

1 Short trains of intra-epidermal electrical stimulation to elicit reliable behavioral and  
2 electrophysiological responses to the selective activation of nociceptors in humans.

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**21 Abstract**

22 Currently, the study of nociception in humans relies mainly on thermal stimulation of heat-  
23 sensitive nociceptive afferents. To circumvent some limitations of thermal stimulation, it was  
24 proposed that intra-epidermal electrical stimulation (IES) could be used as an alternative  
25 method to activate nociceptors selectively. The selectivity of IES relies on the fact that it can  
26 generate a very focal electrical current and, thereby, activate nociceptive free nerve endings  
27 located in the epidermis without concomitantly activating non-nociceptive mechanoreceptors  
28 located more deeply in the dermis. However, an important limitation of IES is that it is  
29 selective for nociceptors only when very low current intensities are used. At these intensities,  
30 the stimulus generates a very weak percept, and the signal-to-noise ratio of the elicited evoked  
31 potentials (EPs) is very low. To circumvent this limitation, it was proposed that the strength of  
32 the nociceptive afferent volley could be increased through temporal summation, using short  
33 trains of repeated IES. Here, we characterized the intensity of perception and EPs elicited by  
34 trains of 2, 3 and 4 IES delivered using a 5-ms inter-stimulus interval. We found that both the  
35 intensity of perception and the magnitude of EPs significantly increased with the number of  
36 pulses. In contrast, the latency of the elicited EPs was not affected by the number of pulses,  
37 indicating that temporal summation did not affect the type of activated fibers and, therefore,  
38 that trains of IES can be used to increase the reliability of stimulus-evoked responses while  
39 still preserving its selectivity for nociceptors.

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## 44 **1. Introduction**

45 During the last decades, investigation of the neurophysiological mechanisms underlying  
46 nociceptive processing and pain perception has relied mainly on the thermal stimulation of  
47 cutaneous A $\delta$ - and C-fiber free nerve endings [17]. For example, thermal stimuli generated by  
48 laser stimulators have been used extensively because of their indisputable selectivity for heat-  
49 sensitive nociceptors [1]. In addition, due to their high power, lasers can generate very steep  
50 heating ramps, and thus elicit synchronous afferent discharges enabling the recording of time-  
51 locked responses such as event-related brain potentials (ERPs) or reaction times [2]. More  
52 recently, intra-epidermal electrical stimulation (IES) [10] and electrical stimulation using a  
53 small surface concentric electrode [11] have been proposed as alternative methods to activate  
54 nociceptors selectively and, thereby, explore nociception [10]. The rationale for these  
55 stimulation techniques relies on the fact that nociceptive free nerve endings are preferentially  
56 located in the epidermis, while non-nociceptive mechanoreceptors are mainly located more  
57 deeply in the dermis. Therefore, pulses of electric current spatially restricted to the epidermis  
58 could activate nociceptors selectively. These alternative methods could circumvent some  
59 limitations of laser stimulation, such as skin overheating and lesion due to stimulus repetition,  
60 and delay or relative desynchronization of the nociceptive afferent volley due to transduction  
61 of thermal energy into a neural impulse. However, these stimulation techniques suffer from  
62 their own limitations, in particular, the need to use low stimulation current intensities to  
63 guarantee its selectivity for nociceptors. Indeed, it has been shown that if IES is delivered  
64 using a strong intensity (e.g. an intensity corresponding to the pain threshold), the stimulus is  
65 not selective for nociceptors because it also activates more deeply located low-threshold  
66 mechanoreceptors [5,18]. In particular, it was shown that selective denervation of nociceptive  
67 free nerve endings by prolonged topical application of capsaicin abolishes the behavioral and  
68 electrophysiological responses to laser stimuli and IES delivered at low current intensities

