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Authors: Van Renterghem J., Dhondt H., Verstraete G., De Bruyne M., Vervaet C., De Beer T.

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The impact of the injection mold temperature upon polymer crystallization and resulting drug release from immediate and sustained release tablets.

Jeroen Van Renterghem¹, Heleen Dhondt¹, Glenn Verstraete², Michiel De Bruyne³, Chris Vervaet², Thomas De Beer¹

¹Laboratory of pharmaceutical process analytical technology, Ottergemsesteenweg 460, 9000, Ghent, Belgium
²Laboratory of pharmaceutical technology, Ottergemsesteenweg 460, 9000, Ghent, Belgium
³Inflammation Research Center, VIB, Ghent, Belgium and Department of Biomedical Molecular Biology, Ghent University, 9052 Ghent, Belgium.

³Department of Plant Systems Biology, VIB, Ghent, Belgium and Department of Plant Biotechnology and Bioinformatics, Ghent University, 9052 Gent, Belgium.

*Corresponding author: Jeroen Van Renterghem
Laboratory of Process Analytical Technology, Ghent University, Ottergemsesteenweg 460, 9000 Ghent, Belgium
TEL: 0032 9 264 8039
FAX: 0032 9 264
E-MAIL: jeroen.vanrenterghem@ugent.be
Abstract

It was the aim of this study to elucidate the impact of the injection mold temperature upon the polymer crystallinity, its microstructure and the resulting drug release from immediate and sustained release tablets containing semi-crystalline polymers. The immediate release formulation contained 20% (w/w) ketoprofen (KETO) in poly (ethylene oxide) (PEO) and the sustained release formulation contained 20 - 40% (w/w) metoprolol tartrate (MPT) in polycaprolactone (PCL). Physical mixtures of drug-polymer were characterized via isothermal crystallization experiments using DSC and rheological measurements to elucidate the impact of the drug solid-state upon the crystallization kinetics. Tablets were prepared using various thermal histories (extrusion barrel temperature and injection mold temperatures). Polymer crystallinity and microstructure in the tablets was characterized via DSC and polarized optical microscopy. The polymer microstructure was altered by the various applied thermal histories. The differences in PEO crystallinity induced by the various mold temperatures did not affect the KETO dissolution from the tablets. On the other hand, MPT (20 - 40% w/w) dissolution from the PCL matrix when extruded at 80 °C and injection molded at 25 and 35 °C was significantly different due to the changes in the polymer microstructure. More perfect polymer crystals are obtained with higher mold temperatures, decreasing the drug diffusion rate through the PCL matrix. The results presented in this study imply that the injection mold temperature should be carefully controlled for sustained release formulations containing hydrophobic semi-crystalline polymers.

Keywords: Solid dispersion, injection molding, semi-crystalline polymers, rheology, crystallization

Abbreviations
1 Introduction
The well-established melt processing technique hot-melt extrusion (HME) can be combined with injection molding (IM) for downstream processing. Herewith, the extrusion step is conducted to melt the raw materials (i.e., drug + polymer) using elevated barrel temperatures and shearing forces from the rotating screw(s), to obtain a homogeneous melt at the end of the die. In a continuous manner, subsequent injection of the melt into a mold cavity yields the final dosage form. In combination, a lot of opportunities for the pharmaceutical industry exist towards production of immediate and sustained release solid dosage forms, depending on the type of polymer used. Another strength of the HME/IM combination is the large variety of drug products that can be produced: matrix tablets (Bruce et al., 2005), implants (Rothen-Weinhold et al., 1999), transdermal drug delivery systems (Crowley et al., 2004; Prodduturi et al., 2005), vaginal rings (Clark et al., 2012).

When using HME in combination with IM, characterization of the drug-polymer melt formulation by rheological measurements is essential to predict the processability. An indicative range of the processing temperature can be established by a rheological characterization of the flow properties of the molten dispersions (Gupta et al., 2012; Parikh et al., 2014; Van Renterghem et al., 2017; Verstraete et al., 2016). When using semi-crystalline polymers as carrier during HME and IM, these polymers are processed above their melting temperature to decrease their melt viscosity sufficiently to allow a steady flow of the polymer through the extruder barrel and into the mold. Upon cooling from the melt, semi-crystalline polymers solidify when nuclei are formed and crystallization commences. Depending on the cooling rate (i.e., mold temperature) and the solid-state of the drug content, the crystallization rate of the polymer is altered as well as the polymer’s crystalline microstructure and crystallinity percentage. Jeong et al. has shown that a small difference in the polymer morphology can influence the drug release from microspheres prepared via solvent evaporation (Jeong et al., 2003). Annealing the PCL microspheres at 25, 40 and 50 °C yielded differences in the drug release. As the size of the lamellae was larger for the annealed sample at 50 °C compared to the 25 °C sample, the drug diffusion was affected. Other researchers also demonstrated that the polymer crystallinity had a significant impact on the drug release from other semi-crystalline polymer systems prepared via solvent
evaporation (Alexis, 2005; Karavelidis et al., 2011; Zilberman, 2005). However, to the best of our knowledge, this thermal history effect upon drug release was not investigated for injection molded matrix systems.

