**Thermic dehorning and ear tagging as atypical portals of entry of Clostridium tetani in ruminants**

*Thermische onthoornen en oormerken als atypische intredeplaatsen van Clostridium tetani bij herkauwers*

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**ABSTRACT**

This paper describes two infections with *Clostridium tetani* (*C. tetani*). One outbreak occurred after dehorning of calves, the second infection happened after ear tagging of a goat. In the first case 3 young Holstein Friesian calves showed generalized stiffness, severe lock-jaw and bloat two weeks after dehorning. The thermal dehorning wounds were identified as the infection sites of *C. tetani* by bacterial culture and PCR. The second case was a three-year old male castrated goat, with generalized stiffness. The animal had been ear tagged one week prior to the onset of the symptoms. *C. tetani* could be cultured from pus on the ear tag. Treatment was attempted in two calves and the goat. Wounds were debrided and disinfected, penicillin and anti-tetanus serum were administered and polyionic perfusions provided. In addition, the goat was vaccinated against tetanus. The goat and one calf fully recovered after 36 and 8 days respectively. To the authors’ knowledge a tetanus outbreak in association with thermal dehorning has not been described previously. Also ear tagging as a possible cause for *C. tetani* infection has not been described in goats.

**SAMENVATTING**


Voor zover bekend werd er nog nooit een tetanusuitbraak geassocieerd met thermisch onthoornen, beschreven. Ook het oormerken werd nog niet beschreven als een mogelijke oorzaak van een *C. Tetani*-infectie bij geiten.

**INTRODUCTION**

Tetanus is a life-threatening toxico-infection caused by the neurotoxin of *C. tetani*. Typical symptoms are increased general muscular rigidity evolving to a complete tetanic state, frequently with a fatal outcome (Linnenbrink et al., 2006). Although tetanus is described in all domestic animals, small ruminants and horses are known to be the most susceptible species (Aslani et al., 1998; Driemeier et al., 2006; Wernery et al., 2004). Tetanus usually affects a single animal, but outbreaks have been sporadically described in farm animals, especially in association with serial injections (Barbosa et al., 2009; Driemeier et al., 2007). Infection occurs as a result of contamination of wounds with spores of *C. tetani*, which desporulate to the vegetative form and produce tetanus toxin when anaerobic conditions are present. The most frequently described infection sites in farm animals are castration, shearing and injection wounds (Aslani et al., 1998; Linnenbrink et al., 2006; Poudel et al., 2009). This case report describes two tetanus cases with an atypical portal of entry, the first being an outbreak of tetanus in dairy calves, shortly after thermal dehorning and the second in a three-year old goat, after ear tagging.
CASE 1

Case history

In May 2011 three Holstein Friesian calves, between 3 and 5 months of age, were referred to the Veterinary Clinic in Merelbeke with complaints of stiffness, recumbency, ruminal bloat and lock-jaw. The symptoms had started 2 to 7 days before presentation. At the farm, all animals were housed in the same pen together with thirteen other calves. Fourteen days before admission, eight of these calves were thermally dehorned. Seven days before admission, one of the recently dehorned calves was found dead. No previous symptoms were noted by the owner, and no necropsy was performed. The animals were vaccinated against IBR, but not against clostridial diseases.

Clinical examination and ancillary diagnostics

All calves showed stiffness in all four limbs, markedly increased muscle tone, pricked ears, lifted tail and bloat. One calf was still able to lie down and get up without assistance and to ingest small amounts of feed and water, whereas the other two calves could not bend their legs, had severe lock-jaw and were unable to eat or drink. The dehorning wounds had a thick crust with pus underneath. No other wounds were detected. Temperature, respiration and heart rate were normal in all calves.

Standard hematology, biochemistry and blood gas analysis only showed a mild to severe dehydration in all calves. The crust was removed from the dehorning wounds and cultured for Clostridium tetani. The crust was suspended in sterile phosphate buffer solution (PBS) and heated at 80°C for 20 minutes. Of this suspension, 100µl was cultivated at 37°C for 24 hours on a blood plate in an anaerobic chamber (Ruskinn Technology) with 84% N₂, 8% CO₂ and 8% H₂. A pure culture of a hemolytic gram-positive rod with terminal spores was obtained (Figure 1a). The final identification of C. tetani was done by PCR (Akbulut et al., 2005).