69 (corresponding to twice the absolute detection threshold;  $0.18 \pm 0.25 \text{mA}$ ) but does not affect  
70 the responses to conventional transcutaneous electrical stimulation and IES delivered at a  
71 stronger intensity of current (2.5 mA) [15]. Thus, there is converging evidence that IES can  
72 activate nociceptors selectively, if and only if low intensities of current are used [14]. The  
73 important drawback is that at such low intensities, a single pulse of IES elicits a very weak  
74 sensation and the signal-to-noise ratio of the elicited ERPs is low, possibly because of the  
75 very small number of recruited afferents. This drawback has probably limited the use of this  
76 technique for pain research, and as a consequence, its availability. To circumvent the lack of  
77 spatial summation, some authors have proposed to deliver short trains of electrical pulses (e.g.  
78 three pulses delivered at a 5-ms inter-stimulus interval) [7,11,13,16,20,21], with the aim of  
79 increasing the strength of the nociceptive afferent volley through temporal summation.  
80 However, in these studies, the latency of the elicited ERPs was not systematically analyzed.  
81 As the latency of ERP components depends on the conduction velocity of the sensory fibers,  
82 and, therefore, on the type of fiber activated by the eliciting stimulus, it is important to ensure  
83 that temporal summation does not affect the type of fibers activated by IES. The aim of the  
84 present study was to compare the magnitude and latency of the perception and ERPs elicited  
85 by trains of 2, 3 or 4 pulses of IES delivered using a 5-ms inter-stimulus interval.

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## 87 **2. Methods**

88 Eleven volunteers took part in the study (4 women, aged from 21 to 45 years) with no prior  
89 history of neurological, psychiatric or chronic pain disorder. Written informed consent was  
90 obtained and all experimental procedures were approved by the local ethics committee and  
91 conformed to the latest revision of the Declaration of Helsinki.

92 IES was delivered to the right hand dorsum using a stainless steel concentric bipolar electrode  
93 developed by Inui et al. [10] (Nihon Kohden, Japan). The electrode consists of a needle  
94 cathode (length: 0.1 mm, Ø: 0.2 mm) surrounded by a cylindrical anode (Ø: 1.4 mm). By  
95 gently pressing the device against the skin, the needle electrode was inserted in the epidermis  
96 of the hand dorsum, within the sensory territory of the superficial radial nerve. In order to  
97 guarantee the selectivity of the nociceptive stimulation, the intensity of the stimulus was  
98 individually adjusted to twice the absolute detection threshold to a single 0.5 ms constant-  
99 current square-wave pulse (DS7 Stimulator, Digitimer Ltd., UK). The detection threshold was  
100 estimated using an adaptive algorithm [3]. After positioning the electrode, single-pulse stimuli  
101 were applied using a staircase procedure, with detection vs. non detection as criterion, by  
102 increasing or decreasing the intensity of the electrical current in steps of 0.01 mA. The  
103 procedure was interrupted after the occurrence of four staircase reversals. The staircase  
104 converged towards the intensity at which the probability of detecting the stimulus was 50%  
105 [3]. The intensity was then set to twice the detection threshold, defined as the average of the  
106 intensity delivered at the four staircase reversals, with an intensity of  $\leq 0.50$  mA as restrictive  
107 criterion [4,6]. If this criterion was not met, the electrode was displaced and the adaptive  
108 staircase procedure was restarted.

109 During a first session, stimuli were applied using a single pulse or a train of 2, 3 or 4 pulses  
110 separated by a 5-ms inter-pulse interval. The different types of stimuli were repeated 5 times  
111 in random order. After each stimulus, the participants were asked to rate the perceived  
112 intensity of the stimulus using a numerical rating scale (NRS) extending from 0 to 100 (0 =  
113 not perceived; 100 = maximum pain; 50 = limit between non-painful and painful domains of  
114 sensation).

115 During a second session, the electroencephalogram (EEG) was recorded using 19 Ag-AgCl  
116 electrodes placed on the scalp according to the International 10-20 system and referenced to

117 linked earlobes (A1-A2). Ocular movements and eye-blinks were recorded using two  
118 additional bipolar electrodes placed at the upper-left and lower-right sides of the left eye. The  
119 signals were amplified, digitized at a 167 Hz sampling rate (PL-EEG, Walter Graphtek,  
120 Germany). Stimuli were applied using a train of 2, 3 or 4 pulses separated by a 5-ms inter-  
121 pulse interval, delivered in a random order in three consecutive blocks of 30 trials each (one  
122 block = 10 trials x 3 stimulus types). Within a block, the inter-train interval varied randomly  
123 from 5 to 10 s (rectangular distribution). Each block was separated by a 2-5 min pause.  
124 Participants were asked to press a button held in the left hand as soon as they perceived the  
125 stimulus. The mean reaction time (RT) recorded relative to stimulus onset was used as a  
126 measure of response speed. RTs greater than 1000 ms were considered as undetected. We also  
127 examined the frequency distribution of RTs according to stimulus type. For this purpose, RTs  
128 were grouped in 100-ms bins extending from 0 to 1000 ms.