It was the aim of this study to investigate the influence of various processing conditions (i.e., extrusion barrel temperature and injection mold temperature) upon the drug release from both immediate and sustained release drug formulations prepared by HME and IM. Physical mixtures of drug and semi-crystalline polymer were first characterized by DSC and rheological experiments to obtain knowledge about their processability and the influence of the drug solid-state upon polymer crystallization. Injection-moulded tablets were prepared using various extrusion barrel temperatures and injection mold temperatures. These tablets were analyzed using DSC to determine the polymer crystallinity and the drug solid-state. Polarized optical microscopy (POM) was used to investigate the polymer microstructure of sustained release tablets containing PCL as matrix. Dissolution tests were performed to investigate the impact of mold temperature (i.e. thermal history) on the resulting drug release.
2 Experimental

2.1 Materials

The immediate release formulation contained 20% (w/w) ketoprofen (KETO) in poly (ethylene oxide) (PEO). KETO (SIMS, Florence, Italy) is a non-steroidal anti-inflammatory drug (NSAID) with poor water solubility (BCS class-II), ideal as model drug for the preparation of immediate release formulations. KETO has a melting point of 94 °C (Sweetman, 2009). PEO (POLYOX™ WSR N10 LEO NF, Dow Chemical Company, Michigan, USA) is a water soluble semi-crystalline polymer with a glass transition temperature of -67 °C and a melting point of 65 °C. The average molecular weight is 100,000 g/mol (Suwardie et al., 2011). The enthalpy of fusion for 100% crystalline PEO is 205 J/g (Zhao et al., 2005). KETO/PEO interactions can be detected even in non-melt-processed blends at temperatures as low as 43 °C (Schachter et al., 2004). Because of these strong interactions, KETO is dissolved in the PEO matrix during the isothermal crystallization (section 2.2.1) and extrusion/injection molding experiments (section 2.2.2).

The sustained release formulation contained 20 and 40% (w/w) metoprolol tartrate (MPT) in PCL. MPT (Utag, Amsterdam, The Netherlands) is a cardio-selective beta blocker belonging to the BCS Class-I drugs and is thus an interesting model drug to prepare extended release formulations. MPT crystals have a melting point of approximately 123 °C (Sweetman, 2009). PCL (Capa™ 6506, Perstorp, Malmö, Sweden) is a biodegradable, semi-crystalline and hydrophobic polyester that is poorly water soluble and is used to obtain a controlled release of the API (Active pharmaceutical ingredient). The drug release from this polymer is via diffusion and biodegradation of the polymer is slow (Jeong et al., 2003; Kamaly et al., 2016). PCL has a melting point of approximately 60 °C, a glass transition temperature of -60 °C and an average molecular weight of 50,000 g/mol. The heat of fusion for the 100% crystalline polymer is 139.5 J/g (Gupta et al., 2012).
2.2 Methods

2.2.1 Characterization of raw materials and physical mixtures

Differential scanning calorimetry

A DSC Q2000 (TA instruments, New Castle, USA) was used to perform all thermal analysis of the raw materials, the physical mixtures and the IM tablets. All experiments were performed using Tzero pans containing approximately 3 mg of sample. Indium was used to calibrate the instrument. Heat-cool-heat experiments were performed as followed: the sample was first equilibrated for 3 min at -70 °C, followed by a heating run at 10 °C/min to 140 °C which is above the drug melting points of KETO and MPT. Also a heating run to a temperature below the melting point of MPT (i.e., 100°C) was performed to study the crystallization of PCL upon cooling when MPT was still in its crystalline state. The second cycle is a cooling run to -70 °C at 10 °C/min during which crystallization of the polymer can be observed. The last cycle is a second heating at the same heating rate and temperature limits as the first heating. All heat-cool-heat experiments were performed in triplicate.

Furthermore, isothermal crystallization experiments were conducted to study the crystallization speed of the polymer at different isothermal temperatures. The experiments were performed as followed: the physical mixture was first equilibrated for 3 min at a temperature above the polymer melting point (see table 1). The sample was then quickly cooled (at 50 °C/min) to the isothermal crystallization temperature and kept isothermal for one hour during which the sample crystallized. The crystallized sample was again heated at 10 °C/min to determine the polymer melting point. The samples containing MPT were equilibrated below (i.e., 80 °C for 20 and 40% (w/w) MPT) and above (i.e., 140 °C for 20% (w/w) MPT) the drug melting temperature to determine the influence of the drug solid-state upon the polymer crystallization speed.
A Haake™ MARS™ III rheometer (Thermo Fisher, Waltham, USA) equipped with a 20 mm parallel plate configuration was used for all rheological experiments. The lower stationary plate can be heated, allowing to impose different thermal treatments on the sample. Physical mixtures were loaded onto the bottom plate and were allowed sufficient time to soften and equilibrate before commencing the tests. Temperature sweep experiments were performed using a heating/cooling rate of 2 °C/min at a constant frequency (1Hz) and deformation (for PEO, PEO-20KETO, PCL = 2% and for PCL-20MPT =0.01%) that was within the linear viscoelastic range. The sample was heated above the drug melting temperature (i.e., 100 °C or 150 °C for -KETO and MPT formulations, respectively) and was then cooled until crystallization was complete, at which point the sample was again reheated at 2 °C/min to above the drug melting point.

