.4 (C-furan), 150.7 (C-7a),
- 685 150.8 (C-2), 159.3 (C-4). HRMS (ESI): calculated for  $C_{16}H_{18}N_3O_5$  ([M+H]<sup>+</sup>): 332.1241, found: 332.1246.
- 4.2.31. 4-Methyl-5-(quinolin-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (44). Compound
- 44 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to a slight impure
- 688 44 (50 mg) as a white solid. 44 was purified by preparative RP-HPLC gradient: 0.2% formic acid in
- water: MeCN at a flow rate of 20 mL/min; The initial gradient composition (95% A/05% B) was held for
- 690 2.0 min, increased to 60% B in 13 min, then increased to 100% B in 1 min as a white solid (33 mg, 0.084)
- 691 mmol) in 22% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ: 2.93 (s, 3 H, CH<sub>3</sub>), 3.57 3.61 (m, 1
- 692 H, H-5''), 3.68 3.72 (m, 1 H, H-5'), 3.97 (q, J = 3.7 Hz, 1 H, H-4'), 4.18 (t, J = 4.2 Hz, 1 H, H-3'), 4.52
- 693 (t, *J* = 5.4 Hz, 1 H, H-2'), 5.13 (br. s., 1 H, OH-5'), 5.22 (br. s., 1 H, OH-3'), 5.46 (br. s., 1 H, OH-2'), 6.33
- 694 (d, J = 5.9 Hz, 1 H, H-1'), 7.58 7.62 (m, 1 H, H-quinoline), 7.76 7.80 (m, 1 H, H-quinoline), 7.95 (d, J
- 695 = 8.5 Hz, 1 H, H-quinoline), 8.01 (d, J = 7.6 Hz, 1 H, H-quinoline), 8.06 (d, J = 8.4 Hz, 1 H, H-quinoline),
- 8.42 -8.44 (m, 2 H, H-quinoline and H-6), 8.75 (s, 1 H, H-2).  $^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 25.2 (CH<sub>3</sub>),
- 697 61.4 (C-5'), 70.4 (C-3'), 74.0 (C-2'), 85.2 (C-4'), 86.7 (C-1'), 115.5 (C-4a), 117.5 (C-5), 121.3 (C-
- 698 quinoline), 126.2 (C-quinoline), 126.2 (C-quinoline), 127.9 (C-quinoline), 128.1 (C-6), 128.4 (C-quinoline),

- 699 129.9 (C-quinoline), 136.5 (C-quinoline), 147.2 (C-quinoline), 151.1 (C-2), 151.5 (C-7a), 153.1 (C-
- 700 quinoline), 161.0 (C-4). HRMS (ESI): calculated for  $C_{21}H_{21}N_4O_4$  ([M+H]+): 393.1557, found: 393.1563.
- 701 4.2.32. 4-Methyl-5-(isoquinolin-3-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (45).
- Compound 45 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 45 (21
- mg, 0.054 mmol) as a white solid in 19% yield for two steps.  $^{1}$ H NMR (400 MHz, DMSO- $d_{6}$ )  $\delta$ : 2.68 (s, 3)
- 704 H, CH<sub>3</sub>), 3.56 3.60 (m, 1 H, H-5''), 3.64 3.70 (m, 1 H, H-5'), 3.96 (q, J = 3.8 Hz, 1 H, H-4'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), 4.16 (q, J = 3.8 Hz, 1 H, H-5'), J = 3.8
- 705 = 4.2 Hz, 1 H, H-3'), 4.51 (q, J = 6.1 Hz, 1 H, H-2'), 5.08 (t, J = 5.5 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz,
- 706 1 H, OH-3'), 5.42 (d, J = 6.4 Hz, 1 H, OH-2'), 6.32 (d, J = 6.3 Hz, 1 H, H-1'), 7.67 7.71 (m, 1 H, H-1')
- 707 quinoline), 7.82 (td, J = 7.5, 1.1 Hz, 1 H, H-quinoline), 8.02 (d, J = 8.1 Hz, 1 H, H-quinoline), 8.11 (s, 1 H,
- 708 H-6), 8.16 8.18 (m, 2 H, 2 H-quinoline), 8.73 (s, 1 H, H-2), 9.44 (s, 1 H, H-quinoline). <sup>13</sup>C NMR (100
- 709 MHz, DMSO-*d*<sub>6</sub>) δ: 24.0 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.1 (C-2'), 85.2 (C-4'), 86.5 (C-1'), 115.8 (C-1')
- 710 4a), 117.4 (C-5), 119.1 (C-6), 126.3 (C-quinoline), 126.5 (C-quinoline), 126.8 (C-quinoline), 127.3 (C-
- 711 quinoline), 127.6 (C-quinoline), 130.9 (C-quinoline), 135.8 (C-quinoline), 146.9 (C-quinoline), 150.9 (C-
- 712 2), 151.1 (C-7a), 151.8 (C-quinoline), 161.0 (C-4). HRMS (ESI): calculated for  $C_{21}H_{21}N_4O_4$  ([M+H]<sup>+</sup>):
- 713 393.1557, found: 393.1549.
- 714 4.2.33. 4-Methyl-5-(1*H*-indol-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (46). The iodo-
- precursor **55** (150 mg, 0.38 mmol), (1-(tert-butoxycarbonyl)-1*H*-indol-2-yl)boronic acid (149 mg, 0.57
- 716 mmol, 1.5 eq.), Pd(OAc)<sub>2</sub> (9 mg, 0.038 mmol, 0.1 eq.), TPPTS (43 mg, 0.076 mmol, 0.2 eq.) and Na<sub>2</sub>CO<sub>3</sub>
- 717 (121 mg, 1.14 mmol, 3 eq.) were added to a 10 mL round-bottom flask purged with argon. MeCN/water
- 718 (1/2 ratio, 3 mL, 6 mL/mmol SM) was added and the reaction mixture was stirred at ambient temperature
- for 5 min, and then at 100 °C. Upon complete conversion of the SM (LC-MS), the reaction mixture was
- cooled to room temperature and evaporated *in vacuo*. The residue was treated with 0.5 M NaOMe in MeOH
- 721 (5 mL) at ambient temperature for 1 h. The mixture was neutralized with 0.5 M aq. HCl and evaporated.
- The residue was purified by column chromatography (5  $\rightarrow$  20% MeOH/DCM) to give 46 (68 mg, 0.18
- 723 mmol) as a light yellow solid in 47% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.61 (s, 3 H,