129 Offline analyses of the EEG data were carried out using Brain Vision Analyzer 1.05 (Brain  
130 Products GmbH, Germany) and Letswave 5 (Université catholique de Louvain, Belgium).  
131 The continuous EEG recordings were band-pass filtered (0.5-45 Hz) and segmented into 2000  
132 ms epochs extending from -500 to +1500 ms relative to stimulus onset. Artifacts produced by  
133 eye blinks and eye movements were corrected using an Independent Component Analysis [9].  
134 Signals were re-referenced according to a common average reference, and baseline-corrected  
135 from -500 to 0 ms. Epochs containing artefacts were identified by visual inspection and  
136 excluded from further analyses (rejected epochs constituted less than 15% of the total number  
137 of epochs). The epochs were then averaged according to the number of pulses (2, 3 or 4).  
138 Furthermore, an additional set of average waveforms were computed to test the effect of  
139 repetition. For each subject, the full set of epochs were split into four blocks according to trial  
140 order (blocks 1 to 4) and number of pulses (2, 3 or 4), yielding 6 average waveforms for each  
141 subject. Within each average waveform, the latency and amplitude of three distinct peaks

142 were measured as follows. First, a negative peak (N2) was identified as the most negative  
143 peak obtained at Cz within 200-300 ms after stimulus onset. Second, a positive peak (P2) was  
144 defined as the most positive peak obtained at Cz within 300-400 ms after stimulus onset. The  
145 peak-to-peak amplitude of the N2-P2 complex was obtained by subtracting the N2 peak  
146 amplitude from the P2 peak amplitude. Third, a negative peak (N1) was identified at the  
147 contralateral electrode T3 re-referenced to Fz, within 120-170 ms after stimulus onset.

148 The effect of the number of stimuli was assessed using an ANOVA for repeated measures  
149 (GraphPad 5, GraphPad Software Inc., CA) with *stimulus type* as within-subject factor with  
150 four levels (1 vs. 2 vs. 3 vs. 4 pulses) for the intensity of perception, and three levels (2 vs. 3  
151 vs. 4 pulses) for RTs and ERP amplitudes and latencies. For the N2-P2 difference, the  
152 ANOVA also included *time* as second within-subject factor (blocks 1 vs. 2 vs. 3 vs. 4), in  
153 order to assess the effect of stimulus repetition. Analyses included data from unperceived  
154 stimuli (NRS = 0 in the first session, trials with no button press in the second session), except  
155 for RTs. Post-hoc analyses using paired *t*-tests were performed when necessary. Significance  
156 level was set at  $p < 0.05$ .

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### 158 **3. Results**

159 The group-level mean  $\pm$ SD of the absolute detection threshold was  $0.09 \pm 0.07$  mA. With an  
160 intensity set at twice the detection threshold, the mean  $\pm$ SD intensity of perception (NRS) was  
161  $25 \pm 15$  for a single pulse,  $40 \pm 18$  for 2 pulses;  $51 \pm 14$  for 3 pulses and  $59 \pm 13$  for 4 pulses, as  
162 shown in Figure 1A. The ANOVA revealed a significant effect of *stimulus type* on the  
163 intensity of perception ( $p < 0.001$ ). All post-hoc *t*-tests were significant: 1 vs. 2 pulses, 2 vs. 3  
164 pulses, and 3 vs. 4 pulses (all  $p < 0.013$ ).

165 The group-level mean  $\pm$ SD of RTs was  $397 \pm 47$ ms,  $341 \pm 49$ ms and  $330 \pm 51$ ms for trains of 2,  
166 3 and 4 pulses, respectively. The ANOVA showed that RTs were significantly different  
167 according to stimulus type ( $p < 0.001$ ). Post-hoc comparisons showed that RTs to 2 pulses  
168 were significantly greater than RTs to 3 pulses ( $p < 0.001$ ), and that RTs to 3 pulses were  
169 significantly greater than RTs to 4 pulses ( $p < 0.010$ ). As shown in Figure 1B, most stimuli  
170 were detected with RTs between 300 and 500 ms, independently of stimulus type. Such RTs  
171 are compatible with the conduction velocity of myelinated A-fibers [3].