Isothermal crystallization experiments can also be performed using a rheometer. First, the sample is heated until the polymer is molten (PEO systems: 100 °C; PCL systems: 80 °C, 100 °C or 140 °C). Then the sample is cooled down to the isothermal crystallization temperature (Tc) (shown in table 1) with a cooling rate of 5 °C/min followed by a time sweep at Tc. The temperature, the frequency (1 Hz) and the deformation (2%) are kept constant and the complex viscosity, loss and storage moduli are determined as a function of time. Upon crystallization, the complex viscosity and moduli strongly increase. The experiment is stopped once a plateau of the viscoelastic properties (G' and G'') is reached or when the rheometer torque reached its maximum.
Data analysis

The experimental data obtained from the isothermal crystallization experiments was fitted by the commonly used Avrami equation (Eq. 1) (Lorenzo et al., 2007; Piorkowska et al., 2006; Schick, 2009).

\[ 1 - V_c(t) = \exp(-kt^n) \]  

Where \( V_c \) is the relative volumetric transformed fraction, \( n \) the Avrami index, \( k \) (min\(^{-n}\)) the overall crystallization rate constant and \( t \) (min) is time. Crystallization starts with the formation of nuclei, followed by a linear growth of these nuclei. In the Avrami fit, some parameters (i.e. \( n \) and \( k \)) quantifying this nucleation and growth are introduced. For the data measured by DSC, an automated tool developed by Lorenzo et al. was used to calculate the Avrami parameters (Lorenzo et al., 2007).

The Avrami fit was performed manually for the data (i.e., \( G' \) as function of time) obtained by rheological measurements. For both DSC and rheological measurements, a linear range (representing 20 - 40% transformed fraction, with \( R^2 > 0.99 \)) was selected from the linearized Avrami equation (Eq. 2) to calculate the Avrami parameters (i.e., \( n \) and \( k \)). In this region, \( n \) and \( k \) are calculated by the respective slope and intercept.

\[
\log \left( \ln \frac{1}{1-V_c(t)} \right) = \log(k) + n\log(t)
\]  

The experimental half crystallization time \( (t_{1/2}) \) can also be obtained by fitting the Avrami equation (Eq. 3). It should be noted that the absolute zero time \( (t_0) \) is determined as the time when the isothermal crystallization temperature is reached.

\[ t_{1/2} = \left( \frac{\ln(2)}{k} \right)^{1/n} \]
2.2.2 Production of injection molded tablets

The Haake™ minilab co-rotating twin-screw extruder (Thermo Fisher, Waltham, USA) was used to extrude the physical mixtures which were prepared with a mortar and pestle. The extruder was used in continuous mode (i.e. the recirculation channel was not used). The Haake™ MiniJet Pro Piston (Thermo Fisher, Waltham, USA) injection molding system was used to prepare the IM tablets (approximately 350 mg, h = 5 mm, d = 10 mm). The molten extrudate is directly inserted from the extruder into the injector by holding the injector against the die of the extruder. The injector’s temperature is set to the same temperature as the extrusion barrel temperature to maintain the thermal history and to prevent crystallization of the polymer before injection into the mold. After proper filling of the injector, the molten mixture was injected into the mold with a pressure of 800 bar during 5 s after which the pressure is kept constant for another 5 s at 500 bar to avoid expansion by relaxation of the polymer. Annealing of the tablets is done in the mold at various set temperatures (25 and 35 °C) to apply different thermal histories upon the tablets during solidification. The mold temperatures were selected based on the results of the rheological temperature sweep experiments which are described in section 3.1.2. The tablets were kept in the mold for a certain time to allow complete solidification and were then ejected by opening the mold. The various process parameters used for HME and IM are shown in table 2. The 40% MPT physical mixture was only extruded at 80 °C because higher temperatures (i.e., 140 °C) resulted in a too low viscous melt, which cannot be extrudate or injection molded. The residence time in the extruder was approximately 100 s, defined as the lag time from the feeder to the die and was measured by using a color tracer (cochineal red).
2.2.3 Tablet characterization

