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Comparison of species classification models of mass spectrometry data: Kernel Discriminant Analysis vs. Random Forest; A case study of Afrormosia (*Pericopsis elata* (Harms) Meeuwen)

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Rationale:

The genus *Pericopsis* includes four species of which only *Pericopsis elata* (Harms) Meeuwen is of commercial interest. Enforcement officers might have difficulties discerning this CITES-listed species (Appendix II) from some other tropical African timber species. Therefore, we tested several methods to separate and identify these species rapidly in order to enable customs to uncover illegal trade. In this study, two classification methods using DART TOFMS data to discern between several species are presented.

Methods:

Metabolome profiles were collected using DART TOFMS analysis of heartwood specimens of all four *Pericopsis* species, *Haplormosia monophylla* (Harms) Harms, *Dalbergia melanoxylon* Guill. & Perr. Harms, and *Milicia excelsa* (Welw.) C.C.Berg. In total, 95 specimens were analysed and the spectra evaluated. The species were chosen based on their importance in CITES, belonging to the same genus, and wood morphological resemblance. Kernel Discriminant Analysis (KDA) and Random Forest classification were used to discern the species.

Results:

DART TOFMS spectra obtained from wood slivers and post-processing analysis using KDA and Random Forest classification separated *Pericopsis elata* from the other *Pericopsis* taxa and its lookalike timbers *Haplormosia monophylla*, *Milicia excelsa*, and *Dalbergia melanoxylon*. Only 50 ions were needed to achieve the highest accuracy.

Conclusions:

DART TOFMS spectra of the taxa were reproducible and the results of the chemometric analysis provided comparable accuracy. *Haplormosia monophylla* was visually distinguished based on the heatmap and was excluded from further analysis. Both classification methods, KDA and Random Forest, were capable of distinguishing *Pericopsis elata* from the other *Pericopsis* taxa, *Milicia excelsa*, and *Dalbergia melanoxylon*, timbers commonly traded.

Key words: *Pericopsis*; DART TOFMS; Random Forest; KDA; Forensic; Timber identification;

INTRODUCTION

Species characteristics and international trade

Pericopsis elata, commonly known as Afrormosia, is an emblematic species of the African rainforests and has been protected by the Convention on International Trade in Endangered Species (CITES)^[1] since 1992. Its heartwood is characterized by high natural durability, mechanical strength, and dimensional stability. This combination makes it suitable for the most demanding applications of wood, especially for exterior joinery. The decorative value of the wood is also appreciated for the production of luxury furniture and parquetry. In some parts of the rainforest belt, the species is common and available in quantities large enough for industrial logging and the international timber trade. The market discovered the species as a precious wood, named Afrormosia, after the Second World War. The *P. elata* populations of Ghana were logged followed by the populations of Côte-d'Ivoire soon after.^[2] These loggings were not based on management plans aiming at a sustainable yield and resulted in overexploitation after only a few decades. The West-African countries are therefore no longer considered as a source of *P. elata* timber. The logging shifted gradually to the Central African rainforests of Cameroon, the Republic of Congo (where the species is relatively rare), and the Kisangani region of the Democratic Republic of the Congo (DRC) in this order. There are two additional *Pericopsis* species in Africa: *P. angolensis* (Baker) Meeuwen and *P. laxiflora* (Baker) Meeuwen, whereas a single species is endemic to Asia: *P. mooniana*. *Pericopsis angolensis* also produces high quality durable timber, but trees of this species are less abundant and too small or poorly shaped for commercial exploitation, except in Mozambique, where the wood is sometimes traded as Muwanga or mixed with harvested *Pericopsis elata* timber.^[2] *Pericopsis laxiflora*, is similar, with the same uses but is not available in large sizes, and is by some researchers considered a subspecies of *Pericopsis angolensis*.^[2] The Asian species, *Pericopsis mooniana*, which ranges from Sri Lanka east to New Guinea and Micronesia is mainly exported from Indonesia to Japan.^[3]