172 Clear ERPs were identified in 9 of the 11 participants. Figure 2 illustrates the group-level  
173 average ERP waveforms elicited by 2, 3 and 4 pulses as well as the group-level scalp  
174 topographies of the N1, N2 and P2 waves. The mean peak latencies and amplitudes of these  
175 components are detailed in Table 1. The peak-to-peak amplitude of the N2-P2 complex was  
176 significantly affected by *stimulus type* ( $p = 0.021$ ) and *time* ( $p = 0.015$ ). The interaction  
177 between the two factors was not significant ( $p = 0.348$ ). This indicates a significant effect of  
178 the numbers of IES pulses and a significant effect of stimulus repetition on the magnitude of  
179 the elicited ERPs. In addition, it suggests that the effect of stimulus repetition was not  
180 different for the different types of stimuli. Separate analyses for N1, N2 and P2 amplitudes  
181 showed a significant effect of *stimulus type* for N1 ( $p = 0.030$ ) and P2 ( $p = 0.039$ ), but not for  
182 N2 ( $p = 0.302$ ). Post-hoc comparisons revealed a significant difference between 2 and 4  
183 pulses for both N1 ( $p = 0.028$ ) and P2 ( $p = 0.044$ ). There was no significant difference  
184 between 2 and 3 pulses (N1:  $p = 0.203$ ; P2:  $p = 0.202$ ), as well as between 3 and 4 pulses (N1:  
185  $p = 0.285$ ; P2:  $p = 0.519$ ). Comparison of the N1, N2 and P2 latencies did not reveal any  
186 significant effect of *stimulus type* (N1:  $p = 0.257$ ; N2:  $p = 0.641$ ; P2:  $p = 0.816$ ).

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#### 188 4. Discussion



189 The present study confirms that when care is taken to position the electrode, the absolute  
190 detection threshold of IES lies at very low intensities. Notably, these absolute detection  
191 threshold values are far below the values at which a single pulse of IES elicits a percept  
192 qualified as painful (e.g.,  $1.6 \pm 0.5$  mA; [5]). Importantly, the temporal summation resulting  
193 from increasing the number of IES pulses significantly increased the intensity of the elicited  
194 percept. Therefore, manipulation of the numbers of pulses represents a viable approach to  
195 increase the strength of the nociceptive stimulus without changing the intensity of the  
196 electrical current and, thus, ensuring that the stimulus remains selective for nociceptors.  
197 Similarly, the amplitude of the ERPs elicited by IES was also increased by the numbers of  
198 pulses. However, the relationship between number of pulses and ERP magnitude was not  
199 exactly similar to the relationship between number of pulses and intensity of perception.  
200 Indeed, whereas increasing the number of pulses appeared to be related to a gradual increase  
201 of perceived intensity, the increase of ERP magnitude with the number of pulses appeared to  
202 reach a plateau between 3 and 4 pulses.

203 There was a significant effect of the number of pulses on reaction time latencies. This could  
204 be explained by either a difference in peripheral conduction time or a difference in the central  
205 processing time required to detect and respond to the incoming sensory input [19]. Increasing  
206 the number of pulses could have led to the activation of additional fibers brought to a  
207 subthreshold potential by the preceding pulses and, hence, could have led to the activation of  
208 faster, non-nociceptive A-fibers. However, this interpretation seems unlikely. Indeed, there  
209 was no effect of the number of pulses on the latency of the elicited ERP components,  
210 suggesting that the ERPs elicited by the different numbers of pulses were related to the  
211 activation of the same type of afferents, i.e. predominantly A $\delta$ -fiber afferents [15]. The effect  
212 of the number of pulses on reaction time latencies was thus probably related to the fact that

213 increasing the number of pulses increases the strength of the nociceptive input and, hence,  
214 facilitates and speeds its detection (e.g. [8]).