**Differential scanning calorimetry**

The IM tablets were analyzed by DSC using the same heating rate and limits as used for the physical mixtures. Two samples were cut with a surgical blade from the edges of the flat-faced radius edged tablets (3 tablets per thermal history setting). The first heating cycle of the IM tablets allows to determine the drug and polymer’s melting temperature (i.e., peak temperature $T_m$), the presence of undissolved API particles and the calculation of the degree of crystallinity using Eq. 4 after processing at various thermal histories (i.e., barrel temperature and mold temperature).

$$X_c = \frac{\Delta H_f}{f \times \Delta H_{f100\%}} \times 100\%$$

(4)

where $X_c$ (%) is the degree of crystallinity, $\Delta H_f$ ($J/g$) is the enthalpy of melting, $f$ is the fraction of drug or polymer and $\Delta H_{f100\%}$ is the enthalpy of melting for a fully crystalline polymer or drug.

**Polarized optical microscopy**

The crystalline microstructure of the sustained release tablets (i.e., PCL and PCL-20MPT extruded at 80 °C) was visualized using a polarized optical microscope (Leica DM 2500P) equipped with a black and white camera (Leica DFC360 FX). Samples were made by cutting the tablets in half (injection direction) with a razor blade, trimmed and mounted on aluminum pin (see Fig. SI1). Semi thin sections of 3µm were obtained by cryosectioning at -40 °C using a Leica UC7 microtome equipped with a Leica EM FC7 cyro chamber. Sectioning was performed in direction of the injection flow, from the center towards to the outer edge of the tablet. To stretch the sample slices, they were placed in a drop of glycerol on a glass slide and were then covered with a coverslip. Sections of PCL-20MPT were
immediately collected on a glass slide in the cryo-chamber. Curled sections were stretched manually with an eyelash. The crystalline microstructure was analyzed semi-quantitatively using ImageJ software (National Institutes of Health, Bethesda, MD) (Supplementary material Fig. SI2).

Drug dissolution tests

Immediately after processing, tablets (n = 5) from each thermal history setting were subjected to in vitro dissolution testing. All formulations were tested using the same phosphate buffer solution (pH = 6.6). A VK 7010 dissolution system with a VK 8000 automatic sampling station (Vankel Industries, Edison, USA) was used. The vessels contained 900 mL of phosphate buffer solution (pH 6.6). The paddle speed and the water bath temperature were kept constant at 100 rpm and 37 ± 0.5 °C, respectively. Samples of 5 mL were withdrawn from the dissolution medium at predetermined time points (5, 10, 15, 30, 45, 60, 75, 90, 120 and 240 min for immediate release and 0.5, 1, 2, 4, 6, 8, 12, 16, 20 and 24 hours for sustained release formulations) and spectrophotometrically (Shimadzu 1650PC, Kyoto, Japan) analyzed at a wavelength of 260 and 274 nm for KETO and MPT, respectively. Drug release profiles were compared using the difference (f1) and similarity factor (f2) (as introduced by Moore and Planner). (Moore and Planner, 1996). The drug release profiles show similarity when the value of f2 is greater than 50 and the value of f1 is less than 15.

3 Results & Discussion

3.1 Characterization of raw materials and physical mixtures

3.1.1 Crystallization kinetics

The half crystallization times as function of the various applied isothermal crystallization temperatures for the samples containing PEO are shown in Fig. 1c. The results clearly show that the
crystallization speed of PEO is slower with increasing isothermal crystallization temperatures. Earlier research has demonstrated that a higher degree of undercooling (i.e., \( \Delta T = T_m^\ast - T_c \)) results in a larger driving force and speeds up crystallization (Crist and Schultz, 2016). The equilibrium melting temperature \( T_m^\ast \) is defined as the melting temperature of an extended chain crystal and is usually determined by the Hoffman-Weeks approach. Therefore, the polymer is crystallized at various isothermal temperatures \( T_c \) and subsequently reheated to determine the peak melting point. A linear fit is applied to the \( T_m \) vs \( T_c \) data and the intercept of the fit with a line \( T_c = T_m \) gives \( T_m^\ast \). Figure 1d shows that \( T_m^\ast \) for the PEO-20KETO (70.41 °C) sample is lower than for the neat polymer (72.61 °C). Consequently, the degree of undercooling is also reduced at a given \( T_c \), and nucleation and growth are much slower for PEO-20KETO due to the lower degree of undercooling at the same \( T_c \) (Fowler et al., 2010). KETO is dissolved in the molten polymer and inhibits chain folding by interacting with the polymer chains, inhibiting the formation and folding of the polymer into lamellae (Marentette and Brown, 1998). For example, the half crystallization time of pure PEO measured by rheology was 3.7 min at 47 °C, whereas it was almost a tenfold (33.5 min) for PEO-20KETO. It should be noted that crystallization of PEO was not fully completed within the experimental time (60 min) at 46 °C for the PEO-20KETO sample measured by DSC (Fig. 1b), therefore the half crystallization calculation was not possible. Furthermore, the rheological half crystallization times were always longer compared to the DSC experiments \( T_c = 46 \) °C not included). This can be explained by the differences in sample mass, the cooling rate differences (5 °C or 50 °C/min) and the sensitivity of the technique for detecting early crystallization. Moreover, these results confirm that the frequency (1Hz) and deformation (2%) used during the rheological tests did not affect the crystallization kinetics as otherwise the rheological measurements should have shown faster crystallization compared to the DSC experiments due to the applied oscillatory stress.
Figure 2 shows the results of the isothermal crystallization experiments for the samples containing PCL. Similar to the PEO results, PCL crystallization speed is also reduced at higher isothermal temperatures. Both DSC and rheological crystallization experiments are complementary and showed the same trend (Fig. 2a - b). Both techniques indicated that the solid-state of the drug in the polymer melt had an impact upon the crystallization speed of PCL. The crystallization speed of PCL was depressed when the PCL-MPT mixture was heated above the melting temperature of MPT and subsequently cooled to the isothermal crystallization temperature. Molten MPT clearly acted as a plasticizer and inhibited PCL crystallization. For example, the half crystallization time measured by DSC was 10.13 min for pure PCL at 43 °C and 21.30 min for the plasticized PCL melt. At 44 °C, crystallization was not completed within the experimental time (60 min) during the DSC run as can be seen in Fig. 2d.

