Antimicrobial activity of an acetic and boric acid solution against 

Staphylococcus pseudintermedius

Gevoeligheid van Staphylococcus pseudintermedius voor de combinatie azijnzuur-boorzuur

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ABSTRACT

Incubation of 10^7 colony forming units/ml of Staphylococcus pseudintermedius in an undiluted, a 1:2 and a 1:4 diluted aqueous 2% acetic acid and 2% boric acid solution resulted in inactivation of the bacteria within 30, 60 and 120 minutes, respectively. This indicates that a combination of these acids might be useful for local treatment of S. pseudintermedius infections. Further clinical studies are necessary, however, to confirm these in vitro results.

SAMENVATTING

Respectievelijk 30, 60 en 120 minuten na het suspenderen van 5x10^7 kolonie vormende eenheden van Staphylococcus pseudintermedius isolaten in 5 ml van een onverdunde, 1/2 en 1/4 verdunde waterige oplossing van 2% azijnzuur en 2% boorzuur, werden geen levende bacteriën meer gevonden. Dit duidt erop dat een combinatie van deze zuren nuttig zou kunnen zijn voor de lokale behandeling van S. pseudintermedius infecties. Bijkomende klinische studies zijn evenwel noodzakelijk om de hier beschreven in vitro bevindingen te bevestigen.

INTRODUCTION

In dogs, coagulase positive staphylococci have been associated with several suppurative conditions, including pyoderma, otitis externa, endometritis, urinary tract infections, conjunctivitis and wound infections (Hermans et al., 2004; Quinn et al., 1999). Staphylococcus aureus has only occasionally been isolated from these lesions and in the literature it is commonly accepted that Staphylococcus intermedius is the main major pathogenic Staphylococcus species associated with dogs (Hermans et al., 2004). However, recent studies have shown that canine staphylococcal isolates previously identified as S. intermedius, in fact belong to the newly described species Staphylococcus pseudintermedius (Bannoehr et al., 2007; Devriese et al., 2006; Sasaki et al., 2007a).

Infections caused by S. pseudintermedius are often treated with antibiotics. However, acquired antimicrobial resistance is frequent in canine isolates (Donné et al., 2000). Although these isolates are usually susceptible to cephalosporins and the combination amoxicillin-clavulanic acid, methicillin resistant S. pseudintermedius strains showing acquired resistance to all beta-lactam antibiotics have been described (Bannoehr et al., 2007; Sasaki et al., 2007b).

As an alternative to antimicrobial therapy, acetic and boric acid have been suggested for local treatment of bacterial infections. Both acids have been shown to exert antibacterial effects on different bacterial species, including staphylococci (Houlsby et al., 1986; Russel and Diez-Gonzalez, 1998).

A 2% acetic acid and 2% boric acid aqueous solution is commercially available as an ear and skin cleaner for use in dogs. In the present study, the bactericidal effect of different dilutions of this product on canine S. pseudintermedius isolates was determined.

MATERIALS AND METHODS

Two isolates (81 and 336), phenotypically identified as S. pseudintermedius (Devriese et al., 2006), were used in this study. Isolate 81 was obtained in 2005 from the uterus of a dog with pyometra. It showed acquired resistance to beta-lactamase susceptible beta-lactam antibiotics, macrolides, lincosamides, tetracyclines and neomycin. Isolate 336 was obtained in 2006 from the umbilicus of a pup that died shortly after birth. Acquired antimicrobial resistance was not detected in this isolate. Both isolates were cultured overnight at 35°C on Columbia agar with 5% sheep blood (Oxoid, Basingstoke, Hampshire, UK) in a 5% CO2 atmosphere.