Because of law enforcement concerns, there is a need for robust identification of *P. elata* from the other *Pericopsis* species and lookalike timbers. There have been documented fraudulent imports of *P. elata* declared as *Milicia excelsa*, a non-CITES listed species. The timber of *P. elata* can also be

confused with *Dalbergia melanoxylon* (CITES App. II) from Africa. Traditional identification of wood has relied on anatomical features such as those in the extensive online database InsideWood.^[4] When searching InsideWood using standardised wood anatomical features of *P. elata*, the results indicate that several other species have similar wood structures such as *Dalbergia melanoxylon* and *Haplormosia monophylla*. *Dalbergia melanoxylon* is a timber species that also occurs in Central Africa, *Haplormosia monophylla* is taxonomically closely related to *P. elata*, also occurs in Africa and is traded by the common name of Idewa. To a lesser extent, the timber of the three other *Pericopsis* species might also be sold or confused with *P. elata*.^[2]

Species identification based on wood anatomy

It is therefore important for law enforcement officers to be able to discriminate between the abovementioned species. The anatomical features described in the IAWA Hardwood List^[5] and used in InsideWood are adequate for narrowing down the number of possible identities of a hardwood sample, but their discriminatory ability is limited for closely related taxa with very similar features. This is the case with *Pericopsis* and *Haplormosia*, which have similar paratracheal axial parenchyma (ranging from scanty paratracheal through vasicentric to aliform to confluent and banded especially in *Haplormosia*), storied axial parenchyma in mainly four-celled strands, and rays generally up to three or four cells wide and storied. *Pericopsis mooniana* appears to have sparser vessels and more distinct aliform and confluent parenchyma than the other *Pericopsis* species, but this observation is based on a single microscope slide in Kew's reference collection and the literature in InsideWood and Plant resources of South-East Asia 5 (PROSEA)^[3]. It becomes more difficult to differentiate between *P. angolensis*, *P. elata* and *P. laxiflora* based on wood anatomical features. Comparing these species using InsideWood leads to minor and variable differences. Only *P. elata* appears to have vasicentric axial parenchyma. However, this was also present in two transverse sections of *P. angolensis*, provided by the Royal Museum for Central Africa (RMCA, Tervuren, Belgium). *Pericopsis laxiflora* appears to have more bands of parenchyma and few to no high rays. This was observed by comparison of two transverse and tangential sections with two and three slides of *P. elata* and *P. angolensis* respectively. Another interesting feature is the presence of unilateral parenchyma in *P. elata*, which is rarer in *P. angolensis* and almost lacking in *P. laxiflora*. The wood of *Milicia excelsa* is very unlikely to be confused with *P. elata* or *Haplormosia monophylla* under the microscope because none of the cells are storied, the rays are wider with a single row of upright cells at the margins and each one often contains a single prismatic crystal. However, *Milicia excelsa* has been confused with *P. elata* based on morphological macroscopic wood features. InsideWood shows extensive anatomical information on *Dalbergia melanoxylon*. As stated before, using InsideWood with the standardised wood anatomical features of *P. elata* may lead to wood

identification as a *Dalbergia* species. Therefore, using the wood anatomical database alone could lead to an incorrect species identification.

Using DART TOFMS data for species identification

Direct Analysis in Real Time (DART) (see Cody *et al.*^[6]) Time-Of-Flight Mass Spectrometry (TOFMS) has shown promise in the analysis of wood and plants. Previous research using DART TOFMS spectra was able to distinguish between two species of American oak (*Quercus alba* L. and *Quercus rubra* L.)^[7], between four species of agarwood (*Aquilaria spp.*)^[8,9], and between *Dalbergia* timbers from Africa, Madagascar and Asia.^[10] Recent research focused on the identification of plant species (*Mitragyna speciosa* (Korth.) Havil. and *Datura*)^[11], discrimination among selected CITES-protected *Araucariaceae*^[12] and differentiating coastal from inland populations of Douglas-fir (*Pseudotsuga menziesii* (Mirb.) Franco) using Random Forest classification algorithms.^[13] The main goal of this study is to determine if we can separate *Pericopsis elata* using DART TOFMS spectra from the following species: *Pericopsis angolensis*, *Pericopsis laxiflora*, *Pericopsis mooniana*, *Milicia excelsa*, *Haplormosia monophylla* and *Dalbergia melanoxylon*. We will also determine (1) which classification technique, Kernel Discriminant Analysis (KDA) or Random Forest, performs better to separate these species, (2) if by using the variable (ions) importance lists retrieved from the Random Forest, we can improve the KDA, and (3) the lowest number of ions needed to separate the species.