In contrast to molten MPT, crystalline MPT increased the crystallization speed of PCL. Crystalline MPT acted as nucleating agent, decreasing the half crystallization time from 10.13 min for PCL to 9.39 min for PCL-20MPT and 9.41 min for PCL-40MPT at 43 °C. Similar results are determined from the rheological measurements (Fig. 2b), supporting the hypothesis. Increasing the MPT concentration from 20 to 40% did not result in a further increase of the polymer crystallization speed, suggesting that the nucleating efficiency of crystalline MPT reached a saturation point. This finding is supported by the equilibrium melting temperature, which slightly increased by the addition of crystalline MPT ($T_m^{PCL}$ PCL = 61.46 °C; $T_m^{PCL-20MPT}$ = 62.36 °C; $T_m^{PCL-40MPT}$ = 61.82 °C). Therefore, the driving force for nucleation and crystal growth of PCL increased by the addition of MPT crystals. Furthermore, the half crystallization times measured by rheology were longer than the half crystallization times measured by DSC, as was also the case for the PEO samples.
3.1.2 Rheological characterization

Figure 3 shows the complex viscosity as function of temperature for samples containing PEO (Fig. 3a) and PCL (Fig. 3b). Neat PEO and the PEO-20KETO mixture showed a decreasing complex viscosity upon heating with a sharp decrease which indicated melting of the PEO crystals. KETO tended to depress the melting point of PEO and reduced the complex viscosity of the drug-polymer mixture compared to pure PEO. Schachter et al. showed that KETO/PEO interactions can be detected even in non-melt processed blends at temperatures as low as 43 °C, hereby explaining the melting point depression upon heating of the physical mixture (Schachter et al., 2004). Crystallization of the polymer upon cooling from the melt occurred at lower temperatures for the PEO-20KETO mixture since nucleation of polymer crystals is inhibited by KETO, confirming the previously described DSC crystallization experiments. As for the PCL-20MPT mixture, the PCL crystals started melting at a temperature of 56.8 °C during the first heating cycle. Moreover, MPT acted as a filler (i.e., increasing the complex viscosity) above the melting point of PCL and below its own melting point (i.e., 123 °C), but plasticized (i.e., lower complex viscosity) the molten mixture when the drug crystals melted. Crystallization of PCL upon cooling is slightly inhibited by molten MPT since the crystallization commenced at a lower temperature compared to pure PCL. During a second heating run of the PCL-20MPT mixture, the viscosity decreased again to the same level as during the cooling run, indicating only melting of the PCL crystals while MPT is dissolved in the matrix. Also, the melting point of PCL during the second heating run was slightly lowered compared to the first heating cycle which can be due to the refolding of the polymer chains into a less stable form compared to the original extended chains. Based on the latter temperature sweep experiments, an extrusion temperature of 80 °C was selected to process formulations while MPT maintained its crystalline state, while PCL/MPT mixtures were extruded at 140 °C in order melt the drug fraction. Furthermore, based on the crystallization kinetics during the temperature sweep test, 25 °C and 35 °C were chosen as mold temperatures to produce the IM tablets. The lowest temperature (25 °C) was selected based on an anticipated fast crystallization, while the highest temperature (35 °C) represented a temperature during which
crystallization was still ongoing during the temperature sweep experiment (showed in Fig. 3 by the vertical dotted line), hence a slower crystallization is expected at this temperature.