- 724 CH<sub>3</sub>), 3.55 3.60 (m, 1 H, H-5''), 3.63 3.68 (m, 1 H, H-5'), 3.96 (q, J = 3.6 Hz, 1 H, H-4'), 4.15 (q, J =
- 725 4.2 Hz, 1 H, H-3'), 4.47 (q, J = 6.0 Hz, 1 H, H-2'), 5.07 (t, J = 5.4 Hz, 1 H, OH-5'), 5.20 (d, J = 4.9 Hz, 1
- 726 H, OH-3'), 5.40 (d, J = 6.4 Hz, 1 H, OH-2'), 6.30 (d, J = 6.1 Hz, 1 H, H-1'), 6.58 (d, J = 1.3 Hz, 1 H, H-
- 727 indole), 7.00 7.04 (m, 1 H, H-indole), 7.09 7.13 (m, 1 H, H-indole), 7.40 (d, *J* = 7.8 Hz, 1 H, H-indole),
- 728 7.56 (d, J = 7.8 Hz, 1 H, H-indole), 8.01 (s, 1 H, H-6), 8.72 (s, 1 H, H-2), 11.37 (s, 1 H, NH). <sup>13</sup>C NMR
- 729 (100 MHz, DMSO-d<sub>6</sub>) δ: 22.6 (CH<sub>3</sub>), 61.5 (C-5'), 70.6 (C-3'), 74.2 (C-2'), 85.3 (C-4'), 86.6 (C-1'), 101.9
- 730 (C-indole), 108.7 (C-5), 111.0 (C-indole), 115.9 (C-4a), 119.1 (C-indole), 119.7 (C-indole), 121.1 (C-indole)
- 731 indole), 125.8 (C-6), 128.3 (C-indole), 131.3 (C-indole), 136.5 (C-indole), 150.6 (C-7a), 151.0 (C-2), 159.7
- 732 (C-4). HRMS (ESI): calculated for  $C_{20}H_{21}N_4O_4$  ([M+H]<sup>+</sup>): 381.1557, found: 381.1577.
- 4.2.34. 4-Methyl-5-(benzofuran-2-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (47).
- 734 Compound 47 was prepared according to General procedure A. 55 (150 mg, 0.38 mmol) gave rise to 47 (20
- 735 mg, 0.052 mmol) as a white solid in 14% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.79 (s, 3 H, CH<sub>3</sub>), 3.55
- -3.63 (m, 1 H, H-5'), 3.64-3.72 (m, 1 H, H-5''), 3.96 (d, J = 3.3 Hz, 1 H, H-4'), 4.16 (d, J = 3.5 Hz, 1 H,
- 737 H-3'), 4.49 (q, J = 5.8 Hz, 1 H, H-2'), 5.11 (t, J = 5.3 Hz, 1 H, OH-5'), 5.21 (d, J = 4.8 Hz, 1 H, OH-3'),
- 738 5.44 (d, J = 6.3 Hz, 1 H, OH-2'), 6.30 (d, J = 6 Hz, 1 H, H-1'), 7.16 (s, 1 H, H-benzofuran), 7.24 7.36 (m,
- 739 2 H, H-benzofuran), 7.66 (d, J = 18.7 Hz, 2 H, H-benzofuran), 8.32 (s, 1 H, H-6), 8.76 (s, 1 H, H-2). <sup>13</sup>C
- 740 NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 23.3 (CH<sub>3</sub>), 61.4 (C-5'), 70.5 (C-3'), 74.2 (C-2'), 85.4 (C-4'), 86.7 (C-1'),
- 741 104.3 (C-benzofuran), 106.1 (C-5), 110.9 (C-benzofuran), 115.0 (C-4a), 120.9 (C-benzofuran), 123.1 (C-
- 742 benzofuran), 124.2 (C-benzofuran), 127.0 (C-6), 128.7 (C-benzofuran), 150.3 (C-benzofuran), 150.9 (C-
- 743 7a), 151.4 (C-2), 154.2 (C-benzofuran), 159.7 (C-4). HRMS (ESI) calculated for  $C_{20}H_{20}N_3O_5^+$  ([M+H<sup>+</sup>]):
- 744 382.1397, found: 382.1391.
- 4.2.35. 4-Methyl-5-(quinolin-8-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (48). Compound
- 48 was prepared according to General procedure B. 57 (200 mg, 0.28 mmol) gave rise to 48 (37 mg, 0.094)
- mmol) as a white solid in 46% yield for two steps. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 1.91 (s, 3 H, CH<sub>3</sub>),
- 748 3.52 3.57 (m, 1 H, H-5''), 3.61 3.66 (m, 1 H, H-5'), 3.95 (d, J = 3.4 Hz, 1 H, H-4'), 4.13 (q, J = 4.2 Hz,