The bactericidal effect of a 2% acetic acid and 2% boric acid aqueous solution (Malacetic Otic, Dermapet, U.S.A.) against these isolates was determined. Approximately 5x10^7 colony forming units (cfu) of each S. pseudintermedius isolate were suspended in 5 ml undiluted, 1:2 and 1:4 diluted (in distilled water) Malacetic Otic. As a control, approximately 5x10^7 cfu of each S. pseudintermedius isolate were suspended in 5 ml distilled water. All suspensions were incubated at 30°C in a linear shaking bath (GLS400, Grant Instruments, Shepreth, UK). At
Table 1. Logarithmic mean and standard deviation of the number of colony forming units (log10 cfu) per ml for Staphylococcus pseudintermedius isolates 81 and 336 after incubation at 30°C in distilled water, and in an undiluted, a 1:2 and a 1:4 diluted aqueous 2% acetic acid and 2% boric acid solution (AA-BA sol.)

<table>
<thead>
<tr>
<th>Suspension</th>
<th>0</th>
<th>30</th>
<th>60</th>
<th>Incubation time (minutes)</th>
<th>120</th>
<th>180</th>
<th>240</th>
<th>300</th>
<th>360</th>
</tr>
</thead>
<tbody>
<tr>
<td>Isolate 81 - distilled water</td>
<td>6.74 ± 0.05</td>
<td>6.84 ± 0.04</td>
<td>6.87 ± 0.03</td>
<td>6.77 ± 0.17</td>
<td>6.79 ± 0.08</td>
<td>6.72 ± 0.08</td>
<td>6.82 ± 0.05</td>
<td>6.69 ± 0.17</td>
<td></td>
</tr>
<tr>
<td>Isolate 81 – 1:1 AA-BA sol.</td>
<td>6.00 ± 0.07</td>
<td>neg.*</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
</tr>
<tr>
<td>Isolate 81 – 1:2 AA-BA sol.</td>
<td>6.53 ± 0.08</td>
<td>2.54 ± 0.28</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
</tr>
<tr>
<td>Isolate 81 – 1:4 AA-BA sol.</td>
<td>6.68 ± 0.13</td>
<td>4.70 ± 0.13</td>
<td>2.66 ± 0.38</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
</tr>
<tr>
<td>Isolate 336 - distilled water</td>
<td>6.91 ± 0.07</td>
<td>6.89 ± 0.03</td>
<td>6.88 ± 0.03</td>
<td>6.82 ± 0.02</td>
<td>6.71 ± 0.17</td>
<td>6.73 ± 0.07</td>
<td>6.79 ± 0.07</td>
<td>6.81 ± 0.05</td>
<td></td>
</tr>
<tr>
<td>Isolate 336 – 1:1 AA-BA sol.</td>
<td>5.22 ± 0.00</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
</tr>
<tr>
<td>Isolate 336 – 1:2 AA-BA sol.</td>
<td>6.49 ± 0.09</td>
<td>2.91 ± 0.09</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
</tr>
<tr>
<td>Isolate 336 – 1:4 AA-BA sol.</td>
<td>6.67 ± 0.04</td>
<td>4.10 ± 0.01</td>
<td>2.80 ± 0.02</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
<td>neg.</td>
</tr>
</tbody>
</table>

*neg.: <1.22

RESULTS

The results are summarized in Table 1. No differences were observed between the two isolates tested. In the control samples without Malacetic Otic, the bacterial titer did not change throughout the experiment. In the undiluted Malacetic Otic solution, the bacteria were inactivated within 30 minutes. Incubation of S. pseudintermedius in a 1:2 and a 1:4 diluted Malacetic Otic solution resulted in inactivation of the bacteria within 60 and 120 minutes, respectively.

DISCUSSION

It can be concluded that, under the conditions tested here, an aqueous solution containing at least 0.5% acetic acid and 0.5% boric acid exerts a bactericidal effect against S. pseudintermedius. This indicates that a combination of these acids might be useful for local treatment of S. pseudintermedius infections.

In the present study, pure cultures of S. pseudintermedius were incubated in an aqueous environment. The in vivo situation is far more complex. Several factors, including the presence of organic material, mixed infections of S. pseudintermedius with other bacteria or fungi, and the presence of bacteria in biofilms (Hae sebrouck et al., 2007) may influence the effects of acetic and boric acid. Therefore, further clinical studies are necessary to confirm the usefulness of these acids in the field.

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REFERENCES