EXPERIMENTAL

Material

Heartwood samples of all *Pericopsis* species, *Milicia excelsa*, *Haplormosia monophylla* and *Dalbergia melanoxylon* were randomly collected. The Table in the Appendix lists the different samples with their geographic provenance, country of origin, the source and number of specimens.

Species validation of the commercial timber samples was done by comparing their spectra against those of curated xylaria reference samples.

DART-TOFMS analysis

The specimens were analysed using a DART-SVP ion source (IonSense, Saugus, MA, USA) coupled to a JEOL AccuTOF 4G LC Mass Spectrometer (Jeol USA, Peabody, USA). Heartwood slivers are put directly in a stream of heated helium gas, produced by the DART ion source. This gas has a temperature of 350°C leading to ionization and the emission of compounds from the wood entering the mass analyser. Spectra were acquired in positive ion mode with the DART ion source parameters and mass spectrometer settings as defined in Evans *et al.*^[12], McClure *et al.*^[10], Lancaster &

Espinoza^[8] and Espinoza *et al.*^[9] The text files of the mass-calibrated, centroided mass spectra were exported using TSS Unity (Shrader Software Solutions, Inc., Grosse Pointe Park, MI, USA) data reduction software and used for further analysis.

Specimen classification methods

A heatmap, showing the intensity of each ion-mass (mass to charge ratio m/z) in the sample (Figure 1), was created using the Mass Mountaineer Mass Spectral Interpretation Tools software (RBC Software, Peabody, MA, USA). Next, KDA was performed with the same Mass Mountaineer software package using a tolerance of 5 mmu and a 1% threshold. KDA is a generalization of linear discriminant analysis (LDA) where the principal components are nonlinearly related to the input variables in the transformed space.^[14] Each specimen is assigned to a class in the grouping variable (in this case species), and KDA then calculates the maximum separation between species classes in a training set. This is then mapped in the non-linear higher-dimensional space.^[12] KDA determines the species separation based on a selection set of appropriate ions. The selection of ions is based on selecting variables that are unique to one species or ions that show higher intensity in one taxon but lower in others. This process is simplified by a visual inspection of the heatmap. Model accuracy is assessed using leave-one-out-cross-validation (LOOCV, see Lever *et al.*^[15]; McClure *et al.*^[10]).

The results of the KDA were compared to the Random Forest method, which is implemented in the RandomForest package^[16] in R. All calculations were done in RStudio (RStudio Team, 2015). Spectral data were exported from Mass Mountaineer (tolerance of 250 mmu and 1% threshold) to Microsoft Excel and imported into RStudio. A Random Forest is best described as a set of n regression or classification trees.^[17,18] The dataset is randomly split into a training and validation dataset, in this case 80 % and 20 % respectively. Each tree is constructed using different subset samples of the training dataset with the objective of classifying each sample to a class in the grouping variable (species). Each node in the tree is split using the best predictor variable, here ion relative abundance, among a randomly chosen subset of predictor variables.^[16,19] In total 10,000 classification trees were created to build the Random Forest with 50 randomly chosen ions at every node split. Model accuracy is determined by the out-of-the bag principle (OOB). At each bootstrap iteration the samples that were not used in the training set are used to validate the current tree in that bootstrap iteration.^[16] The overall OOB is reported as the estimation of the error rate, indicating the misclassification of samples. Instead of using the OOB classification error, we report the compliment, or the Random Forest classification accuracy, to compare with the validation rate of LOOCV in KDA (OOB error + Random Forest classification accuracy = 1).^[13] Before determining the performance of the Random Forest through the validation dataset, the classification error of the samples per species in the training set is given. This is a first indication of which species will be problematic. Finally, the performance