- 749 1 H, H-3'), 4.53 (q, J = 6.1 Hz, 1 H, H-2'), 5.03 (t, J = 5.3 Hz, 1 H, OH-5'), 5.18 (d, J = 4.8 Hz, 1 H, OH-
- 750 3'), 5.40 (d, J = 6.5 Hz, 1 H, OH-2'), 6.31 (d, J = 6.3 Hz, 1 H, H-1'), 7.57 (dd, J = 8.3, 4.1 Hz, 1 H, H-
- 751 quinoline), 7.70 (dd, J = 8.1, 7.1 Hz, 1 H, H-quinoline), 7.81 (dd, J = 7.0, 1.5 Hz, 1 H, H-quinoline), 7.85
- 752 (s, 1 H, H-6), 8.07 (dd, J = 8.3, 1.4 Hz, 1 H, H-quinoline), 8.46 (dd, J = 8.3, 1.7 Hz, 1 H, H-quinoline), 8.68
- 753 (s, 1 H, H-2), 8.82 (dd, J = 4.1, 1.8 Hz, 1 H, H-quinoline). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 21.4 (CH<sub>3</sub>),
- 754 61.6 (C-5'), 70.6 (C-3'), 73.9 (C-2'), 85.2 (C-4'), 86.7 (C-1'), 114.6 (C-5), 118.4 (C-4a), 121.6 (C-
- 755 quinoline), 125.2 (C-6), 126.3 (C-quinoline), 128.1 (C-quinoline), 128.3 (C-quinoline), 131.4 (C-quinoline),
- 756 133.7 (C-quinoline), 136.4 (C-quinoline), 147.0 (C-quinoline), 150.3 (C-quinoline), 150.5 (C-7a), 150.6
- 757 (C-2), 159.4 (C-4). HRMS (ESI): calculated for  $C_{21}H_{21}N_4O_4$  ([M+H]+): 393.1557, found: 393.1568.
- 758 4.2.36. 4-Methyl-5-(1*H*-indol-7-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (49). Compound
- 49 was prepared according to General procedure A. 55 (120 mg, 0.31 mmol) gave rise to 48 (75 mg, 0.20
- 760 mmol) as a white solid in 64% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.11 (s, 3 H, CH<sub>3</sub>), 3.52 3.57 (m,
- 761 1 H, H-5''), 3.62 3.67 (m, 1 H, H-5'), 3.94 (q, J = 3.5 Hz, 1 H, H-4'), 4.16 (q, J = 4.8 Hz, 1 H, H-3'), 4.51
- 762 (q, J = 5.8 Hz, 1 H, H-2'), 5.02 (t, J = 5.4 Hz, 1 H, OH-5'), 5.18 (d, J = 5.0 Hz, 1 H, OH-3'), 5.35 (d, J = 5.0 Hz, 1 H, OH-5'), 5.35 (d, J = 5.0 Hz, 1 H, OH-
- 763 6.3 Hz, 1 H, OH-2'), 6.32 (d, J = 5.8 Hz, 1 H, H-1'), 6.51 (dd, J = 2.9, 1.9 Hz, 1 H, H-indole), 7.06 7.11
- 764 (m, 2 H, 2 H-indole), 7.27 (t, J = 2.8 Hz, 1 H, H-indole), 7.60 (dd, J = 6.6, 2.3 Hz, 1 H, H-indole), 7.86 (s,
- 765 1 H), 8.70 (s, 1 H), 10.83 (br. s., 1 H, NH). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 21.1 (CH<sub>3</sub>), 61.4 (C-5'),
- 766 70.5 (C-3'), 74.1 (C-2'), 85.0 (C-4'), 86.9 (C-1'), 101.5 (C-indole), 113.0 (C-5), 117.2 (C-4a), 118.2 (C-
- 767 indole), 118.9 (C-indole), 119.7 (C-indole), 123.1 (C-indole), 125.1 (C-6), 125.7 (C-indole), 127.8 (C-indole)
- 768 indole), 135.5 (C-indole), 150.5 (C-7a), 150.7 (C-2), 159.3 (C-4). HRMS (ESI): calculated for C<sub>20</sub>H<sub>21</sub>N<sub>4</sub>O<sub>4</sub>
- 769 ([M+H]<sup>+</sup>): 381.1557, found: 381.1553.
- 770 4.2.37. 4-Methyl-5-(1-methyl-1*H*-indol-7-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-*d*]pyrimidine (50).
- Compound **50** was prepared according to General procedure A. **55** (150 mg, 0.38 mmol) gave rise to 49 (75
- mg, 0.11 mmol) as a white solid in 30% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.07 (s, 3 H, CH<sub>3</sub>), 3.22
- 773 (d, J = 14.3 Hz, 3 H, CH<sub>3</sub>), 3.51 3.58 (m, 1 H, H-5''), 3.60 3.68 (m, 1 H, H-5'), 3.93 3.96 (m, 1 H, H-5'')

- 774 4'), 4.11 4.16 (m, 1 H, H-3'), 4.47 (dq, J = 15.7, 5.8 Hz, 1 H, H-2'), 5.03 (dt, J = 9.0, 5.4 Hz, 1 H, OH-
- 5'), 5.18 (d, J = 4.6 Hz, 1 H, OH-3'), 5.40 (dd, J = 6.2, 4.3 Hz, 1 H, OH-2'), 6.30 (t, J = 5.6 Hz, 1 H, H-1'),
- 776 6.51 (d, J = 3.1 Hz, 1 H, H-indole), 7.05 (dtd, J = 19.8, 7.3, 7.3, 1.8 Hz, 2 H, 2 H-indole), 7.27 (dd, J = 4.4,
- 3.2 Hz, 1 H, H-indole), 7.61 7.63 (m, 1 H, H-indole), 7.88 (d, J = 5.8 Hz, 1 H, H-6), 8.70 (s, 1 H, H-2).
- 778  $^{13}$ C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 20.9 (CH<sub>3</sub>), 35.3 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.1 (C-2'), 85.1 (C-
- 4'), 86.7 (C-1'), 100.6 (C-indole), 113.2 (C-5), 117.6 (C-indole), 118.4 (C-4a), 118.7 (C-indole), 120.5 (C-indole), 120
- 780 indole), 125.3 (C-indole), 125.6 (C-6), 129.1 (C-indole), 131.5 (C-indole), 134.8 (C-indole), 149.9 (C-7a),
- 781 151.0 (C-2), 159.3 (C-4). HRMS (ESI): calculated for  $C_{21}H_{23}N_4O_4$  ([M+H]<sup>+</sup>): 395.1714, found: 395.1738.
- 4.2.38. 4-Methyl-5-(dibenzo[b,d]furan-4-yl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (51).
- Compound **51** was prepared according to General procedure A. **55** (150 mg, 0.38 mmol) gave rise to **51** (13
- 784 mg, 0.030 mmol) as a white solid in 8% yield. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 2.22 (s, 3 H, CH<sub>3</sub>), 3.53
- 785 3.61 (m, 1 H, H-5'), 3.62 3.70 (m, 1 H, H-5''), 3.97 (d, J = 3.4 Hz, 1 H, H-4'), 4.15 (d, J = 3.9 Hz, 1 H,
- 786 H-3'), 4.55 (q, J = 5.9 Hz, 1 H, H-2'), 5.05 (t, J = 5.3 Hz, 1 H, OH-5'), 5.20 (dd, J = 4.7, 2.1 Hz, 1 H, OH-
- 787 3'), 5.44 (dd, J = 6.4, 2.0 Hz, 1 H, OH-2'), 6.33 (d, J = 6.1 Hz, 1 H, H-1'), 7.44 (t, J = 7.6 Hz, 1 H, H-
- 788 dibenzofuran), 7.47 7.56 (m, 2 H, H-dibenzofuran), 7.59 (dd, *J* = 7.0, 0.8 Hz, 1 H, H-dibenzofuran), 7.67
- 789 (d, J = 2.8 Hz, 1 H, H-dibenzofuran), 8.05 (s, 1 H, H-6), 8.21-8.23 (m, 2 H, H-dibenzofuran), 8.75 (s, 1 H,
- 790 H-2). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 21.2 (CH<sub>3</sub>), 61.5 (C-5'), 70.6 (C-3'), 74.0 (C-2'), 85.3 (C-4'),
- 791 86.8 (C-1'), 110.5 (C-5), 111.8 (C-dibenzofuran), 116.8 (C-4a), 119.0 (C-dibenzofuran), 120.6 (C-
- 792 dibenzofuran), 121.4 (C-dibenzofuran), 123.3 (3 C, C-dibenzofuran), 123.7 (C-dibenzofuran), 125.9 (C-6),
- 793 127.8 (C-dibenzofuran), 129.2 (C-dibenzofuran), 150.6 (C-7a), 151.0 (C-2), 153.7 (C-dibenzofuran), 155.4
- 794 (C-dibenzofuran), 159.4 (C-4). HRMS (ESI) calculated for  $C_{24}H_{22}N_3O_5^+$  ([M+H<sup>+</sup>]): 432.1554, found:
- 795 432.1543.
- 796 **4.2.39.** 4-Methyl-5-((*E*)-2-(pyridin-2-yl)vinyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-*d*]pyrimidine (52).
- 797 The iodo-precursor **55** (150 mg, 0.38 mmol), Pd(OAc)<sub>2</sub> (9 mg, 0.038 mmol, 0.1 eq.) and TPPTS (43 mg,
- 798 0.076 mmol, 0.2 eq.) were added to a 10 mL round-bottom flask, exchanged with a gas atmosphere with