Treatment and further evolution

One calf was euthanized immediately at arrival due to the graveness of the symptoms and the bad prognosis. The dehorning wounds of the remaining calves were thoroughly cleaned with H₂O₂. The calves received IV 10 ml of anti-tetanus serum® (Intervet), daily injections with procaine benzylpenicillin (10 mg/kg, IM, Duphapen, Pfizer A.H.) and thiamine (10mg/kg, IV, tid) for 6 days and polyionic perfusion with 10% glucose. A second calf was euthanized 2 days after arrival in the clinic due to worsening of the symptoms. In the third calf, which showed the best clinical condition at arrival, the symptoms gradually improved and the patient could leave the clinic 7 days after presentation with minimal signs of stiffness.

CASE 2

Case history

In June 2011, a castrated male goat of 3 years of age was presented at the clinic with symptoms of apathy, generalized stiffness, dysphagia and reduced ruminal sounds. At home, the veterinarian suspected indigestion or ruminal acidosis and treated the animal with butylscopolamine (Buscopan®, Boehringer Ingelheim BV), flunixine meglumine (Bedozane®, Eurovet), a bicarbonate perfusion and supplementation with vitamins and minerals. The animal was kept in a hobby setting on pasture together with 5 apparently healthy other goats. Although all animals were officially registered since birth, their ear tags had only been placed a week before presentation at the clinic. The animal had been vaccinated once against C. tetani before castration at 6 months of age, but without continuation of the vaccination program.

![Figure 1a](image1a.png)

Figure 1a. Spores and bacteria of Clostridium tetani with a typical drum-stick shape isolated from the crust of the dehorning wounds in case 1 (gram-staining-1000x).

![Figure 1b](image1b.png)

Figure 1b. Spores and bacteria of Clostridium tetani with a typical drum-stick shape isolated from the ear tags in case 2 (Malachite Green – 1000x).
Clinical examination and ancillary diagnostics

Upon admission, the goat showed obvious stiffness of the limbs and was alert. Rectal temperature, pulse and respiration rates were normal. Elevated abdominal tension, pricked ears, elevated tail and subtle ruminal bloat were present. The uptake of food and water was impossible due to severe lock-jaw. The animal could still defecate and urinate in a normal way.

The goat was slightly dehydrated. No other deviations were found on standard blood examination. The ruminal fluid had a normal macroscopic aspect and a pH of 7.3 (normal values 6-7). Multiple living protozoa were detected on microscopic examination (10x magnification). Standard parasitological examination resulted in a strongylo count of 8100 eggs per gram faeces and an Eimeria spp. count of 450 oöcysts per gram faeces. Of the detected oöcysts roughly 10% belonged to the virulent species Eimeria ninakohlyakimovae. The ear tags were removed after permission of the official authority responsible for food animal identification. From pus sticking to the ear tags, an anaerobic culture was performed as described in case 1 and Clostridium tetani could be cultured. Further identification was done as described in case 1. A smear of the purified colonies was stained with Malachite Green and safranin and the typical drum-stick shaped rods were visible, as demonstrated in Figure 1b.

Treatment and further evolution

The crusts were removed and the wounds were disinfected with H2O2. The goat received 4 ml of anti-tetanus serum® (Intervet), and daily injections with procaine benzylpenicillin (10 mg/kg, IM, Duphapen, Pfizer A.H.) and thiamine (10 mg/kg, IV, tid) for 6 days and a polyionic perfusion with 10% glucose. Additional treatment and further evolution