3.1.3 Differential scanning calorimetry

The thermograms for physical mixtures containing PEO are shown in figure 4a. The first heating cycle confirmed the previously described rheological temperature sweep results since the melting point of PEO was depressed from 65.57 °C (pure PEO) to 63.54 °C (PEO-20KETO) and no melting endotherm representing KETO crystals was observed, suggesting dissolution of KETO crystals in the PEO matrix. The cooling cycle showed a higher crystallization temperature upon cooling of pure PEO (40.92 °C) compared to the PEO-20KETO mixture (32.92 °C), confirming the previously described crystallization inhibition theory. The thermograms of samples containing PCL are shown in figure 4b. The first cycle showed melting of PCL and a second melting peak for MPT at 117 °C which is lower than pure MPT (123 °C), suggesting partial dissolution of MPT crystals in the PCL matrix. Cooling of molten MPT mixture showed a slightly lower polymer crystallization temperature (28.58 °C) as compared to pure PCL (29.25 °C). On the other hand, when the PCL-20MPT mixture was only heated to 100 °C and subsequently cooled, a higher crystallization temperature is measured (30.77 °C). This can be explained by the nucleating effect of MPT crystals. The 40% MPT mixture showed the same trend as the crystallization temperature was further increased to 31.33 °C.

3.2 Tablet characterization

3.2.1 Solid-state analysis and crystalline microstructure
The DSC results from the IM tablets are summarized in table 3. KETO was completely transformed from the crystalline to the amorphous state during extrusion and subsequent injection molding. Also, MPT was partially transformed to the amorphous state when extruded at 140 °C, but remained crystalline when extruded at 80 °C. The crystallinity of PEO during annealing in the mold increased significantly (from 66.29 to 70.14%) when annealed at 35 °C compared to annealing at 25 °C. On the other hand, the PCL-20MPT and PCL-40MPT formulation processed at 80 °C and 140 °C did not show a significant increase in the polymer crystallinity. However, the peak melting temperature did increase (from 50.40 to 51.92 °C for PCL-20MPT; from 55.95 to 56.96 °C for PCL-40MPT), suggesting that the polymer microstructure had changed. The reason for the polymer microstructural differences originates from the various applied cooling rates. Higher cooling rates are obtained when cooling from the melt to a lower mold temperature (25 °C), therefore giving less time to the melt to form more perfect crystals due to the quick transition from a molten material to a structured crystalline solid state. In order to confirm this hypothesis, the microstructure of the PCL crystals was visualized using POM. The crystalline microstructure of pure PCL consists of impinged spherulite structures (Fig. 5). Injection molding of pure PCL at 25 °C (Fig. 5a) gave smaller structures compared to the sample collected after injection molding at 35 °C (Fig. 5b). A semi-quantitative analysis was performed and provided as supplementary material (Fig SI2). Moreover, the incorporation of MPT crystals (Fig. 5 c-d) in the melt reduced the size of the PCL microstructure at both injection mold temperatures (i.e., 25, 35 °C) compared to neat PCL.

3.2.2 Drug dissolution

Immediate release formulation

The cumulative release of KETO from the PEO matrix tablets is shown in figure 6. The release of KETO from the IM tablets reached 100% within 4 hours. Calculated similarity (f2 = 70) and difference factor (f1 = 5) indicated that there is no significant difference between the dissolution profiles of the tablets injection molded at 25 °C and 35 °C. For immediate release formulations containing rapidly
erodible and water soluble semi-crystalline polymers, the drug dissolution is mainly controlled by swelling and erosion of the polymer. For this low molecular weight PEO, erosion is the dominant mechanism according to Cantin et al. (Cantin et al., 2016). Therefore, differences in polymer crystallinity and microstructure are insignificant for controlling the drug dissolution for this type of formulation. However, the drug release can be controlled by addition of erosion-promoting substances. Crospovidone showed to increase the KETO release from PEO melt processed tablets by increasing the rate of erosion of the PEO matrix (Schachter et al., 2004). Furthermore, the molecular weight of PEO is well known to have an impact on the drug release (Cantin et al., 2016). High molecular weight PEO is often used to provide sustained release of the API (Monteyne et al., 2016).

**Sustained release formulation**

The drug release profiles from the formulations containing 20% (w/w) MPT are shown in Fig. 7a. As a result of the poor wettability of the PCL matrix, a full release of the drug cannot be obtained within 24 hours. The MPT release profile of the 20% MPT formulation showed a significant increase of the total drug release when extruded above the drug melting point. This is due to MPT being partially transformed to the amorphous state during extrusion (see table 3). However, no significant difference in drug release was found between the tablets prepared at various mold temperatures (25 and 35 °C) after extrusion at 140 °C. The variance of the drug release profile was higher for the tablets extruded at 140 °C, which can be explained by the large variance in amorphous drug obtained (see table 3). On the other hand, the drug release is significantly lower from tablets produced after extrusion at 80 °C due to the crystalline state of the drug. Moreover, the drug release profiles from annealed tablets molded at 25 °C and 35 °C were not significantly different according to the calculated similarity (f2 = 61) and difference factor (f1 = 86). However, the curves showed a certain similarity in shape but a more pronounced burst-effect was observed for tablets prepared at mold temperature of 25 °C. It is well known that polymer crystallinity and morphology has an impact on the water vapor permeability (Duan and Thomas, 2014). Larger and more perfect crystals present a stronger barrier for water diffusion. By
increasing the drug load to 40% (w/w), a faster and more complete release of MPT was found (Fig. 7b).