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799
                            argon. Next, TEA (106 μL, 0.76 mmol, 2 eq.) and DMF (2 mL, 5 mL/mmol SM) were added in the reaction
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                            mixture via syringes. The reaction mixture was stirred at ambient temperature for 5 min, and then stirred at
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                             100 °C. When the completion of the reaction was observed via LC-MS analysis, the reaction was cooled to
                            room temperature. The reaction mixture was evaporated in vacuo and purified by column chromatography
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803
                            (0 \rightarrow 10\% \text{ MeOH/DCM}) to give 52 (55 mg, 0.15 mmol) as a pink solid in 39% yield. <sup>1</sup>H NMR (400 MHz,
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                            DMSO-d_6) \delta: 2.87 (s, 3 H, CH<sub>3</sub>), 3.53 - 3.63 (m, 1 H, H-5"), 3.64 - 3.78 (m, 1 H, H-5"), 3.94 (q, J = 3.8
805
                            Hz, 1 H, H-4'), 4.14 - 4.17 (m, 1 H, H-3'), 4.47 (q, J = 6.0 Hz, 1 H, H-2'), 5.11 (t, J = 5.6 Hz, 1 H, OH-5'),
806
                            5.19 \text{ (d, } J = 5.0 \text{ Hz, } 1 \text{ H, } \text{OH-3'}), 5.41 \text{ (d, } J = 6.3 \text{ Hz, } 1 \text{ H, } \text{OH-2'}), 6.24 \text{ (d, } J = 6.1 \text{ Hz, } 1 \text{ H, } \text{H-1'}), 7.18 \text{ (d, } J = 6.0 \text{ Hz, } 1 \text{ H, 
                            J = 16.0 \text{ Hz}, 1 \text{ H}, \text{HC} = \text{CH}, 7.22 - 7.26 \text{ (m, 1 H, H-Pyr)}, 7.52 \text{ (d, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 \text{ H}, \text{H-Pyr)}, 7.78 \text{ (td, } J = 7.9 \text{ Hz}, 1 
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808
                            7.7, 1.8 Hz, 1 H, H-Pyr), 7.98 (d, J = 16.0 Hz, 1 H, HC=CH), 8.31 (s, 1 H, H-6), 8.53 - 8.61 (m, 1 H, H-
809
                            Pyr), 8.67 (s, 1 H, H-2). ^{13}C NMR (100 MHz, DMSO-d_6) \delta: 23.1 (CH<sub>3</sub>), 61.5 (C-5'), 70.5 (C-3'), 74.0 (C-
810
                            2'), 85.2 (C-4'), 86.5 (C-1'), 113.6 (C-5), 115.8 (C-4a), 121.8 (C-Pyr), 122.0 (C-Pyr), 123.5 (C=C), 123.7
                            (C-6), 127.6 (C=C), 136.8 (C-Pyr), 149.5 (C-Pyr), 150.8 (C-7a), 151.0 (C-2), 155.1 (C-Pyr), 159.4 (C-4).
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                            HRMS (ESI): calculated for C_{19}H_{21}N_4O_4 ([M+H]<sup>+</sup>): 369.1557, found: 369.1571.
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                                                           4-Methyl-5-(2-(pyridin-2-yl)ethyl)-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (53).
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                            Compound 52 (50 mg, 0.14 mmol) was dissolved in MeOH (2 mL). The flask was purged into a N<sub>2</sub>
                            atmosphere. Next, a cat. amount of Pd(OH)<sub>2</sub>/C was added into the reaction solution. Then, the N<sub>2</sub> gas was
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816
                            exchanged with H<sub>2</sub> gas (balloon; bubbling). The reaction mixture was stirred at ambient temperature until
                            LC-MS analysis showed full conversion of 52. Then, the balloon was removed and the mixture was filtered
817
                            over Celite and rinsed by MeOH. The filtrate was evaporated until dryness. The residue was purified by
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819
                            column chromatography (0 \rightarrow 10% MeOH/DCM) to give 53 (32 mg, 0.086 mmol) as a white solid in 64%
820
                            yield. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ: 2.78 (s, 3 H, CH<sub>3</sub>), 3.04 - 3.15 (m, 2 H, CH<sub>2</sub>), 3.19 - 3.27 (m, 2 H,
                            CH_2), 3.47 - 3.57 (m, 1 H, H-5''), 3.58 - 3.67 (m, 1 H, H-5'), 3.89 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 Hz, 1 H, H-4'), 4.08 (q, J = 3.6 
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                            4.5 \text{ Hz}, 1 H, H-3'), 4.35 \text{ (q, } J = 6.0 \text{ Hz}, 1 H, H-2'), 5.05 \text{ (t, } J = 5.4 \text{ Hz}, 1 H, OH-5'), 5.14 \text{ (d, } J = 4.9 \text{ Hz}, 1
823
                            H, OH-3'), 5.29 (d, J = 6.4 Hz, 1 H, OH-2'), 6.16 (d, J = 6.1 Hz, 1 H, H-1'), 7.23 (ddd, J = 7.4, 4.9, 1.1 Hz,
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        1 H, H-Pyr), 7.33 (d, J = 7.8 Hz, 1 H, H-Pyr), 7.55 (s, 1 H, H-6), 7.71 (td, J = 7.6, 1.9 Hz, 1 H, H-Pyr), 8.48
        -8.55 (m, 1 H, H-Pyr), 8.59 (s, 1 H, H-2). {}^{13}C NMR (100 MHz, DMSO-d_6) \delta: 22.3 (CH<sub>3</sub>), 26.3 (CH<sub>2</sub>), 38.5
825
        (CH<sub>2</sub>), 61.7 (C-5'), 70.6 (C-3'), 73.8 (C-2'), 84.9 (C-4'), 86.4 (C-1'), 114.9 (C-5), 116.8 (C-4a), 121.4 (C-1')
826
        Pyr), 122.9 (C-Pyr), 123.3 (C-6), 136.5 (C-Pyr), 149.1 (C-Pyr), 150.5 (C-2), 150.7 (C-7a), 158.9 (C-4),
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828
        160.7 (C-Pyr). HRMS (ESI): calculated for C_{19}H_{23}N_4O_4 ([M+H]<sup>+</sup>): 371.1714, found: 371.1731.
        4.2.41. 4-Methyl-5-phenylthio-N7-(β-D-ribofuranosyl)-pyrrolo[2,3-d]pyrimidine (54[39]). The iodo-
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        precursor 55 (150 mg, 0.38 mmol), thiophenol (42 mg, 0.38 mmol, 1 eq.), CuI (4 mg, 0.019 mmol, 0.05
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        eq.) and K<sub>2</sub>CO<sub>3</sub> (131 mg, 0.95 mmol, 2.5 eq.) were added to a 10 mL round-bottom flask purged with argon.
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        Ethyleneglyol (106 μL, 1.90 mmol, 5 eq.) and iPrOH (2 mL, 5 mL/mmol SM) were added and the reaction
        mixture was stirred at ambient temperature for 5 min, and then at 130 °C. Upon reaction completion (LC-
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834
        MS), the reaction mixture was cooled to room temperature and evaporated in vacuo. The residue was
        purified by column chromatography (0 \rightarrow 10% MeOH/DCM) to give 54 (130 mg, 0.35 mmol) as a white
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        solid in 92% yield. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ: 2.61 (s, 3 H, CH<sub>3</sub>), 3.55 - 3.60 (m, 1 H, H-5"), 3.65
        -3.703 (m, 1 H, H-5'), 3.96 (q, J = 3.4 Hz, 1 H, H-4'), 4.14 (q, J = 4.5 Hz, 1 H, H-3'), 4.46 (q, J = 5.7 Hz,
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        1 H, H-2'), 5.11 (t, J = 5.3 Hz, 1 H, OH-5'), 5.19 (d, J = 4.9 Hz, 1 H, OH-3'), 5.44 (d, J = 6.3 Hz, 1 H, OH-
        2'), 6.25 (d, J = 5.9 Hz, 1 H, H-1'), 7.05 - 7.80 (m, 2 H, H-Ph), 7.13 - 7.17 (m, 1 H, H-Ph), 7.26 - 7.30 (m,
839
        2 H, H-Ph), 8.27 (s, 1 H, H-6), 8.73 (s, 1 H, H-2). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 20.6 (CH<sub>3</sub>), 61.3 (C-
840
841
        5'), 70.4 (C-3'), 74.3 (C-2'), 85.3 (C-4'), 86.9 (C-1'), 100.4 (C-5), 117.6 (C-4a), 125.4 (2 C, C-Ph), 125.5
        (C-Ph), 129.3 (2 C, C-Ph), 133.9 (C-6), 138.6 (C-Ph), 151.4 (C-7a), 151.6 (C-2), 159.7 (C-4). HRMS (ESI):
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4.3. *In vitro* evaluation