Tetanus is a fatal toxico-infection, caused by toxins of Clostridium tetani (Poudel et al., 2009). This bacterium is able to produce two important toxins, namely tetanoylsgin and tetanospasmin. The first has the ability to lyse cell membranes, causing tissue damage and stimulating the development of an anaerobic environment. The latter is responsible for the neurological symptoms by inhibiting the release of inhibitory neurotransmitters, causing generalized muscle spasm and alterations of autonomic control. Because the toxin is slowly transported retrograde from the peripheral nerves to the central nervous system, incubation periods of 3 to 18 days are described (Driemeier et al., 2006; Linnenbrink et al., 2006). C. tetani produces very resistant spores that can survive boiling temperatures for several minutes (Dixit et al., 2005; Linnenbrink et al., 2006). In order to desporulate to a vegetative form, the spores need an environment with low oxygen tension, typically generated under the crust of penetrating wounds (Linnenbrink et al., 2006). Although the advanced symptoms might resemble those seen in traumatic reticuloperitonitis, white muscle disease, grass tetany, menigitis, cerebrocortical necrosis, lead intoxication, strychnine-intoxication and to a lesser extent laminitis, poly-arthritis, esophageal obstruction and other causes of ruminal tympany (Stöber, 2002). However, in the case of tetanus the symptoms quickly evolve to the pathognomonic tonic-clonic cramps of the antigravitational muscles, protrusion of the third eye lid, stiff ears and a raised tail base. Although tetanus typically is an individual problem, outbreaks have been described in several animal species and humans, due to very different entry routes. Most famous are human outbreaks in intravenous drug users due to the use of contaminated needles (Sun K., 1994; Gormley et al., 2004). Also in ruminants multiple outbreaks have been described following injections with contaminated needles, for example after injection with anthelmintics in sheep and cattle or after rabies vaccination in buffalo’s (Driemeier et al., 2006; Barbosa et al., 2009). Although goats and sheep are more sensitive than cattle, (Aslani et al., 1998) outbreaks have more frequently been described in young cattle, possibly due to the more frequent application of mass injections in calves (Driemeier et al., 2006).

The present case report describes an outbreak of tetanus in three animals of the same herd, following thermal dehorning. Since the ban on chemical dehorning in 2008, thermal dehorning is the standard dehorning method for calves of 4 to 8 weeks of age (Nantier, 2009). The dehorning iron is heated to 700 degrees Celsius and is then placed upon the horn stumps, burning the epithelium and surrounding tissue and resulting in permanent removal of the horn stumps. After 1 to 2 days a thick impermeable crust is formed which encloses the dehorning wounds (Nantier, 2009). The temperature reached on the iron is kept for several minutes and is probably lethal to spores. Therefore, it is unlikely that the iron itself serves as a vector for C.
tetani (Dixit et al., 2005). Environmental contamination of the dehorning wound with faecal material and dirt from the stable environment seems a more likely infection route (Driemeier et al., 2006; Linnenbrink et al., 2006). Additionally, the thermal treatment also causes the perfect circumstances for the creation of an anaerobic environment by destroying the blood vessels and creating a thick impermeable crust. These factors predispose to the growth and toxin production of C. tetani (Nitzscheke et al., 2008). Burning wounds are a well known risk for tetanus in humans whereas no reports in animals were found (Cassel et al., 2002). This is the first time thermal dehorning is described as a cause of a tetanus infection. As this method is nowadays the standard dehorning procedure in Belgium and since most calves are not vaccinated against tetanus at older age and no cases of tetanus have been reported. Therefore, the occurrence of tetanus associated with ear tagging in this adult goat is likely an unfortunate incident.

Although the insertion of ear tags is routinely done in young ruminants, including goats, only one outbreak of tetanus in lambs following ear tag insertion at 7 to 8 days of age has been described (Aslani et al., 1998). It is not clear whether the older age of the animal was a risk factor for the occurrence of tetanus. However, permethrin-impregnated ear tags are relatively frequently administered in cattle and small ruminants at older age and no cases of tetanus have been reported. A solid vaccination scheme for tetanus is the most effective preventive measure, both for adults as through maternal immunity in newborns (Gall et al., 2011). Vaccination is advisable in highly susceptible species such as small ruminants and in animals with high economical or emotional value. Double primo-vaccination at minimally 8 and 12 weeks of age followed by yearly boosters provide a lasting protection later in life. Performing dehorning and ear tagging as clean as possible and local administration of antimicrobial sprays might decrease the risk. Inspection of the wounds a few days later and disinfection when necessary are advisable. The preventive application of antitetanus serum might be an option when ear tagging or dehorning valuable unvaccinated animals.

These cases illustrate the importance of a continued alertness when executing routine invasive interventions as ear tagging and dehorning. Although small ruminants are more sensitive than cattle and although extra attention should be given to tetanus prophylaxis when working with goats or sheep, tetanus may also occur in young cattle.

REFERENCES