215 Taken together, our results indicate that increasing the number of pulses constitutes an  
216 appropriate procedure to increase the signal-to-noise ratio of the brain responses elicited by  
217 IES without jeopardizing its selectivity for nociceptive afferents. In contrast, previous studies  
218 showed that increasing the intensity of the electrical current decreases significantly the  
219 latencies of the elicited ERPs, suggesting that in this case, the elicited ERPs are no longer  
220 related to the activation of A $\delta$ -fibers but to the concomitant activation of non-nociceptive A $\beta$ -  
221 fibers [5,18]. As compared to laser stimulation, IES could be particularly useful in  
222 experimental contexts requiring short inter-stimulus intervals, stimulation at various locations  
223 or combination with other types of stimuli. Of course, the fact that the selectivity of IES  
224 depends crucially on current intensity constitutes an important limitation of the method,  
225 especially to explore nociceptive processing in patients with sensory disorders. However, this  
226 limitation could be partially circumvented by the use of a very strict procedure to define  
227 current intensity, as well as the use of short trains to increase the strength of the nociceptive  
228 afferent volley through temporal summation.

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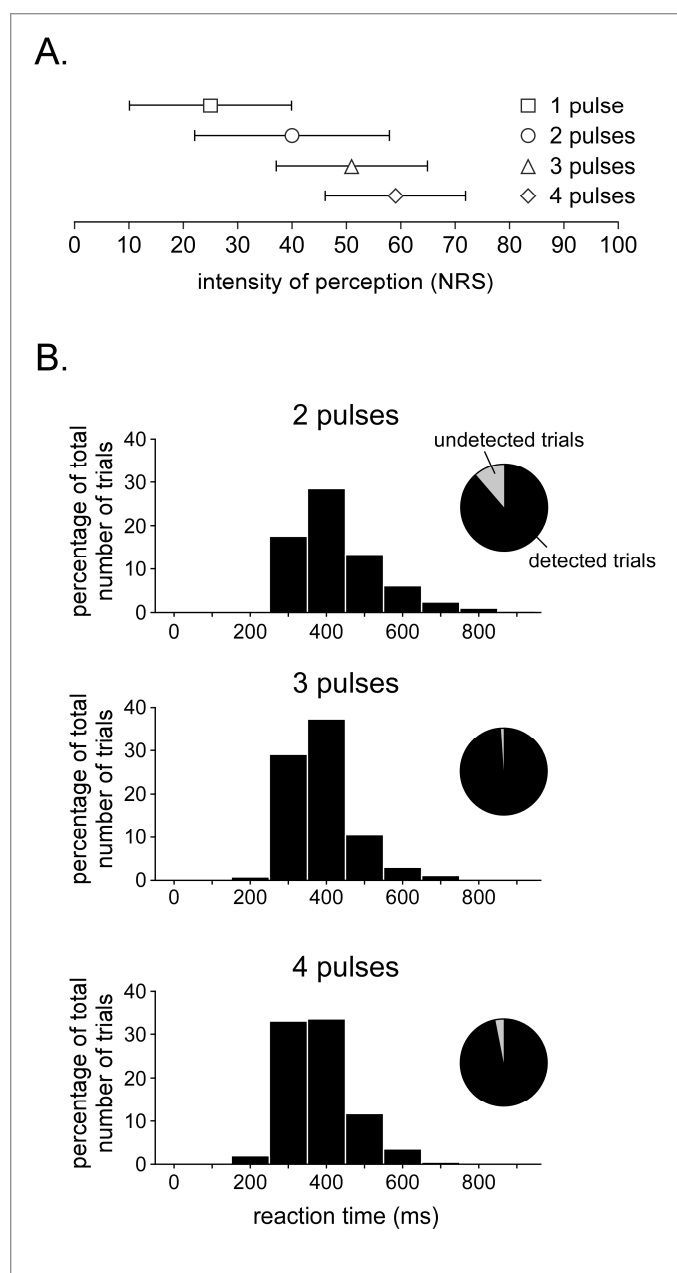
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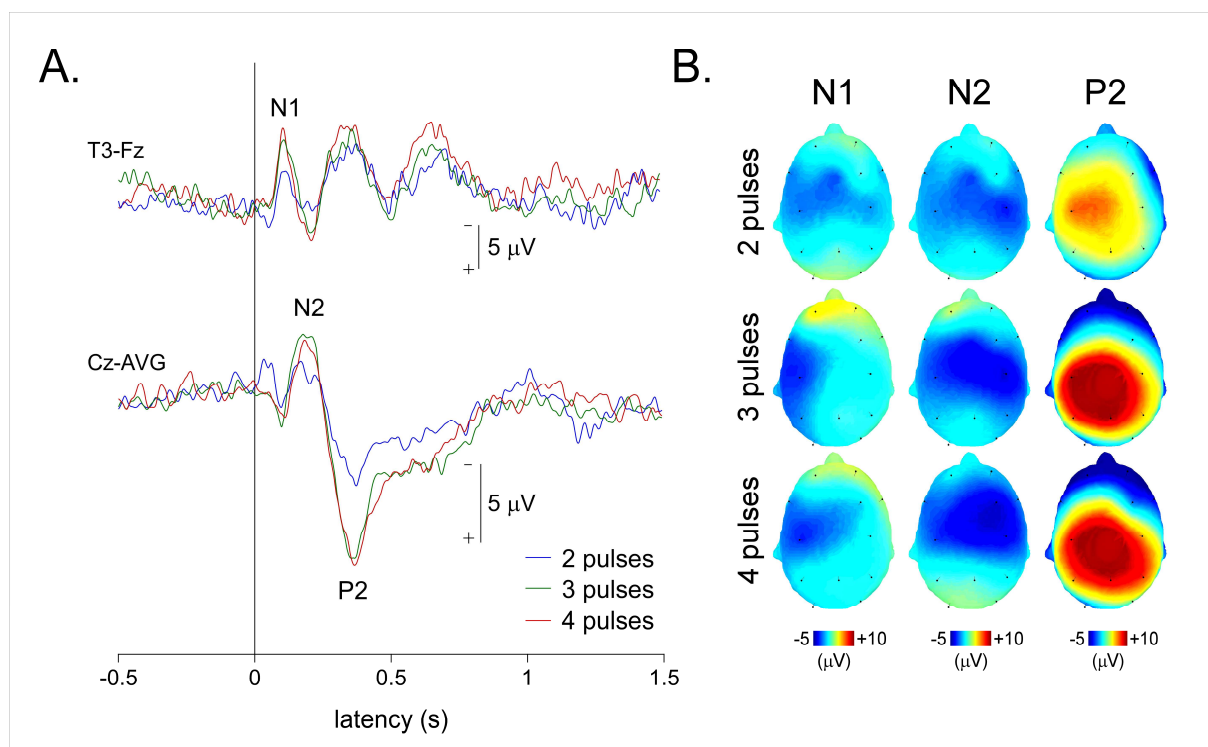
## 292 Figure legends