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calculated for  $C_{18}H_{20}N_3O_4S$  ([M+H]<sup>+</sup>): 374.1169, found: 374.1158.

**4.3.1. Test Substances and Formulations** Test compounds were formulated in 100% dimethyl sulfoxide (DMSO) at 20 mM. The compounds were 4-fold serially diluted to obtain ten concentrations and an in-test concentration starting at 64 μM.

Intracellular amastigote assay of L. infantum (MHOM/MA(BE)/67/ITMAP263, WT strain): amastigotes (spleen-derived from an infected donor hamster) were maintained on primary peritoneal mouse macrophages (M $\phi$ , from Swiss mice) in RPMI-1640 medium, supplemented with 200 mM L-glutamine, 16.5 mM NaHCO<sub>3</sub> and 5% inactivated fetal calf serum. All cultures and assays were conducted at 37 °C and 5% CO<sub>2</sub> atmosphere. Assays were performed with 190  $\mu$ L of M $\phi$ /parasite inoculum (3 × 10<sup>4</sup> cells + 4.5 × 10<sup>5</sup> parasites/well). The M $\phi$  were infected for 48 h, and 10  $\mu$ L of compound dilutions were added after 2 h of infection. Parasite growth was compared to untreated-infected controls (set as 100% growth) and uninfected controls (set as 0% growth). The incubation lasted for 5 days, and parasite burdens (mean number of amastigotes/M $\phi$ ) were microscopically measured after staining with a 10% Giemsa solution. The results were given as percentage (%) reduction in growth compared to controls and was used to calculate the IC<sub>50</sub> value. MIL was included as the positive drug control.