of the Random Forest classification is determined using the validation dataset to test the model. Several measures for variable importance can be assessed as well, which in this case indicate specific ions that are key for differentiating among species. The first measure is the Gini-index or Mean Decrease in Impurity (MDI), which is used to quantify the impurity in each node.^[18] A second measure, based on permutation of the OOB data, is the Mean Decrease in Accuracy (MDA) and aims at improving the accuracy. The difference in prediction accuracy is a good indicator of variable importance.^[18] For a comprehensive review of MDI vs MDA the authors refer to Alex Perrier's "Feature Importance in Random Forest"^[20].

The list of ions, ranked by variable-importance were then used for KDA to determine if the Random Forest generated ions give a higher classification accuracy than the empirical ion selection described above. Experiments were conducted with different numbers of ions (5, 10, 20, 30, 50, 100, 200, and 256 ions) based on the importance values from the Random Forest.

Model comparison

The KDA results were compared to the Random Forest classification under two different conditions. The KDA of the *Pericopsis* and the lookalike species was based on 65 ions. This experiment excluded *Haplormosia monophylla* because of the small sample size. The second KDA consisted of only *Pericopsis* taxa, and 248 ions were used for the classification. The species used and their respective sample sizes are listed in Table 1.

RESULTS AND DISCUSSION

Figure 1 shows the heatmap for the different species analysed. *Haplormosia monophylla*'s chemotype is present in the heat map but was removed from the classification models due to the small sample size. However, this species could be separated from the other species by looking at the heatmap. As shown in Figure 1, the chemotypes for *H. monophylla*, *M. excelsa*, and *D. melanoxydon* are different from the chemotypes of the *Pericopsis* species. Additionally, some differences appear among the chemotypes of the *Pericopsis* taxa. For example, *P. laxiflora* seems to have more intense ions around 409 *m/z*. *P. angolensis* can be differentiated by looking at the ions around 285 and 299 *m/z* and a lack of consistent intensity around 205 *m/z*. The surprising find that the chemotype of *P. laxiflora* is dissimilar to the chemotype of *P. angolensis* erodes the support for the hypothesis that *P. laxiflora* could be a subspecies of *P. angolensis*^[2]. However, a larger sample size of *P. laxiflora* is needed to statistically test this observation. Only *P. mooniana* has ions at 260 and 273 *m/z*, these are absent in the other *Pericopsis* species. Figures 2 and 3 show the KDA graphical representation of two different datasets. Figure 2 shows that the KDA classification algorithms clustered each of the *Pericopsis*

species separately, whereas Figure 3 shows the separation of the protected *P. elata* from the other species.

The main goal of this study was to separate the CITES-listed *Pericopsis elata* from the other species in its genus and from its lookalikes. The LOOCV (KDA) was 95.79%, and the classification accuracy of the Random Forest was 96.05%, indicating that both KDA, with the empirically chosen ions, and Random Forest enabled us to correctly identify *Pericopsis elata* to a satisfactory level. Table 2 shows the confusion matrix, which summarizes the classification of the training data set for the Random Forest. As can be seen, *P. elata* shows the lowest classification accuracy, but it is still high. Only two out of 16 samples are misclassified. Afterwards, the Random Forest is validated using the prediction data. Table 3 shows the result for the classification of the prediction data. In this example, the Random Forest classified all samples correctly. Next, we tried to differentiate between *Pericopsis* species. The LOOCV was 88.89% and the random forest accuracy was 93.75%. These results are, however, based on an unbalanced dataset, with only five samples for *Pericopsis laxiflora* compared to, for example, the 21 samples from *Pericopsis angolensis*. Although the final model performance is satisfactory, it might impact the model variability and the handling of misclassifications.^[13] The overall classification accuracy is not significantly affected; however, this is often not an appropriate performance measure in learning extremely unbalanced data.^[21] Using these small unbalanced datasets increases the risk of leaving a certain species out of the training dataset bootstrapping, skewing the model towards the more abundant species. Possible solutions are suggested by Chen *et al.*^[21] This was however outside the scope of the current study and should be investigated further.