Similar results are found for the 40% (w/w) MPT formulation wherein the tablets produced at 25 °C showed a significant (factor f2 = 41 and f1 = 84) higher percentage drug release after 24 hours compared to tablets prepared at a mold temperature of 35 °C. These differences in drug release between both injection mold temperatures can be explained by the various crystal morphologies obtained (shown in previous section). Larger and more perfect crystal lattices were obtained in case a higher mold temperature was used. As a result, the drug diffusion rate through the water insoluble matrix was decreased. Furthermore, the polymer microstructure of the tablets can change over time during long term storage. Therefore, crystal growth and polymer degradation are likely to become more important for the control of drug release at later time points.

4 Conclusion

The impact of the injection mold temperature upon the polymer crystallinity and microstructure and hence the drug release was investigated. Isothermal crystallization experiments showed that the temperature and the drug solid-state had an impact on the polymer crystallization kinetics. Molten or dissolved drug inhibited the polymer crystallization, whereas a crystalline drug acted as filler/nucleating agent, enhancing the polymer crystallization upon cooling from the melt. The polymer crystallinity increased at higher mold temperatures for the PEO tablets but did not significantly change for PCL tablets. The drug release from the PEO matrix was not affected by the various polymer crystallinities induced by a difference in mold temperature. This is due to the erosion mechanism, controlling the drug release for the immediate release formulation. On the other hand, the drug release from the PCL matrix, which is purely diffusion controlled, was influenced by the polymer microstructure. It is shown in this study that the polymer microstructure can be controlled by applying various thermal histories. Larger polymer crystals are obtained using higher mold temperatures, reducing the drug diffusion rate through the polymer matrix. These results imply that the injection
mold temperature should be carefully controlled for sustained release formulations containing hydrophobic semi-crystalline polymers.

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6 References

Alexis, F., 2005. Factors affecting the degradation and drug-release mechanism of poly(lactic acid) and poly[(lactic acid)-co-(glycolic acid)]. Polym. Int. 54, 36–46. doi:10.1002/pi.1697


Table 1: Isothermal crystallization experiments.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Melt equilibration temperature (°C)</th>
<th>Isothermal crystallization temperatures (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PEO</td>
<td>100</td>
<td>46 – 47 – 48 – 49 – 50 – 51</td>
</tr>
<tr>
<td>PEO-20KETO</td>
<td>100</td>
<td>42 – 43 – 44 – 45 – 46 – 47</td>
</tr>
<tr>
<td>PCL</td>
<td>100</td>
<td>39 – 40 – 41 – 42 – 43 – 44</td>
</tr>
<tr>
<td>PCL-20MPT</td>
<td>80 and 140</td>
<td>39 – 40 – 41 – 42 – 43 – 44</td>
</tr>
</tbody>
</table>
### Table 2: Process parameters for HME and IM.