- **4.3.2 Cytotoxicity** towards  $M\phi$  was microscopically defined as the lowest concentration causing cell detachment, lysis and granulation. This was done by semi-quantitative scoring of at least 500 cells distributed over adjacent microscopic areas. The results were given as the percentage (%) reduction in cell viability compared to untreated controls. The percentage was used to calculate  $CC_{50}$  value.
- **4.3.3 Drug-resistant strains of** *L. infantum* Compounds were assayed on promastigotes of *L. infantum* MHOM/FR/96/LEM3323-Cl4 (Sb-resistant) and derived isogenic strains with additional resistance against MIL (LEM3323-Cl4 MIL5Cl3) or PMM (LEM3323-Cl4 PMM). Promastigotes were cultured at room temperature in HOMEM promastigote medium (Gibco®, Life technologies) supplemented with 10% FCS. Mφ were infected by metacyclic promastigotes at 2:1 multiplicity of infection (MOI). Infected Mφ were incubated at 37 °C and 5% CO<sub>2</sub> atmosphere. After 24 h incubation, 10 μL of compound dilutions were added. After 96 h incubation, parasite burdens (mean number of amastigotes/Mφ) were assessed

872 controls and used to calculate the EC<sub>50</sub> values. 4.3.4 Intracellular amastigote assay of *T. cruzi* (Tulahuen CL2 β-galactosidase reporter strain) 873 874 Compounds were evaluated in vitro as described in previous report, as was the in vitro cytotoxicity assay 875 against MRC-5 fibroblasts.[40] 876 In vitro microsomal stability assay. Liver microsomes of mouse, hamster and human (pooled) were supplied 877 commercially from Corning® and stored at -80 °C. NADPH generating system solutions A and B, and UGT reaction mix solutions A and B stored at -20 °C. Testing compounds and reference diclofenac were 878 879 formulated in 100% DMSO at 10 mM. The in vitro microsomal stability assays were performed exactly as 880 described previously. [40] The results were given as the percentage of remaining parent compound. The 881 percentages from two independent assays were used to calculate mean and STDEV. 4.4. In vivo evaluation 882 **4.4.1. Test substances and formulations 32** was formulated in 10% (v/v) PEG 400 and 1% Tween 80 (v/v) 883 in water at 25 mg/mL envisaging a dosing volume of 200  $\mu$ L/100 g. The formulation at 25 mg/mL resulted 884 885 in a thick fine white suspension. MIL was formulated at 20 mg/mL in water. **4.4.2.** Efficacy in the early curative hamster model Female golden hamsters (BW 75-85 g) were 886 purchased from Janvier-France and kept in quarantine for at least 5 days before starting the experiment. 887 888 Food for laboratory rodents and drinking water were available ad libitum. The animals were randomly 889 allocated to experimental units of 5 animals based on live body weight at the start of the experiment (Day 0 = day of infection). L. infantum (MHOM/MA(BE)/67) amastigates were obtained from the spleens of 890 heavily infected donor hamsters were purified using two centrifugation steps and diluted to prepare an 891 infection inoculum containing  $2 \times 10^7$  amastigotes/100 µL phosphate buffered saline (PBS). The infection 892

microscopically. The results were given as the percentage of reduction in parasite growth compared to

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- inoculum was administered intracardially (IC). Animals were treated orally s.i.d. for 10 days. Dosing started at 21 dpi.
- G1: Vehicle-treated infected control (VIC): 10% PEG 400 and 1% Tween 80 in water s.i.d. PO for 10
- 896 days (200  $\mu$ L) [n = 5]
- **G2**: MIL: 40 mg/kg s.i.d. PO for 5 days (200  $\mu$ L) [n = 5]
- **G3: 32**: 50 mg/kg s.i.d. PO for 10 days (200 μL/100g) [n = 5]
- 899 Amastigote burdens in the different target organs (liver, spleen, bone-marrow) were determined 9 days after 900 the last treatment (i.e. day 39 of the experiment). The organs of individual animals were weighed (except 901 bone-marrow); impression smears were stained with Giemsa for microscopic evaluation of the total 902 amastigote burden (= mean number of amastigotes per cell × number of cells counted (minimum 500 nuclei). Percentage reduction compared to the burdens in the vehicle-treated infected control animals (VIC) is used 903 904 as a measure for drug activity. Possible 'viable' residual burdens were assessed using the promastigote 905 back-transformation assay (incubation of small pieces of tissue in promastigote medium at room 906 temperature with qualitative assessment of the presence of promastigotes after 2 weeks of incubation).
- 4.4.3. *In vivo* toxicity The animals were observed daily for the occurrence of clinical or adverse effects. Allanimals were weighed twice weekly to monitor the general health status.
- 4.4.4. Pharmacokinetics For G3 and G4, a series of blood samples were collected from the sublingual vein on the first and the last day of treatment. To reduce the number of samplings per individual animal and the associated the stress involved in the repeated blood sampling, subgroups of 3 and 2 animals were sampled on alternate time points. Samples were collected at 0.5, 1, 2, 4 and 8 h (before second dose) and at 24 h (before the morning dose of day 2 = 16 h post last treatment dose). On day 9, samples were taken pre-dose (0) and at 0.5, 1, 2, 4 and 8 h, and 24 h (16 h post last treatment dose). All samples were collected as DBS for analytical detection of parent compound using LC-MS/MS.
- Bio-analysis used LC-MS/MS (UPLC) (Waters Aquity<sup>TM</sup>) coupled with tandem quadrupole mass spectrometry (MS<sup>2</sup>) (Waters Xevo<sup>TM</sup>), equipped with an electrospray ionization (ESI) interface and

operated in multiple reaction monitoring (MRM) mode. The optimal MS parameters and control of the chromatographic separation conditions had been tuned before (**Table 1 in Supplementary material**). For calibration and validation, standard curves in blood were made and spotted onto DBS cards. The standard PK parameters were determined using appropriate software (TopFit):  $C_{max}$ ,  $T_{max}$ ,  $AUC_{0-24h}$ , Cl and Vd. The calculated parameters are based upon the mean values per time point using non-compartmental analysis. The areas under the curve ( $AUC_{0-8h}$  and  $AUC_{0-24h}$ ) were calculated using the linear trapezoidal rule.

#### **Ethics statement**

The use of mice for the isolation of peritoneal macrophages and hamsters for splenic *Leishmania infantum* amastigotes was approved by the ethical committee of the University of Antwerp [UA-ECD 2019-10]. The use of laboratory rodents was carried out in strict accordance to all mandatory guidelines (EU directives, including the Revised Directive 2010/63/EU on the Protection of Animals used for Scientific Purposes that came into force on 01/01/2013, and the declaration of Helsinki in its latest version).

### Supplementary material

Copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra of synthesized compounds and UPLC-MS/MS conditions for the determination **32** and microphotographs of Giemsa-stained tissue imprints of various organs (liver, spleen and bone marrow) of vehicle-, MIL - and **32**-treated animals and mean concentration (ng/mL) of **32** in blood after a single dose (Day 1: 50 mg/kg) and after repeated-dose administration (Day 9: 50 mg/kg b.i.d. for 10 days) in infected hamsters.

#### **Conflicts of interest**

The authors declare no conflict of interest.