Supplementary goals of this study were to determine if empirically selected variables (ions) provided the same level of accuracy as the ions selected by the Random Forest algorithm and the minimum number of ions required to obtain the highest classification accuracy. Random Forest provides a two ways of determining variable (i.e., ions) importance, the MDI and MDA. The Random Forest analysis produces a ranked list of variables (ions) that have the highest importance in separating classes, and in this case, the most valuable ions for separating *P. elata* from the other lookalike species. This ranked list of ions was used to perform a KDA analysis, and the results were compared to the Random Forest-selected ions.

Table 4 compares the accuracy of both approaches using the calculated LOOCV. Additionally,

Table 4 shows the results when different numbers of ions were used and the resulting LOOCV based on the most important ions according to MDI and MDA (ranging from 5 to 256 ions). It is clear that the highest LOOCV accuracy, for the lowest number of ions, was obtained with the 50 most important ions selected by the MDI algorithm of Random Forest.

CONCLUSION

We have demonstrated that the identification of *P. elata* can be obtained using DART TOFMS spectra. Although the heatmaps of the *Pericopsis* taxa appear to be equivocal, the statistical post-processing of the spectra can be used to identify species with high accuracy. The chemotypes shown in the heatmap (Figure 1) of *M. excelsa*, *D. melanoxylon*, and *H. monophylla* are very different from the four other *Pericopsis* species, and minor differences on the chemotypes of the *Pericopsis* taxa can also be observed. The chemotype of *P. laxiflora* is dissimilar to the chemotype of *P. angolensis*. This is not in support of the hypothesis that *P. laxiflora* could be a subspecies of *P. angolensis*^[2]. A larger sample size of *P. laxiflora* will allow to statistically test this observation..

Taxa classification using KDA and Random Forest algorithms have similar and satisfactory success rates, showing that both methods can be used to determine the species identity. A recurring challenge when performing KDA is to determine which variable to use, this may lead to overfitting and bias through arbitrary selection of discriminant ions. However, Random Forest algorithms are based on dataset training, multiple bootstrap iterations, and not on analyst judgment and do incorporate all ions detected among samples. The results from Random Forest classification analysis are therefore more objective. It was, however, interesting to note that KDA through manual selection of ions provides similar classification accuracy to Random Forest. We show that the variable importance measures included in the Random Forest can however aid in the ion choice for KDA. For this case we conclude only 50 ions are needed to achieve the best accuracy. We conclude that, in addition to wood anatomy, suspected wood samples can be analysed with DART TOFMS and resulting spectra can be evaluated using either KDA (LOOCV 95.79%) or Random Forest (96.05%).

Ultimately, DART TOFMS and post processing analysis of the spectra provides robust identifications of timbers when traditional wood anatomical methods are equivocal or if the required expertise is unavailable, and these tools provide an additional method for combatting illegal timber trade. The success of this method naturally depends on the availability of samples to ensure balanced datasets. The authors would like to take this opportunity to invite xylaria throughout the world to share their vast collections.

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FIGURES

Figure 1: Heatmap of the ions present in the analysed samples. Y-axis indicates sample number and the x-axis indicates the mass to charge ratio. The intensity of the red squares in the heatmap correlates with the abundance of the ions in the samples.

Figure 2: Kernel Discriminant Analysis of the ions present in the four *Pericopsis*-species.

Figure 3: Kernel Discriminant Analysis of the ions present in *Pericopsis elata*, *Pericopsis* (other), *Dalbergia melanoxylon* and *Milicia excelsa*.

TABLES

Table 1: Total sample number per species group.