293

294 Figure 1. A. Group-level average intensity of the perception elicited by 1, 2, 3 and 4 pulses of  
 295 IES delivered to the right hand dorsum (the whisker plots represent the standard deviation  
 296 across participants). B. The histograms represent the frequency distribution of the reaction-  
 297 times to IES using 2, 3 and 4 pulses (bins of 100 ms). The pie charts represent the proportion  
 298 of detected (black) and undetected (grey) trials.

299



300

301 Figure 2. A. Group-level average waveforms of the ERPs elicited by 2, 3 and 4 pulses of IES  
302 delivered to the right hand dorsum obtained at electrode Cz (average reference) and T3 (Fz  
303 reference). B. Group-level average scalp topographies of the N1, N2 and P2 waves elicited by  
304 2, 3 and 4 pulses of IES.

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313 Table 1

|                 |                       | Number of pulses of IES |             |             |
|-----------------|-----------------------|-------------------------|-------------|-------------|
|                 |                       | 2                       | 3           | 4           |
| <hr/>           |                       |                         |             |             |
| Latencies (ms)  |                       |                         |             |             |
|                 | N1                    | 149 ±102                | 151 ±109    | 144 ±102    |
|                 | N2                    | 228 ±111                | 220 ±106    | 223 ±101    |
|                 | P2                    | 369 ±113                | 363 ±118    | 367 ±105    |
| Amplitudes (µV) |                       |                         |             |             |
|                 | N1                    | -6.7 ±3.3               | - 11.6 ±7.3 | - 12.0 ±5.8 |
|                 | N2                    | -5.4 ±2.3               | - 6.5 ±3.7  | - 7.3 ±3.4  |
|                 | P2                    | 9.1 ±5.7                | 13.4 ±8.5   | 12.8 ±7.9   |
|                 | N2-P2 difference (µV) | 14.5 ±5.6               | 19.9 ±9.9   | 20.1 ±9.7   |
| <hr/>           |                       |                         |             |             |

314

315 Table 1. Latencies and amplitudes (mean ± standard deviation) of the ERP components N1, N2  
316 and P2, and the N2-P2 amplitude difference, according to the number of pulses of the intra-  
317 epidermal stimulation.