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#### REFERENCE

- 952 [1] S. Burza, S.L. Croft, M. Boelaert, Leishmaniasis, Lancet, 392 (2018) 951-970.
- 953 [2] World Health Organization, Leishmaniasis, updated May 2021. https://www.who.int/en/news-
- 954 room/fact-sheets/detail/leishmaniasis.
- 955 [3] D. Pace, Leishmaniasis, J Infect, 69 Suppl 1 (2014) S10-18.
- 956 [4] M. Akhoundi, K. Kuhls, A. Cannet, J. Votypka, P. Marty, P. Delaunay, D. Sereno, A Historical Overview
- of the Classification, Evolution, and Dispersion of Leishmania Parasites and Sandflies, PLoS Negl Trop Dis,
- 958 10 (2016) e0004349.
- 959 [5] D. Sacks, A. Sher, Evasion of innate immunity by parasitic protozoa, Nat Immunol, 3 (2002) 1041-1047.
- 960 [6] F. Chappuis, S. Sundar, A. Hailu, H. Ghalib, S. Rijal, R.W. Peeling, J. Alvar, M. Boelaert, Visceral
- leishmaniasis: what are the needs for diagnosis, treatment and control?, Nat Rev Microbiol, 5 (2007) 873-
- 962 882
- 963 [7] P.M. Gillespie, C.M. Beaumier, U. Strych, T. Hayward, P.J. Hotez, M.E. Bottazzi, Status of vaccine
- research and development of vaccines for leishmaniasis, Vaccine, 34 (2016) 2992-2995.
- 965 [8] F.J. Perez-Victoria, M.P. Sanchez-Canete, K. Seifert, S.L. Croft, S. Sundar, S. Castanys, F. Gamarro,
- Mechanisms of experimental resistance of Leishmania to miltefosine: Implications for clinical use, Drug
- 967 Resist Updat, 9 (2006) 26-39.
- 968 [9] D.L. Looker, J.J. Marr, R.L. Berens, Mechanisms of action of pyrazolopyrimidines in Leishmania
- 969 donovani, J Biol Chem, 261 (1986) 9412-9415.
- 970 [10] G.D. Campagnaro, H.P. de Koning, Purine and pyrimidine transporters of pathogenic protozoa -
- onduits for therapeutic agents, Med Res Rev, 40 (2020) 1679-1714.
- 972 [11] H.P. de Koning, D.J. Bridges, R.J. Burchmore, Purine and pyrimidine transport in pathogenic protozoa:
- 973 from biology to therapy, FEMS Microbiol Rev, 29 (2005) 987-1020.
- 974 [12] J.M. Boitz, B. Ullman, A conditional mutant deficient in hypoxanthine-guanine
- 975 phosphoribosyltransferase and xanthine phosphoribosyltransferase validates the purine salvage pathway
- 976 of Leishmania donovani, J Biol Chem, 281 (2006) 16084-16089.

- 977 [13] M. Berg, P. Van der Veken, A. Goeminne, A. Haemers, K. Augustyns, Inhibitors of the Purine Salvage
- 978 Pathway: A Valuable Approach for Antiprotozoal Chemotherapy?, Curr Med Chem, 17 (2010) 2456-2481.
- 979 [14] M. Gottlieb, D.M. Dwyer, Leishmania donovani: surface membrane acid phosphatase activity of promastigotes, Exp Parasitol, 52 (1981) 117-128.
- 981 [15] J.J. Marr, R.L. Berens, D.J. Nelson, Purine metabolism in Leishmania donovani and Leishmania 982 braziliensis, Biochim Biophys Acta, 544 (1978) 360-371.
- 983 [16] F. Hulpia, G.D. Campagnaro, M. Scortichini, K. Van Hecke, L. Maes, H.P. de Koning, G. Caljon, S. Van
- Calenbergh, Revisiting tubercidin against kinetoplastid parasites: Aromatic substitutions at position 7
- improve activity and reduce toxicity, Eur J Med Chem, 164 (2019) 689-705.
- 986 [17] F. Hulpia, D. Mabille, G.D. Campagnaro, G. Schumann, L. Maes, I. Roditi, A. Hofer, H.P. de Koning, G.
- 987 Caljon, S. Van Calenbergh, Combining tubercidin and cordycepin scaffolds results in highly active
- candidates to treat late-stage sleeping sickness, Nat Commun, 10 (2019) 5564.
- 989 [18] J. Bouton, A. Furquim d'Almeida, L. Maes, G. Caljon, S. Van Calenbergh, F. Hulpia, Synthesis and
- evaluation of 3'-fluorinated 7-deazapurine nucleosides as antikinetoplastid agents, Eur J Med Chem, 216 (2021) 113290.
- 992 [19] F. Hulpia, J. Bouton, G.D. Campagnaro, I.A. Alfayez, D. Mabille, L. Maes, H.P. de Koning, G. Caljon, S.
- 993 Van Calenbergh, C6-O-alkylated 7-deazainosine nucleoside analogues: Discovery of potent and selective
- anti-sleeping sickness agents, Eur J Med Chem, 188 (2020) 112018.
- 995 [20] C. Lin, L. Ferreira de Almeida Fiuza, C. Cardoso Santos, D. Ferreira Nunes, O. Cruz Moreira, J. Bouton,
- 996 I. Karalic, L. Maes, G. Caljon, F. Hulpia, C.S.M. de Nazare, S. Van Calenbergh, 6-Methyl-7-Aryl-7-
- Deazapurine Nucleosides as Anti-Trypanosoma cruzi Agents: Structure-Activity Relationship and in vivo Efficacy, ChemMedChem, 16 (2021) 2231-2253.
- 999 [21] C. Lin, F. Hulpia, C.F. da Silva, D. Batista, K. Van Hecke, L. Maes, G. Caljon, M.N.C. Soeiro, S. Van
- 1000 Calenbergh, Discovery of Pyrrolo[2,3-b]pyridine (1,7-Dideazapurine) Nucleoside Analogues as Anti-1001 Trypanosoma cruzi Agents, J Med Chem, 62 (2019) 8847-8865.
- 1002 [22] J. Bouton, L. Ferreira de Almeida Fiuza, C. Cardoso Santos, M.A. Mazzarella, M.N.C. Soeiro, L. Maes, I.
- 1003 Karalic, G. Caljon, S. Van Calenbergh, Revisiting Pyrazolo[3,4-d]pyrimidine Nucleosides as Anti-
- 1004 Trypanosoma cruzi and Antileishmanial Agents, J Med Chem, 64 (2021) 4206-4238.
- 1005 [23] J. Bouton, L. Maes, I. Karalic, G. Caljon, S. Van Calenbergh, Synthesis and evaluation of a collection of
- 1006 purine-like C-nucleosides as antikinetoplastid agents, Eur J Med Chem, 212 (2021) 113101.
- 1007 [24] P. Perlikova, A. Krajczyk, E. Dolezelova, M. Slapnickova, N. Milisavljevic, L.P. Slavetinska, E. Tloust'ova,
- 1008 S. Gurska, P. Dzubak, M. Hajduch, A. Zikova, M. Hocek, Synthesis and Antitrypanosomal Activity of 6-
- 1009 Substituted 7-Methyl-7-deazapurine Nucleosides, ACS Infect Dis, 7 (2021) 917-926.
- 1010 [25] P. Perlikova, G. Rylova, P. Naus, T. Elbert, E. Tloustova, A. Bourderioux, L.P. Slavetinska, K. Motyka, D.
- 1011 Dolezal, P. Znojek, A. Nova, M. Harvanova, P. Dzubak, M. Siller, J. Hlavac, M. Hajduch, M. Hocek, 7-(2-
- Thienyl)-7-Deazaadenosine (AB61), a New Potent Nucleoside Cytostatic with a Complex Mode of Action,
- 1013 Mol Cancer Ther, 15 (2016) 922-937.
- 1014 [26] C. Lin, F. Hulpia, I. Karalic, L. De Schepper, L. Maes, G. Caljon, S. Van Calenbergh, 6-Methyl-7-
- deazapurine nucleoside analogues as broad-spectrum antikinetoplastid agents, Int J Parasitol Drugs Drug
- 1016 Resist, 17 (2021) 57-66.
- 1017 [27] P. Naus, O. Caletkova, P. Konecny, P. Dzubak, K. Bogdanova, M. Kolar, J. Vrbkova, L. Slavetinska, E.
- 1018 Tloust'ova, P. Perlikova, M. Hajduch, M. Hocek, Synthesis, cytostatic, antimicrobial, and anti-HCV activity
- of 6-substituted 7-(het)aryl-7-deazapurine ribonucleosides, J Med Chem, 57 (2014) 1097-1110.
- 1020 [28] A. Bourderioux, P. Naus, P. Perlikova, R. Pohl, I. Pichova, I. Votruba, P. Dzubak, P. Konecny, M. Hajduch,
- 1021 K.M. Stray, T. Wang, A.S. Ray, J.Y. Feng, G. Birkus, T. Cihlar, M. Hocek, Synthesis and significant cytostatic
- activity of 7-hetaryl-7-deazaadenosines, J Med Chem, 54 (2011) 5498-5507.
- 1023 [29] N. Kudo, M. Perseghini, G.C. Fu, A versatile method for Suzuki cross-coupling reactions of nitrogen
- 1024 heterocycles, Angew Chem Int Ed Engl, 45 (2006) 1282-1284.