Group	Species	Samples
<i>Pericopsis elata</i>	<i>Pericopsis elata</i>	22
<i>Pericopsis</i> (others)	<i>Pericopsis angolensis</i>	21
	<i>Pericopsis laxiflora</i>	5
	<i>Pericopsis mooniana</i>	10
	<i>Dalbergia melanoxylon</i>	19
<i>Milicia excelsa</i>	<i>Milicia excelsa</i>	18
Sum		95

Table 2: Confusion matrix from the Random Forest of dataset 1 (all the species) based on the training set. Note that the classification accuracy is shown and not the classification error.

	<i>Dalbergia melanoxylon</i>	<i>Milicia excelsa</i>	<i>Pericopsis</i> (other)	<i>Pericopsis elata</i>	Classification accuracy
<i>Dalbergia melanoxylon</i>	16				100.00
<i>Milicia excelsa</i>		13	1		92.86
<i>Pericopsis</i> (other)			30		100.00
<i>Pericopsis elata</i>			2	14	87.50

Table 3: Classification of the prediction data set for dataset 1 (all the species).

	<i>Dalbergia melanoxylon</i>	<i>Milicia excelsa</i>	<i>Pericopsis</i> (other)	<i>Pericopsis elata</i>
<i>Dalbergia melanoxylon</i>	3			
<i>Milicia excelsa</i>		4		
<i>Pericopsis</i> (other)			6	
<i>Pericopsis elata</i>				6

Table 4: The LOOCV for the KDA based on manually chosen ions, based on the most important ions according to MDI and MDA and the OOB estimation of error rate from the Random Forest for dataset 1 (all the species).

%	MDI	MDA
LOOCV – 5	70.53	70.53
LOOCV – 10	76.84	76.84
LOOCV – 20	94.74	94.74
LOOCV – 30	92.63	94.74
LOOCV – 40	94.74	93.68
LOOCV – 50	95.79	92.63
LOOCV – 100	94.74	95.79
LOOCV – 200	95.79	95.79
LOOCV – 256	95.79	95.79
LOOCV – manual		95.79
OOB		3.95
1 – OOB		96.05

APPENDIX

Table A.1: Geographic provenance, country of origin, source and the number of specimens per species. The sources are: Cook Woods, Klamath Falls, OR, USA (CW); Eisenbrand Inc. Exotic Hardwoods, Torrance, CA, USA (EIEH); Forest Products Laboratory, Madison, WI, USA (FPL); Naturalis Biodiveristy Center, Leiden, Netherlands (NBC Naturalis) and the Royal Museum for Central Africa, Tervuren, Belgium (RMCA).

Species	CITES	Geographic provenance	Origin	Source	n
<i>Dalbergia melanoxylon</i> Guill. & Perr.	App II	Africa	Tanzania	EIEH	19
TOTAL					19
<i>Haplormosia monophylla</i> (Harms) Harms	/		DRC	RMCA	1
			Liberia	FPL	2
				RMCA	1
TOTAL					4
<i>Milicia excelsa</i> (Welw.) C.C.Berg	/	Africa	Angola	RMCA	1
			Burundi	RMCA	1
			Cameroon	RMCA	1
			DRC	RMCA	14
			Ghana	RMCA	1
TOTAL					18
<i>Pericopsis angolensis</i> (Baker) Meeuwen	/	Africa	Angola	FPL	4
				RMCA	3
			Congo	FPL	1
			DRC	FPL	2
				RMCA	2
			Mozambique	RMCA	1
			Tanzania	FPL	2
			Zambia	FPL	4
				RMCA	2
TOTAL					21
<i>Pericopsis elata</i> (Harms) Meeuwen	App II	Africa	DRC	CW	17
			Ghana	FPL	4
			Unknown	FPL	1
TOTAL					22
<i>Pericopsis laxiflora</i> (Baker) Meeuwen	/	Africa	Nigeria	RMCA	1
			Senegal	RMCA	2
			Sudan	RMCA	1
			Togo	FPL	1
TOTAL					5
<i>Pericopsis mooniana</i> Thwaites	/	Africa	Togo	FPL	1
			Zambia	FPL	2
		Asia	Sri Lanka	FPL	2
		Unknown		NBC Naturalis	5
TOTAL					10