<table>
<thead>
<tr>
<th>Formulation</th>
<th>Screw speed</th>
<th>Barrel temperature</th>
<th>Mold temperature</th>
<th>Solidification time</th>
</tr>
</thead>
<tbody>
<tr>
<td>80 % PEO 20 % KETO</td>
<td>150 rpm</td>
<td>100 °C</td>
<td>25 °C</td>
<td>1 min</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>35 °C</td>
<td>1 min</td>
</tr>
<tr>
<td>80 % PCL 20 % MPT</td>
<td>150 rpm</td>
<td>80 °C</td>
<td>25 °C</td>
<td>1 min</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>35 °C</td>
<td>2 min</td>
</tr>
<tr>
<td></td>
<td>150 rpm</td>
<td>140 °C</td>
<td>25 °C</td>
<td>2 min</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>35 °C</td>
<td>3 min</td>
</tr>
<tr>
<td>60 % PCL 40 % MPT</td>
<td>150 rpm</td>
<td>80 °C</td>
<td>25 °C</td>
<td>1 min</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>35 °C</td>
<td>1 min</td>
</tr>
</tbody>
</table>
Table 3: Polymer and drug crystallinity from IM tablets reported as average values with standard deviation between brackets (n = 6). E = extrusion barrel temperature; M = mold temperature. Tm = polymer peak melting point.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Tm</th>
<th>Polymer crystallinity (%)</th>
<th>Drug crystallinity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(°C)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>80% PEO 20% KETO</td>
<td>55.45</td>
<td>66.29 (0.94)</td>
<td>Amorphous</td>
</tr>
<tr>
<td>E = 100 °C M = 25 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>80% PEO 20% KETO</td>
<td>56.08</td>
<td>70.14 (0.50)</td>
<td>Amorphous</td>
</tr>
<tr>
<td>E = 100 °C M = 35 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>80% PCL 20% MPT</td>
<td>55.99</td>
<td>50.40 (0.39)</td>
<td>95.48</td>
</tr>
<tr>
<td>E = 80 °C M = 25 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>80% PCL 20% MPT</td>
<td>56.75</td>
<td>51.92 (0.27)</td>
<td>96.96</td>
</tr>
<tr>
<td>E = 80 °C M = 35 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>80% PCL 20% MPT</td>
<td>56.58</td>
<td>52.63 (0.28)</td>
<td>20.98</td>
</tr>
<tr>
<td>E = 140 °C M = 25 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>80% PCL 20% MPT</td>
<td>56.99</td>
<td>52.72 (0.32)</td>
<td>13.11</td>
</tr>
<tr>
<td>E = 140 °C M = 35 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>60% PCL 40% MPT</td>
<td>55.95</td>
<td>48.57 (0.46)</td>
<td>98.65</td>
</tr>
<tr>
<td>E = 80 °C M = 25 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>60% PCL 40% MPT</td>
<td>56.96</td>
<td>50.85 (0.32)</td>
<td>99.67</td>
</tr>
<tr>
<td>E = 80 °C M = 35 °C</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Figure 1: Heat flow during isothermal crystallization for (a) PEO and (b) PEO-20KETO. c) Half crystallization time vs isothermal crystallization temperature for PEO samples measured by DSC and Rheology. d) Hoffman-Weeks plot for samples containing PEO.
Figure 2: Half crystallization time as function of isothermal crystallization temperatures for samples containing PCL measured by (a) DSC and (b) Rheology. Heat flow as function of isothermal time for PCL-20MPT below (c = 80 °C) and above (d = 140 °C) drug melting point and PCL-40MPT (e) below the drug melting point. (f) Hoffman-Weeks plot for samples containing PCL samples.
Figure 3: Rheological temperature sweep experiments for (a) PEO and PEO-20KETO and (b) PCL and PCL-20MPT. HR = heating run, CL = Cooling run. Vertical dotted lines indicate 25 °C and 35 °C, which were chosen as mold temperatures.
Figure 4: DSC thermograms of the first heating (solid line) and subsequent cooling phase (dashed line) of (a) PEO and PEO-20KETO, and (b) PCL, PCL-20MPT and PCL-40MPT mixtures.
Figure 5: PCL extruded at 80 °C and injection molded at a) 25 °C and b) 35 °C. PCL-20MPT extruded at 80 °C and injection molded at c) 25 °C and d) 35 °C. Magnification = 10 x
Figure 6: Cumulative drug release of PEO-20KETO formulation from tablets (n = 5) prepared at mold temperatures of 25 °C (blue dots) and 35 °C (red squares).
Figure 7: Cumulative drug release of (a) PCL-20MPT and (b) PCL-40MPT tablets (n = 5). E = extrusion temperature, M = mold temperature. The variance of the drug release is estimated by the standard deviation.
Supplementary information for

The impact of the injection mold temperature upon polymer crystallization and resulting drug release from immediate and sustained release tablets.

Jeroen Van Renterghem*, Heleen Dhondt, Glenn Verstraete, Michiel De Bruyne, Chris Vervaet, Thomas De Beer

*Corresponding author: Jeroen Van Renterghem

Laboratory of Process Analytical Technology, Ghent University, Ottergemsesteenweg 460, 9000 Ghent, Belgium
TEL: 0032 9 264 8039
FAX: 0032 9 264
E-MAIL: jeroen.vanrenterghem@ugent.be
Trimming process.

Figure S1: Trimming procedure to obtain samples before sectioning into thin (3µm) samples using a microtome. a) IM tablet attached to the entry section (i.e., gate). b) Tablet cut in half with a razor blade. f) Trimming of the tablet and mounting on an aluminum pin. g) Final sample obtained before cryosectioning.
Semi-quantitative analysis of POM images:

The background of the original images (Fig. SI2 a,b) was subtracted using the background threshold option from imageJ. The remaining black structures of image c and d are related to the crystal structures (i.e., spherulites). The structures of figure c (IM temperature: 25 °C) are smaller and of greater number (5.57 µm, count: 8632) compared to the structures (7.77 µm, count: 7920) of figure d (IM temperature: 35 °C).

Figure SI2: Original POM images of PCL extruded at 80 °C a) IM = 25 °C and b) IM = 35 °C. The same POM images after subtraction of the background c) IM = 25 °C d) IM = 35 °C. Scale bar = 50µm, magnification = 20x, IM = injection mold temperature.