- 1025 [30] F.U. Rutaganira, J. Barks, M.S. Dhason, Q. Wang, M.S. Lopez, S. Long, J.B. Radke, N.G. Jones, A.R.
- 1026 Maddirala, J.W. Janetka, M. El Bakkouri, R. Hui, K.M. Shokat, L.D. Sibley, Inhibition of Calcium Dependent
- 1027 Protein Kinase 1 (CDPK1) by Pyrazolopyrimidine Analogs Decreases Establishment and Reoccurrence of
- 1028 Central Nervous System Disease by Toxoplasma gondii, J Med Chem, 60 (2017) 9976-9989.
- 1029 [31] F.Y. Kwong, S.L. Buchwald, A general, efficient, and inexpensive catalyst system for the coupling of
- 1030 aryl iodides and thiols, Org Lett, 4 (2002) 3517-3520.
- 1031 [32] X.A.F. Cook, A. de Gombert, J. McKnight, L.R.E. Pantaine, M.C. Willis, The 2-Pyridyl Problem:
- 1032 Challenging Nucleophiles in Cross-Coupling Arylations, Angew Chem Int Ed Engl, 60 (2021) 11068-11091.
- 1033 [33] M. Van den Kerkhof, D. Mabille, E. Chatelain, C.E. Mowbray, S. Braillard, S. Hendrickx, L. Maes, G.
- 1034 Caljon, In vitro and in vivo pharmacodynamics of three novel antileishmanial lead series, Int J Parasitol
- 1035 Drugs Drug Resist, 8 (2018) 81-86.
- 1036 [34] H. Lu, C. Chen, C. Klaassen, Tissue distribution of concentrative and equilibrative nucleoside
- transporters in male and female rats and mice, Drug Metab Dispos, 32 (2004) 1455-1461.
- 1038 [35] L.P. Jordheim, D. Durantel, F. Zoulim, C. Dumontet, Advances in the development of nucleoside and
- nucleotide analogues for cancer and viral diseases, Nat Rev Drug Discov, 12 (2013) 447-464.
- 1040 [36] M. De Rycker, B. Baragana, S.L. Duce, I.H. Gilbert, Challenges and recent progress in drug discovery
- 1041 for tropical diseases, Nature, 559 (2018) 498-506.
- 1042 [37] M.C. Field, D. Horn, A.H. Fairlamb, M.A. Ferguson, D.W. Gray, K.D. Read, M. De Rycker, L.S. Torrie,
- 1043 P.G. Wyatt, S. Wyllie, I.H. Gilbert, Anti-trypanosomatid drug discovery: an ongoing challenge and a
- 1044 continuing need, Nat Rev Microbiol, 15 (2017) 217-231.
- 1045 [38] M.L. Sykes, V.M. Avery, Approaches to protozoan drug discovery: phenotypic screening, J Med Chem,
- 1046 56 (2013) 7727-7740.
- 1047 [39] M. Klecka, L.P. Slavetinska, E. Tloustova, P. Dzubak, M. Hajduch, M. Hocek, Synthesis and cytostatic
- activity of 7-arylsulfanyl-7-deazapurine bases and ribonucleosides, Medchemcomm, 6 (2015) 576-580.
- 1049 [40] F. Hulpia, K. Van Hecke, C. Franca da Silva, D. da Gama Jaen Batista, L. Maes, G. Caljon, C.S.M. de
- 1050 Nazare, S. Van Calenbergh, Discovery of Novel 7-Aryl 7-Deazapurine 3'-Deoxy-ribofuranosyl Nucleosides
- with Potent Activity against Trypanosoma cruzi, J Med Chem, 61 (2018) 9287-9300.